

## THEMED ISSUE: GPCR

### REVIEW

# Prostanoid receptor antagonists: development strategies and therapeutic applications

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Identification of the primary products of cyclo-oxygenase (COX)/prostaglandin synthase(s), which occurred between 1958 and 1976, was followed by a classification system for prostanoid receptors (DP, EP<sub>1</sub>, EP<sub>2</sub> . . .) based mainly on the pharmacological actions of natural and synthetic agonists and a few antagonists. The design of potent selective antagonists was rapid for certain prostanoid receptors (EP<sub>1</sub>, TP), slow for others (FP, IP) and has yet to be achieved in certain cases (EP<sub>2</sub>). While some antagonists are structurally related to the natural agonist, most recent compounds are 'non-prostanoid' (often acyl-sulphonamides) and have emerged from high-throughput screening of compound libraries, made possible by the development of (functional) assays involving single recombinant prostanoid receptors. Selective antagonists have been crucial to defining the roles of PGD<sub>2</sub> (acting on DP<sub>1</sub> and DP<sub>2</sub> receptors) and PGE<sub>2</sub> (on EP<sub>1</sub> and EP<sub>4</sub> receptors) in various inflammatory conditions; there are clear opportunities for therapeutic intervention. The vast endeavour on TP (thromboxane) antagonists is considered in relation to their limited pharmaceutical success in the cardiovascular area. Correspondingly, the clinical utility of IP (prostacyclin) antagonists is assessed in relation to the cloud hanging over the long-term safety of selective COX-2 inhibitors. Aspirin apart, COX inhibitors broadly suppress all prostanoid pathways, while high selectivity has been a major goal in receptor antagonist development; more targeted therapy may require an intermediate position with defined antagonist selectivity profiles. This review is intended to provide overviews of each antagonist class (including prostamide antagonists), covering major development strategies and current and potential clinical usage.

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**Abbreviations:** Ang II, angiotensin II; BMP, bone morphogenetic protein; BP, blood pressure; ClogP, predicted *n*-octanol/water partition coefficient; COX, cyclo-oxygenase; CPC, chronic persistent cough; PG, prostaglandin; EP<sup>+/+</sup>/EP<sup>-/-</sup>, system involving wild-type/homozygous gene-deleted EP receptor; FA, free acid; NSAID, non-steroidal anti-inflammatory agent; PAF, platelet-activating factor; PGIS, prostacyclin synthase; PLC, phospholipase C; PRP, platelet-rich plasma; rc, recombinant; SAR, structure–activity relationships; TM, transmembrane; TXA<sub>2</sub>, thromboxane A<sub>2</sub>; TXS, thromboxane synthase

### Introduction

The pharmacological classification for prostanoid receptors was developed on the basis that each receptor preferentially recognizes one of the major natural prostaglandins (PGs). Thus, PGD<sub>2</sub> preferentially activates the DP receptor, PGE<sub>2</sub> pref-

erentially activates the EP receptor, with the same applying to PGF<sub>2α</sub>/FP receptor and PGI<sub>2</sub> (prostacyclin)/IP receptor (Coleman *et al.*, 1994b). In the case of the TP receptor, both thromboxane A<sub>2</sub> (TXA<sub>2</sub>) and its precursor PGH<sub>2</sub> are potent agonists. There are two distinct subtypes of DP receptor, DP<sub>1</sub> and DP<sub>2</sub>; the latter has also been called CRTh2 (chemoattractant receptor-homologous molecule expressed on T helper 2 cells). Of the four EP receptor subtypes, EP<sub>1</sub> and EP<sub>3</sub> generally elicit excitatory actions, while EP<sub>2</sub> and EP<sub>4</sub> elicit inhibitory actions on cell function. All prostanoid receptors belong to the G protein-coupled receptor superfamily of cell-surface

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receptors. Each has seven transmembrane (TM)-spanning segments and may couple to one or more signal-transduction processes. In addition, mRNA splicing variants have been discovered for DP<sub>1</sub>, EP<sub>1</sub>, EP<sub>3</sub>, EP<sub>4</sub>, FP and TP receptors (see Pierce and Regan, 1998).

The repertoire of bioactive oxygenated lipids and signal transduction mechanisms is expanded by homodimerization and heterodimerization between different prostanoid receptors. For example, an isoprostane binding site is created by IP/TP $\alpha$  heterodimerization (Wilson *et al.*, 2004). Moreover, the prostamide (prostaglandin-ethanolamide) recognition site appears to result from heterodimerization of wild type and alternatively spliced FP receptor variants (Liang *et al.*, 2008). Prostanoid receptors may also complex with non-prostanoid receptors, for example, EP<sub>1</sub> receptor with  $\beta_2$ -adrenoceptor, but discussion of these interactions is outside the scope of this review. The receptor/second messenger nomenclature used in the review conforms to this journal's Guide to Receptors and Channels (Alexander *et al.*, 2008).

### Assay systems and antagonist development

The initial pharmacological differentiation of prostanoid receptors relied heavily on isolated tissue studies, with ileum, trachea and vas deferens of the guinea pig being especially important (Jones *et al.*, 1982; Coleman *et al.*, 1984; 1987; 1994a; Dong *et al.*, 1986; see Chen *et al.*, 2001 for experimen-

tal details). Isolated tissue preparations are still used today as they often reflect phenomena that occur in the integrated, living mammal. In addition, they still provide evidence for new receptor entities, as shown by the critical role of the cat iris preparation in the elucidation of prostamide pharmacology (Matias *et al.*, 2004; Woodward *et al.*, 2007; 2008). Studies on isolated and cultured cells have also been important in building the pharmacological classification of prostanoid receptors (Coleman *et al.*, 1984; Eglen and Whiting, 1988; Woodward *et al.*, 1995a,b).

Although of low throughput, isolated tissue studies have been critical to the discovery of prostanoid antagonists, particularly for DP<sub>1</sub>, EP<sub>1</sub> and TP receptors. However, it is remarkable that antagonists for other receptors (EP<sub>2</sub>, EP<sub>3</sub>, FP and IP) have been slow to emerge following discovery of the natural ligand/receptor; several reasons may account for this situation. Partial agonism is found in closely related analogues of the natural ligands for DP<sub>1</sub> and TP receptors (Table 1), thereby providing inroads to pure antagonists. However, partial agonism may not always be readily recognized. For example, taprostene, an early analogue of PGE<sub>2</sub> (Müller *et al.*, 1983), was only shown to be an IP partial agonist in 2004 (Chan and Jones, 2004). Furthermore, partial agonism may not always translate into pure antagonism. Thus, some of the many 'non-prostanoid prostacyclin mimetics' synthesized (see later) show IP partial agonism (Merritt *et al.*, 1991a; Jones *et al.*, 1997; Seiler *et al.*, 1997; Kam *et al.*, 2001), but IP antagonists have apparently not emerged from this grouping. Moreover, a

**Table 1** Prostanoid receptor agonists relevant to defining antagonist profiles

Prostanoid receptor	Full agonist		Partial agonist
	High selectivity	Moderate selectivity	
DP <sub>1</sub>	BW-245C		BW-192C86 <sup>a</sup>
DP <sub>2</sub>	15(R) PGD <sub>2</sub> , 15(R)-15-methyl PGD <sub>2</sub> 13,14-dihydro-15-oxo PGD <sub>2</sub>		
EP <sub>1</sub>	ONO-DI-004	17-Phenyl PGE <sub>2</sub>	Iloprost <sup>b,c</sup>
EP <sub>2</sub>	ONO-AE1-259, CAY-10399 <sup>d</sup>	Butaprost-FA, CP-533536 <sup>e</sup> 19(R)-hydroxy PGE <sub>2</sub> <sup>f</sup>	
EP <sub>3</sub>	ONO-AE-248, SC-46275	Sulprostone, MB-28767	ONO-AP-324 <sup>g</sup>
EP <sub>4</sub>	ONO-AE1-329, tetrazolo PGE <sub>1</sub> <sup>h</sup>	(PGE <sub>2</sub> ) <sup>i</sup>	
FP	Fluprostenol,latanoprost-FA	Cloprostenol	AL-8810 <sup>j,k,l</sup>
IP	Cicaprost	AFP-07, iloprost	Octimibate <sup>m</sup> , taprostene <sup>n</sup>
TP	STA <sub>2</sub> , U-46619		CTA <sub>2</sub> <sup>o</sup> , PTA <sub>2</sub> <sup>o,p</sup> , U-44069 <sup>p</sup>

Information on the non-referenced agonists may be obtained from Jones (2004) and this journal's Guide to Receptors and Channels edited by Alexander *et al.* (2008). In older publications, fluprostenol = ICI-81008, cloprostenol = ICI-80996, cicaprost = ZK-96480 and iloprost = ZK-36374. FA, free acid.

<sup>a</sup>Series of bicyclic-hydantoin prostanoids (Leff and Giles, 1992).

<sup>b</sup>Dong *et al.* (1986).

<sup>c</sup>Boie *et al.* (1997).

<sup>d</sup>Tani *et al.* (2002).

<sup>e</sup>Paralkar *et al.* (2003).

<sup>f</sup>Woodward *et al.* (1993a).

<sup>g</sup>Jones *et al.* (1998).

<sup>h</sup>Analogue 19a in Billot *et al.* (2003).

<sup>i</sup>Utility on high-sensitivity EP<sub>4</sub> systems.

<sup>j</sup>Griffin *et al.* (1999).

<sup>k</sup>Woodward *et al.* (2007).

<sup>l</sup>Sharif *et al.* (2008).

<sup>m</sup>Merritt *et al.* (1991a,b).

<sup>n</sup>Chan and Jones (2004).

<sup>o</sup>Armstrong *et al.* (1985).

<sup>p</sup>Jones *et al.* (1982).

large range of compounds block TP receptors (see later); this expansiveness may simply not apply to other prostanoid receptors. Finally, there has not been widespread commercial drive to develop EP<sub>2</sub> antagonists, for example, owing to a perceived lack of therapeutic utility (see later).

Dramatic progress in antagonist development was seen following cloning of the various prostanoid receptors in the early 1990s: DP<sub>1</sub> (Boie *et al.*, 1995), DP<sub>2</sub> (Hirai *et al.*, 2001), EP<sub>1</sub> (Funk *et al.*, 1993a), EP<sub>2</sub> (Regan *et al.*, 1994), EP<sub>3</sub> (Yang *et al.*, 1994), EP<sub>4</sub> (Bastien *et al.*, 1994), FP (Abramovitz *et al.*, 1994), IP (Namba *et al.*, 1994), TP (Hirata *et al.*, 1991). Stable over-expression of each prostanoid receptor in carrier cell lines allowed high-throughput radioligand binding and functional studies using 96- or even 384-well plate format. Thus, chemical library screening resulted in the discovery of new non-prostanoid scaffolds as leads, from which potent and selective agonists and antagonists were designed. Inspection of Figures 1–8 will reveal the prevalence of aryl-sulphonamido linkages in the various antagonist classes. Non-prostanoid structures provide a practical approach to obtaining new prostanoid-based therapies, as the issues of bioavailability, metabolic stability and cost of synthesis that surround PG analogues are avoided.

### Selective agonists for prostanoid receptors

Selective agonists are the pharmacological counterparts of antagonists and, as such, fulfil an important role in receptor characterization. In terms of prostanoid receptors, three of the natural ligands, PGH<sub>2</sub>/TXA<sub>2</sub> and PGI<sub>2</sub>, are unstable under physiological conditions and are usually replaced by U-46619 and cicaprost respectively (Table 1). In addition, there are potential problems with certain EP agonists synthesized as C1 methyl esters, for example, butaprost (EP<sub>2</sub>) and SC-46275 (EP<sub>3</sub>); full agonist potency is only realized after (enzymatic) hydrolysis of the ester within the tissue. Ono Pharmaceuticals have synthesized selective agonists for EP<sub>1</sub> (ONO-DI-004), EP<sub>2</sub> (ONO-AE1-259), EP<sub>3</sub> (ONO-AE-248) and EP<sub>4</sub> (ONO-AE1-329) receptors (Suzawa *et al.*, 2000). However, the rather modest potencies of ONO-DI-004 and ONO-AE-248 may restrict their utility in full Schild antagonism protocols (R.L. Jones *et al.*, 2008, submitted). Partial agonists for DP<sub>1</sub>, EP<sub>1</sub>, EP<sub>3</sub>, FP, IP and TP receptors are known (Table 1); their use in antagonist protocols may present difficulties of interpretation.

As drugs, notably for systemic administration, selective prostanoid mimetics are a high-risk proposition because of the myriad of unwanted side effects that may occur. As such, the future of prostanoid-based therapies appears to reside in the main in selective PG synthase inhibitors and prostanoid receptor antagonists.

### Antagonist protocols

The Schild protocol for inferring the nature of competition and determining the affinity constant of an antagonist remains the gold standard (see Colquhoun, 2007). It is applicable to both isolated tissue and the ever-increasing number of recombinant (rc) receptor/cell-based assay methods usually involving Ca<sup>2+</sup> mobilization or cAMP generation. While the

latter methods are given to high throughput and precision, care must be taken with high-affinity antagonists in the Ca<sup>2+</sup> assays as the agonist response is usually measured as the peak of the transient Ca<sup>2+</sup> signal, which may occur before re-equilibration of antagonist occupancy is complete. The majority of the data given in Table 2 relate to pA<sub>2</sub> values derived using the Schild protocol, with emphasis on human, guinea pig and rat isolated preparations. Binding data (preferably pK<sub>i</sub>) are given where functional information is not available. There are considerable binding data on mouse prostanoid receptors, but little affinity data in functional systems.

Inhibition-curve (or Cheng-Prusoff) protocols have been used infrequently in prostanoid receptor studies. Bley *et al.* (2006) estimated the pA<sub>2</sub> of the IP antagonists RO-1138452 and RO-3244794 using carbacyclin as the fixed-concentration agonist in a human rc-IP receptor – cAMP assay, but failed to use the modified form of the Cheng-Prusoff equation (Craig, 1993; Lazareno and Birdsall, 1993; Leff and Dougall, 1993). These protocols have the advantages of operating over a lower agonist concentration range than Schild protocols and providing direct observation of the rate of onset of antagonism. The latter is important in recognizing the slow approach to steady state that occurs with high-affinity antagonists at low concentration, and also with highly lipophilic antagonists (Jones *et al.*, 2008) that regularly emerge from combinatorial chemistry – high-throughput screening. Ultimately, it is important and even preferable to define the pharmacology in the human target tissue; human rc-receptor assays are a useful accompaniment. Successful drugs require the correct pharmacological attributes, but physical chemical properties are also important.

### DP<sub>1</sub> receptor antagonists

#### Development

Antagonists for, what we now know to be, the DP<sub>1</sub> receptor subtype, were first described in the 1970s and early 1980s. Examples include N-0164 (MacIntyre and Gordon, 1977), diphloretin phosphate (Westwick and Webb, 1978) and desacetyl-1-nantradol (Horne, 1984), but none of these compounds exhibit the potency and selectivity essential for unambiguous receptor classification. The simple xanthone-carboxylic acid AH-6809 (Keery and Lumley, 1988) has sufficient DP<sub>1</sub> affinity (pA<sub>2</sub> = 5.9–6.6, Table 2), but it has been mainly employed as an EP<sub>1</sub> antagonist (see later). Indeed, the hydantoin derivative, BW-A868C (Figure 1) has been the only selective, surmountable and competitive DP<sub>1</sub> antagonist (pA<sub>2</sub> > 9 for the human subtype) readily available to pharmacologists (Giles *et al.*, 1989; Lydford *et al.*, 1996c). The N-benzyl substituent is crucial for antagonist activity; in a related series of bicyclic-hydantoin analogues a progression from full agonism to virtually pure antagonism is seen with hydrogen, methyl, ethyl and *n*-propyl substituents on N<sup>10</sup> (Giles and Leff, 1992). BW-A868C also has low affinity (pA<sub>2</sub> = 5.1) for the EP<sub>4</sub> subtype (Lydford *et al.*, 1996c). ZK-138357 (Schering AG) is a moderate-affinity DP<sub>1</sub> antagonist (Table 2) with some structural similarity to BW-A868C.

Recently, the potential pathological role of PGD<sub>2</sub>, especially in allergic disorders, has been revived resulting in the discovery and evaluation of highly selective DP<sub>1</sub> antagonists of

**Table 2** Affinities of prostanoid receptor antagonists in functional isolated preparations

Antagonist	Species	Tissue system	Agonist	pA <sub>2</sub>	Reference
<b>DP<sub>1</sub> receptor</b>					
AH-6809	Human	Neutrophil/superoxide release	BW-245C	6.55	Wheeldon and Vardey (1993)
		Platelet/aggregation	BW-245C	6.59	Lydford <i>et al.</i> (1996a)
			PGD <sub>2</sub>	6.27 <sup>a</sup>	Keery and Lumley (1988)
	Cow	Embryonic tracheal fibroblast/cAMP	PGD <sub>2</sub>	6.36	Ito <i>et al.</i> (1990)
	Rabbit	Saphenous vein	BW-245C	5.93	Lydford <i>et al.</i> (1996c)
BW-A868C	Human	Neutrophil/superoxide release	BW-245C	9.46	Lydford <i>et al.</i> (1996a)
		Platelet/aggregation	BW-245C	9.26	Giles <i>et al.</i> (1989)
		Pulmonary vein	PGD <sub>2</sub>	7.84	Walch <i>et al.</i> (1999)
		Uterus (non-pregnant)	BW-245C	8.3	Senior <i>et al.</i> (1992)
		Uterus (pregnant)	BW-245C	8.6	Senior <i>et al.</i> (1993)
	Cow	Embryonic tracheal fibroblast/cAMP	BW-245C	8.0	Crider <i>et al.</i> (1999)
	Dog	Nasal vein	BW-245C	7.3	Liu <i>et al.</i> (1996a)
		Tracheal epithelium/Cl <sup>-</sup> secretion	BW-245C	8.16	Liu <i>et al.</i> (1996b)
	Rabbit	Jugular vein	BW-245C	8.73	Giles <i>et al.</i> (1989)
		Saphenous vein	BW-245C	8.50	Lydford <i>et al.</i> (1996c)
MK-0524 (Laropiprant)	Human	rc-DP <sub>1</sub> /HEK-293E/binding	[ <sup>3</sup> H]-PGD <sub>2</sub>	10.5 <sup>b</sup>	Sturino <i>et al.</i> (2007)
		Platelet/cAMP	PGD <sub>2</sub>	10.05 <sup>c</sup>	Sturino <i>et al.</i> (2007)
ONO-AE3-237	Human	rc-DP <sub>1</sub> /CHO/binding	[ <sup>3</sup> H]-PGD <sub>2</sub>	7.74	Torisu <i>et al.</i> (2004c)
S-5751	Human	Platelet/cAMP	PGD <sub>2</sub>	9.02 <sup>b,c</sup>	Arimura <i>et al.</i> (2001)
	Guinea pig	Platelet/cAMP	PGD <sub>2</sub>	7.50 <sup>b,c</sup>	Arimura <i>et al.</i> (2001)
ZK-138357	Human	Neutrophil/superoxide release	BW-245C	7.25	Lydford <i>et al.</i> (1996a)
	Rabbit	Saphenous vein	BW-245C	5.05	Lydford <i>et al.</i> (1996a)
	Rat	Peritoneal mast cell	BW-245C	-6.0	Chan <i>et al.</i> (2000)
Compound 1	Human	Platelet/binding	[ <sup>3</sup> H]-PGD <sub>2</sub>	6.22	Mitsumori <i>et al.</i> (2003a)
Compound 2	Human	Platelet/binding	[ <sup>3</sup> H]-PGD <sub>2</sub>	7.62	Mitsumori <i>et al.</i> (2003a)
Compound 3	Human	rc-DP <sub>1</sub> /HEK-293-Gα15/Ca <sup>2+</sup>	BW-245C	[-7.4]	Krauss <i>et al.</i> (2005)
Compound 4	Human	rc-DP <sub>1</sub> /CHO/binding	[ <sup>3</sup> H]-PGD <sub>2</sub>	8.27	Torisu <i>et al.</i> (2004c)
Compound 5	Human	rc-DP <sub>1</sub> /HEK-293/binding	[ <sup>3</sup> H]-PGD <sub>2</sub>	9.0	Beaulieu <i>et al.</i> (2008)
<b>DP<sub>2</sub> receptor</b>					
BAY-u3405 (Ramatroban)	Human	rc-DP <sub>2</sub> /CHO/GTPγS binding	PGD <sub>2</sub>	7.44	Mathiesen <i>et al.</i> (2006)
		Eosinophil/shape change	PGD <sub>2</sub>	-8.0 <sup>a</sup>	Mathiesen <i>et al.</i> (2006)
K-117	Human	rc-DP <sub>2</sub> /HEK-293/binding	[ <sup>3</sup> H]-PGD <sub>2</sub>	8.26	Mimura <i>et al.</i> (2005)
K-604	Human	rc-DP <sub>2</sub> /HEK-293/binding	[ <sup>3</sup> H]-PGD <sub>2</sub>	7.96	Mimura <i>et al.</i> (2005)
TM-30089 <sup>d</sup> (CAY-10471)	Human	rc-DP <sub>2</sub> /HEK-293/binding	[ <sup>3</sup> H]-PGD <sub>2</sub>	8.74	Mathiesen <i>et al.</i> (2006)
				9.22	Ulven and Kostenis (2005)
Compound 6	Human	rc-DP <sub>2</sub> /pre-B L1.2/Ca <sup>2+</sup>	PGD <sub>2</sub>	[6.8]	Bauer <i>et al.</i> (2002)
Compound 7	Human	rc-DP <sub>2</sub> /HEK-293/binding	[ <sup>3</sup> H]-PGD <sub>2</sub>	8.64 <sup>c</sup>	Birkinshaw <i>et al.</i> (2006)
Compound 8	Human	rc-DP <sub>2</sub> /HEK-293/binding	[ <sup>3</sup> H]-PGD <sub>2</sub>	[9.40] <sup>c</sup>	Bonnert and Rasul (2004)
Compound 9	Human	rc-DP <sub>2</sub> /CHO/binding	[ <sup>3</sup> H]-PGD <sub>2</sub>	7.17	Armer <i>et al.</i> (2005)
		Eosinophil/shape change	PGD <sub>2</sub>	7.13 <sup>c</sup>	Armer <i>et al.</i> (2005)
		Th2-lymphocyte/chemotaxis	PGD <sub>2</sub>	7.17 <sup>c</sup>	Armer <i>et al.</i> (2005)
Compound 10	Human	rc-DP <sub>2</sub> /CHO/Ca <sup>2+</sup>	PGD <sub>2</sub>	[8.53]	Fretz <i>et al.</i> (2005)
Compound 11	Human	rc-DP <sub>2</sub> /HEK-293/binding	[ <sup>3</sup> H]-PGD <sub>2</sub>	[9.0]	Bonnert <i>et al.</i> (2005c)
<b>EP<sub>1</sub> receptor</b>					
AH-6809	Human	rc-EP <sub>1</sub> /HEK-293E/reporter gene	Iloprost	-6.4	Durocher <i>et al.</i> (2000)
		Pulmonary vein	Sulprostone	5.52	Walch <i>et al.</i> (2001)
	Guinea pig	Ileum	PGE <sub>2</sub>	6.8	Coleman <i>et al.</i> (1987)
		Ileum	PGE <sub>2</sub>	7.39	Eglen and Whiting (1988)
		Ileum	PGE <sub>1</sub>	7.42	Eglen and Whiting (1988)
		Ileum	16,16-DM PGE <sub>2</sub>	7.59	Eglen and Whiting (1988)
		Trachea	16,16-DM PGE <sub>2</sub>	7.48	Eglen and Whiting (1988)
		Trachea	17-Phenyl PGE <sub>2</sub>	7.35	Lawrence <i>et al.</i> (1992)
GW-848687	Human	rc-EP <sub>1</sub> /not given/reporter gene	PGE <sub>2</sub>	9.1	Giblin <i>et al.</i> (2007)
MF-266-1	Human	rc-EP <sub>1</sub> /HEK-293/Ca <sup>2+</sup>	PGE <sub>2</sub>	7.8	Clark <i>et al.</i> (2008)
ONO-8711	Mouse	rc-EP <sub>1</sub> /CHO/binding	[ <sup>3</sup> H]-PGE <sub>2</sub>	8.77	Watanabe <i>et al.</i> (1999)
	Human	rc-EP <sub>1</sub> /CHO/binding	[ <sup>3</sup> H]-PGE <sub>2</sub>	9.22	Watanabe <i>et al.</i> (1999)
ONO-8713	Mouse	rc-EP <sub>1</sub> /not given/binding	[ <sup>3</sup> H]-PGE <sub>2</sub>	9.5	Narumiya and Fitzgerald (2001)
SC-19220	Guinea pig	Ileum	PGE <sub>2</sub>	5.5	Sanner (1969)
		Ileum	PGE <sub>2</sub>	5.6	Bennett and Posner (1971)
		Trachea	PGF <sub>2α</sub>	6.6	Farmer <i>et al.</i> (1974)
SC-51089	Human	rc-EP <sub>1</sub> /HEK-293E/reporter gene	Iloprost	6.94	Durocher <i>et al.</i> (2000)
	Guinea pig	Ileum	PGE <sub>2</sub>	6.5	Hallinan <i>et al.</i> (1993)
		Ileum	PGE <sub>2</sub>	6.7	Sametz <i>et al.</i> (2000)
SC-51322	Human	rc-EP <sub>1</sub> /HEK-293E/reporter gene	Iloprost	8.80	Durocher <i>et al.</i> (2000)
	Guinea pig	Ileum	PGE <sub>2</sub>	8.1	Hallinan <i>et al.</i> (1994)
		Trachea	17-Phenyl PGE <sub>2</sub>	8.45	Hung <i>et al.</i> (2006)
Compound 13	Mouse	rc-EP <sub>1</sub> /CHO/Ca <sup>2+</sup>	PGE <sub>2</sub>	8.25 <sup>c</sup>	Naganawa <i>et al.</i> (2006)
Compound 14	Human	rc-EP <sub>1</sub> /HEK-293E/binding	[ <sup>3</sup> H]-PGE <sub>2</sub>	8.0	Ruel <i>et al.</i> (1999)
Compound 15	Human	rc-EP <sub>1</sub> /CHO/Ca <sup>2+</sup>	PGE <sub>2</sub>	8.2	Hall <i>et al.</i> (2007b)
<b>EP<sub>2</sub> receptor</b>					
AH-6809	Human	rc-EP <sub>2</sub> /COS-7/cAMP	PGE <sub>2</sub>	-6.5	Woodward <i>et al.</i> (1995)
		Bronchus	PGE <sub>2</sub>	5.78	Norel <i>et al.</i> (1999)
	Guinea pig	Trachea	PGE <sub>2</sub>	5.7 <sup>e</sup>	KJ Ong and RL Jones (unpublished)

Table 2 Continued

Antagonist	Species	Tissue system	Agonist	pA <sub>2</sub>	Reference
EP <sub>3</sub> receptor					
DG-041	Human	rc-EP <sub>3</sub> /Chem-1/Ca <sup>2+</sup>	PGE <sub>2</sub>	8.09 <sup>c</sup>	Singh <i>et al.</i> (2009)
L-798106	Guinea pig	Aorta	17-Phenyl PGE <sub>2</sub> <sup>f</sup>	7.96	Jones <i>et al.</i> (2008)
		Trachea	Sulprostone	7.82	Clarke <i>et al.</i> (2004)
		Vas deferens	Sulprostone	7.48	Clarke <i>et al.</i> (2004)
L-826266	Human	rc-EP <sub>3</sub> /HEK-293E/Ca <sup>2+</sup>	PGE <sub>2</sub>	7.97	R.L. Jones <i>et al.</i> (2008, submitted)
		Erythroleukaemia cell/cAMP	Sulprostone	8.35	Clark <i>et al.</i> (2008)
	Guinea pig	Aorta	17-Phenyl PGE <sub>2</sub>	7.58	R.L. Jones <i>et al.</i> (2008, submitted)
ONO-AE3-240	Mouse	rc-EP <sub>3</sub> /not given/Ca <sup>2+</sup>	PGE <sub>2</sub>	8.8	Amano <i>et al.</i> (2003)
Compound 17	Human	Erythroleukaemia cell/cAMP	Sulprostone	6.89	Gallant <i>et al.</i> (2002)
Compound 18	Human	rc-EP <sub>3</sub> /HEK-293E/binding	[ <sup>3</sup> H]-PGE <sub>2</sub>	7.7	Juteau <i>et al.</i> (2001)
Compound 19	Human	rc-EP <sub>3</sub> /HEK-293E/cAMP	PGE <sub>2</sub>	8.22	Belley <i>et al.</i> (2005)
EP <sub>4</sub> receptor					
AH-23848	Sheep	Ductus arteriosus	PGE <sub>2</sub>	-5.2 <sup>a</sup>	Bouayad <i>et al.</i> (2001)
	Human	Middle cerebral artery	PGE <sub>2</sub>	5.7	Davis <i>et al.</i> (2004)
	Pig	Saphenous vein	PGE <sub>2</sub>	5.0	Coleman <i>et al.</i> (1994a)
	Rabbit	Saphenous vein	PGE <sub>2</sub>	4.96	Lydford <i>et al.</i> (1996b)
	Mouse	rc-EP <sub>4</sub> /CHO/cAMP	PGE <sub>2</sub>	5.3	Nishigaki <i>et al.</i> (1995)
BGC-20-1531	Human	rc-EP <sub>4</sub> /HEK-293E/cAMP	PGE <sub>2</sub>	7.6	Maubach <i>et al.</i> (2009)
		Middle cerebral artery	PGE <sub>2</sub>	7.8	Maubach <i>et al.</i> (2009)
	Dog	Middle meningeal artery	PGE <sub>2</sub>	7.7	Maubach <i>et al.</i> (2009)
CJ-023423	Human	rc-EP <sub>4</sub> /HEK-293/cAMP	PGE <sub>2</sub>	8.3	Nakao <i>et al.</i> (2007)
	Rat	rc-EP <sub>4</sub> /HEK-293/cAMP	PGE <sub>2</sub>	8.2	Nakao <i>et al.</i> (2007)
CJ-042794	Human	rc-EP <sub>4</sub> /HEK-293/cAMP	PGE <sub>2</sub>	8.6	Murase <i>et al.</i> (2008b)
	Rat	rc-EP <sub>4</sub> /HEK-293/cAMP	PGE <sub>2</sub>	8.7	Murase <i>et al.</i> (2008a)
GW-627368	Human	rc-EP <sub>4</sub> /HEK-293/cAMP	PGE <sub>2</sub> <sup>b</sup>	7.9	Wilson <i>et al.</i> (2006)
		Pulmonary vein	ONO-AE1-329	7.06	Foudi <i>et al.</i> (2008)
	Pig	Saphenous vein	PGE <sub>2</sub>	9.2	Wilson <i>et al.</i> (2006)
	Rabbit	Saphenous vein	PGE <sub>2</sub> <sup>g</sup>	≥8.5	Jones and Chan. (2005)
L-161982	Human	rc-EP <sub>4</sub> /HEK-293/cAMP	PGE <sub>2</sub>	-8.5	Machwate <i>et al.</i> (2001)
		BEAS-2B cell/CRE reporter	ONO-AE1-329	9.14	L.M. Ayer and M.A. Giembycz (unpublished)
		Middle cerebral artery	PGE <sub>2</sub>	8.4	Davis <i>et al.</i> (2004)
	Rat	rc-EP <sub>4</sub> /HEK-293/binding	[ <sup>3</sup> H]-PGE <sub>2</sub>	7.50	Machwate <i>et al.</i> (2001)
		Periosteal cell/cAMP	PGE <sub>2</sub>	7.0 <sup>c</sup>	Machwate <i>et al.</i> (2001)
MF-498	Human	rc-EP <sub>4</sub> /HEK-293/cAMP	PGE <sub>2</sub>	8.77 <sup>c</sup>	Clark <i>et al.</i> (2008)
ONO-AE2-227	Mouse	rc-EP <sub>4</sub> /CHO/cAMP	PGE <sub>2</sub>	8.0 <sup>c</sup>	Mutoh <i>et al.</i> (2002)
ONO-AE3-208	Mouse	rc-EP <sub>4</sub> /not given/binding	[ <sup>3</sup> H]-PGE <sub>2</sub>	8.89	Kabashima <i>et al.</i> (2002)
Compound 20	Human	rc-EP <sub>4</sub> /HEK-293E/cAMP	PGE <sub>2</sub>	8.49 <sup>c</sup>	Burch <i>et al.</i> (2008)
FP receptor					
AS-604872	Human	rc-FP/HEK-293E/PI	PGF <sub>2α</sub>	7.33 <sup>c</sup>	Cirillo <i>et al.</i> (2007)
THG-113	Pig	Retinal blood vessel	PGF <sub>2α</sub>	[6.34] <sup>c</sup>	Peri <i>et al.</i> (2006)
THG-113.31	Pig	Retinal blood vessel	PGF <sub>2α</sub>	[8.00] <sup>c</sup>	Peri <i>et al.</i> (2006)
THG-113.824	Pig	Retinal blood vessel	PGF <sub>2α</sub>	[8.96] <sup>c</sup>	Peri <i>et al.</i> (2006)
THG-113.825	Pig	Retinal blood vessel	PGF <sub>2α</sub>	7.21 <sup>c</sup>	Peri <i>et al.</i> (2006)
Prostamide F receptor					
AGN-204396	Cat	Iris sphincter	Prostamide F <sub>2α</sub>	5.64	Woodward <i>et al.</i> (2007)
IP receptor					
RO-1138452	Human	rc-IP/CHO/cAMP	Carbacyclin	9.0	Bley <i>et al.</i> (2006)
		Pulmonary artery	Cicaprost	8.20	Jones <i>et al.</i> (2006)
	Guinea pig	Aorta	Cicaprost	8.39	Jones <i>et al.</i> (2006)
	Rabbit	Mesenteric artery	Cicaprost	8.12	Jones <i>et al.</i> (2006)
RO-3244794	Human	rc-IP/CHO/cAMP	Carbacyclin	8.5	Bley <i>et al.</i> (2006)
		BEAS-2B cell/CRE reporter	Taprostene	9.24	L.A. Ayer and M.A. Giembycz (unpublished)
		rc-IP/not given/cAMP	(Not given)	[8.12] <sup>h</sup>	Keitz <i>et al.</i> (2004)
Compound 21	Rat	UMR-106 osteosarcoma cell/cAMP	Iloprost	6.41	Nakae <i>et al.</i> (2005)
Compound 22	Rat	UMR-106 osteosarcoma cell/cAMP	Iloprost	6.32	Nakae <i>et al.</i> (2005)
Compound 23	Rat	UMR-106 osteosarcoma cell/cAMP	Iloprost	6.32	Nakae <i>et al.</i> (2005)
Compound 24	Human	Platelet membrane/cAMP	Iloprost	7.8 <sup>c</sup>	Brescia <i>et al.</i> (2007)
TP receptor					
AA-2414	Human	Bronchus	U-46619	7.7	Itoh <i>et al.</i> (1993)
(Seratrodast)	Pig	Coronary artery	U-44069	9.0	Imura <i>et al.</i> (1990)
	Guinea pig	Aorta	U-46619	8.5	Zhang <i>et al.</i> (1996)
		Trachea	U-46619	7.69	Ashida <i>et al.</i> (1989)
	Rat	Aorta	U-46619	7.8	Zhang <i>et al.</i> (1996)
AH-23848	Human	Bronchus	U-46619	8.5	Coleman and Sheldrick (1989)
		Bronchus	U-46619	6.9	McKenniff <i>et al.</i> (1988)
		Hand vein	U-46619	8.4	Arner <i>et al.</i> (1991)
		Lung parenchyma	U-46619	8.7	McKenniff <i>et al.</i> (1988)
		Platelet/agggregation	U-46619	8.05	Tymkewycz <i>et al.</i> (1991)
	Guinea pig	Lung parenchyma	U-46619	8.7	McKenniff <i>et al.</i> (1988)
		Trachea	U-46619	8.7	McKenniff <i>et al.</i> (1988)
		Trachea	U-46619	9.76	Tymkewycz <i>et al.</i> (1991)

Table 2 Continued

Antagonist	Species	Tissue system	Agonist	pA <sub>2</sub>	Reference	
BAY-u3405 (Ramatroban)	Rat	Aorta	U-46619	8.47	Tymkewycz <i>et al.</i> (1991)	
		Lung parenchyma	U-46619	6.9	Norman <i>et al.</i> (1992)	
		Platelet/agggregation	U-46619	8.19	Tymkewycz <i>et al.</i> (1991)	
	Human	Bronchus	U-46619	8.8	McKenniff <i>et al.</i> (1991)	
		Pulmonary vein	U-46619	8.94	Walch <i>et al.</i> (2001)	
	Guinea pig	Lung parenchyma	U-46619	7.7	Norman <i>et al.</i> (1992)	
		Trachea	U-46619	8.7	McKenniff <i>et al.</i> (1991)	
BM-13177 (Sulotroban)	Rat	Lung parenchyma	U-46619	8.6	McKenniff <i>et al.</i> (1991)	
	Human	Platelet/agggregation	U-46619	6.31	Karasawa <i>et al.</i> (1991a)	
	Guinea pig	Aorta	U-46619	5.58	Karasawa <i>et al.</i> (1991b)	
BM-13505	Rabbit	Jugular vein	U-46619	6.01	Giles <i>et al.</i> (1989)	
		Hand vein	U-46619	7.9	Arner <i>et al.</i> (1991)	
	Human	Platelet/agggregation	U-46619	7.75	Karasawa <i>et al.</i> (1991a)	
		Uterus (non-pregnant)	U-46619	7.4	Senchyna and Crankshaw (1996)	
		Guinea pig	Aorta	U-46619	6.89	Dubé <i>et al.</i> (1992)
			Aorta	U-46619	7.22	Karasawa <i>et al.</i> (1991b)
			Aorta	U-46619	7.3	Zhang <i>et al.</i> (1996)
		Lung parenchyma	U-46619	7.0	Norman <i>et al.</i> (1992)	
		Trachea	U-46619	7.73	Dubé <i>et al.</i> (1992)	
		Trachea	U-46619	7.5	Ogletree and Allen (1992)	
Rat	Aorta	U-46619	8.2	Zhang <i>et al.</i> (1996)		
	Aorta	U-46619	8.6	Ogletree and Allen (1992)		
BMS-180291 (Ifetroban)	Human	Lung parenchyma	U-46619	7.5	Norman <i>et al.</i> (1992)	
		Platelet membrane/binding	[ <sup>3</sup> H]-SQ-29548	8.4	Ogletree <i>et al.</i> (1993)	
	Guinea pig	Aorta	U-46619	9.8	Zhang <i>et al.</i> (1996)	
CV-4151	Human	Platelet/agggregation	U-46619	5.2	Watts <i>et al.</i> (1991)	
	Rabbit	Aorta	U-44069	5.9	Imura <i>et al.</i> (1988)	
EP-092	Human	Bronchus	U-46619	7.37	Featherstone <i>et al.</i> (1990)	
		Bronchus	U-46619	6.8	McKenniff <i>et al.</i> (1988)	
		Lung parenchyma	U-46619	8.9	McKenniff <i>et al.</i> (1988)	
		Platelet /agggregation	U-46619	7.73	Tymkewycz <i>et al.</i> (1991)	
		Uterine artery	U-46619	8.5	Baxter <i>et al.</i> (1995)	
	Guinea pig	Lung parenchyma	U-46619	8.7	McKenniff <i>et al.</i> (1988)	
		Trachea	U-46619	7.29	Featherstone <i>et al.</i> (1990)	
		Trachea	U-46619	8.7	McKenniff <i>et al.</i> (1988)	
		Trachea	U-46619	8.02	Tymkewycz <i>et al.</i> (1991)	
		Aorta	U-46619	8.55	Tymkewycz <i>et al.</i> (1991)	
EP-169	Rat	Lung parenchyma	U-46619	7.1	Norman <i>et al.</i> (1992)	
		Platelet agggregation	U-46619	7.80	Tymkewycz <i>et al.</i> (1991)	
		Platelet/agggregation	U-46619	8.30	Tymkewycz <i>et al.</i> (1991)	
	Human	Pulmonary artery	U-46619	~8.2	Qian <i>et al.</i> (1994)	
		Trachea	U-46619	8.77	Tymkewycz <i>et al.</i> (1991)	
		Trachea	U-46619	8.73	Tymkewycz <i>et al.</i> (1991)	
Glibenclamide	Rat	Platelet/agggregation	U-46619	8.48	Tymkewycz <i>et al.</i> (1991)	
		Internal mammary artery	U-46619	6.3 <sup>i</sup>	Stanke <i>et al.</i> (1998)	
	Human	Saphenous vein	U-46619	6.7 <sup>i</sup>	Stanke <i>et al.</i> (1998)	
		Coronary artery	U-46619	6.2	Cocks <i>et al.</i> (1990)	
		Aorta	U-46619	6.08 <sup>a</sup>	Pfister <i>et al.</i> (2004)	
		Aorta	U-46619	<5.0	Kemp and McPherson (1998)	
		Aorta	U-46619	6.13	Kemp and McPherson (1998)	
GR-32191 (Vapiprost)	Human	Bladder (detrusor)	U-46619	8.27	Palea <i>et al.</i> (1998)	
		Bronchus	U-46619	8.77	Featherstone <i>et al.</i> (1990)	
		Bronchus	U-46619	8.40	Armour <i>et al.</i> (1989)	
	Guinea pig	Platelet/binding	[ <sup>3</sup> H]-GR-32191	8.66	Armstrong <i>et al.</i> (1993)	
		Pulmonary artery	U-46619	8.18	Lumley <i>et al.</i> (1989)	
		Saphenous vein	U-46619	8.93	Furci <i>et al.</i> (1991)	
		Umbilical artery	U-46619	8.0	Boersma <i>et al.</i> (1999)	
		Uterine artery	U-46619	8.5	Baxter <i>et al.</i> (1995)	
		Uterus (non-pregnant)	U-46619	8.6	Senchyna and Crankshaw (1996)	
		Uterus (pregnant)	U-46619	8.5 <sup>a</sup>	Senior <i>et al.</i> (1993)	
		Rat	Aorta	U-46619	8.77	Lumley <i>et al.</i> (1989)
			Aorta	U-46619	9.4	Ogletree and Allen (1992)
			Trachea	U-46619	8.26	Featherstone <i>et al.</i> (1990)
GR-83783 <sup>j</sup> GR-108774 <sup>k</sup> ICI-192605	Human	Trachea	U-46619	9.43	Tymkewycz <i>et al.</i> (1991)	
		Trachea	U-46619	10.0	Ogletree and Allen (1992)	
	Rat	Aorta	U-46619	7.87	Lumley <i>et al.</i> (1989)	
		Aorta	U-46619	7.49	Furci <i>et al.</i> (1991)	
		Aorta	U-46619	8.41	Tymkewycz <i>et al.</i> (1991)	
		Aorta	U-46619	8.3	Ogletree and Allen (1992)	
		Trachea	U-46619	8.31	Lydford and McKechnie (1994)	
		Aorta	U-46619	7.5	Campbell <i>et al.</i> (1991a)	
		Aorta	U-46619	9.2	Campbell <i>et al.</i> (1991b)	
		Platelet/agggregation	U-46619	8.16	Brewster <i>et al.</i> (1988)	
Human	Umbilical artery	U-46619	8.1	Boersma <i>et al.</i> (1999)		
	Umbilical vein	U-46619	9.07	Daray <i>et al.</i> (2003)		
	Uterus (non-pregnant)	U-46619	9.2	Senchyna and Crankshaw (1996)		
Rat	Aorta	U-46619	8.4	Brewster <i>et al.</i> (1988)		

Table 2 Continued

Antagonist	Species	Tissue system	Agonist	pA <sub>2</sub>	Reference
I-PTA-OH	Guinea pig	Lung parenchyma	U-46619	5.6	Norman <i>et al.</i> (1992)
	Rat	Lung parenchyma	U-46619	5.8	Norman <i>et al.</i> (1992)
I-SAP	Human	Platelet/aggregation	U-46619	8.01 <sup>k</sup>	Naka <i>et al.</i> (1992)
KW-3635	Human	Platelet/aggregation	U-46619	8.88	Karasawa <i>et al.</i> (1991a)
	Guinea pig	Aorta	U-46619	7.74	Karasawa <i>et al.</i> (1991b)
L-655240	Guinea pig	Aorta	U-44069	8.0	Hall <i>et al.</i> (1987)
	Guinea pig	Aorta	U-44069	8.0	Hall <i>et al.</i> (1987)
ONO-11120	Human	Platelet/binding	[ <sup>125</sup> I]-PTA-OH	7.71 <sup>c</sup>	Narumiya <i>et al.</i> (1986)
		Platelet/aggregation	U-46619	7.49	Tymkewycz <i>et al.</i> (1991)
	Guinea pig	Trachea	U-46619	8.07	Tymkewycz <i>et al.</i> (1991)
	Rat	Aorta	U-46619	7.14	Tymkewycz <i>et al.</i> (1991)
		Platelet/aggregation	U-46619	7.38	Tymkewycz <i>et al.</i> (1991)
ONO-NT-126	Human	Astrocytoma cell/PI	STA <sub>2</sub>	10.0	Nakahata <i>et al.</i> (1990)
Ridogrel <sup>l</sup>	Human	Platelet/aggregation	U-46619	5.7	Watts <i>et al.</i> (1991)
	Rat	Tail artery	U-46619	5.5	Janssens <i>et al.</i> (1990)
(±)-S-145	Human	Astrocytoma cell/PI	STA <sub>2</sub>	8.48	Nakahata <i>et al.</i> (1990)
(Domitroban) <sup>l</sup>		Platelet membrane/binding	[ <sup>3</sup> H]-(-)-S-145	9.35	Kishino <i>et al.</i> (1991)
	Rat	Aorta smooth muscle cell/binding	[ <sup>3</sup> H]-SQ-29548	9.5 <sup>a</sup>	Hanasaki <i>et al.</i> (1988)
S-18886	Rabbit	Saphenous vein	U-46619	8.9	Cimetière <i>et al.</i> (1998)
(Terutroban)					
SQ-29548	Human	Astrocytoma cell/PI	STA <sub>2</sub>	8.08	Nakahata <i>et al.</i> (1990)
		Immortalized ciliary epithelial cell/PI	U-46619	7.7 <sup>m</sup>	Sharif <i>et al.</i> (2002)
		Corpus cavernosum	U-46619	9.0	Angulo <i>et al.</i> (2002)
		Umbilical artery	U-46619	7.6	Boersma <i>et al.</i> (1999)
		Umbilical vein	U-46619	7.96	Daray <i>et al.</i> (2003)
		Uterus (non-pregnant)	U-46619	8.2	Senchyna and Crankshaw (1996)
	Pig	Coronary artery	U-46619	8.8 <sup>a</sup>	Kromer and Tippins (1996)
	Rabbit	Aorta	U-46619	7.95	Yoshida <i>et al.</i> (2007)
	Guinea pig	Aorta	U-46619	7.96	Dubé <i>et al.</i> (1992)
		Aorta	U-46619	8.9	Ogletree and Allen (1992)
		Aorta	U-46619	8.5	Zhang <i>et al.</i> (1996)
		Lung parenchyma	U-46619	7.7	Norman <i>et al.</i> (1992)
		Trachea	U-46619	8.70	Dubé <i>et al.</i> (1992)
		Trachea	U-46619	8.9	Ogletree and Allen (1992)
	Rat	Aorta	U-46619	9.2	Zhang <i>et al.</i> (1996)
		Lung parenchyma	U-46619	7.2	Norman <i>et al.</i> (1992)
SQ-30741	Human	Coronary artery	U-46619	7.54	Maassen VanDenBrink <i>et al.</i> , 1996)
	Human	Umbilical artery	U-46619	7.0	Boersma <i>et al.</i> (1999)
	Guinea pig	Aorta	U-46619	8.1	Ogletree and Allen (1992)
		Trachea	U-46619	8.6	Ogletree and Allen (1992)
	Rat	Aorta	U-46619	7.9	Ogletree and Allen (1992)
YM-158	Guinea pig	Trachea	U-46619	8.81 <sup>n</sup>	Arakida <i>et al.</i> (1998)
Z-335	Human	Platelet membrane/binding	[ <sup>3</sup> H]-SQ-29548	7.52	Tanaka <i>et al.</i> (1998)
		Platelet/shape change <sup>o</sup>	U-46619	8.02	Yoshida <i>et al.</i> (2007)
	Rabbit	Aorta	U-46619	8.64	Yoshida <i>et al.</i> (2007)
ZD-1542 <sup>i</sup>	Guinea pig	Lung parenchyma	U-46619	8.5	Brownlie <i>et al.</i> (1993)
		Trachea	U-46619	8.3	Brownlie <i>et al.</i> (1993)
	Rat	Aorta	U-46619	8.51	Brownlie <i>et al.</i> (1993)

pA<sub>2</sub> values relate to functional assays. Recombinant (rc-) systems: prostanoid receptor followed by the carrier cell line and second messenger measurement. Smooth muscle preparations: contraction or relaxation of induced tone. Platelets: all data from plasma-free platelet suspensions. Where pA<sub>2</sub> values are not available, pK<sub>i</sub>/pK<sub>D</sub> values derived from radioligand binding are given (italics). Affinity values in square brackets derive from the patent literature.

Structures of compounds: 1–5, Figure 1; 6–12, Figure 2; 13–15, Figure 3; 16–19, Figure 4; 20, Figure 5; 21–24, Figure 7.

BEAS, human bronchial epithelium; CHO, Chinese hamster ovary; COS-7, African green monkey kidney; HEK, human embryonic kidney; CRE, cAMP response element; 16,16-DM PGE<sub>2</sub>, 16,16-dimethyl PGE<sub>2</sub>; PI, phosphoinositide.

<sup>a</sup>Our calculation.

<sup>b</sup>Appreciable affinity for corresponding TP receptor.

<sup>c</sup>pIC<sub>50</sub>.

<sup>d</sup>Insurmountable antagonism in functional DP<sub>2</sub> systems.

<sup>e</sup>1 μM SC-51322 present.

<sup>f</sup>Replacement for sulprostone, which had slow onset and offset.

<sup>g</sup>Possible interference by EP<sub>2</sub> system.

<sup>h</sup>pK<sub>i</sub>.

<sup>i</sup>pK<sub>b</sub> for non-competitive antagonism.

<sup>j</sup>Combined TP antagonist/TXS inhibitor.

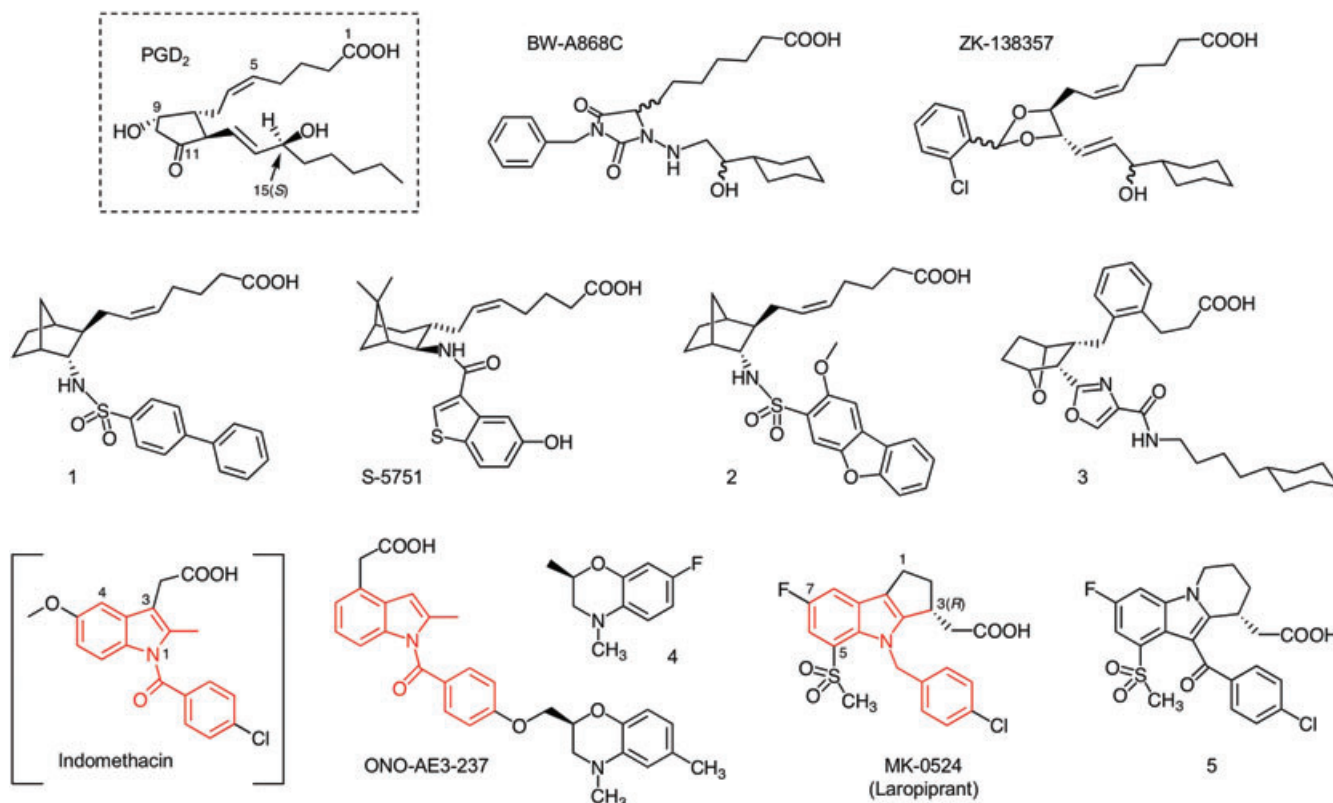
<sup>k</sup>Shape change seen.

<sup>l</sup>Data for (+)- and (-)-enantiomers in Kishino *et al.* (1991).

<sup>m</sup>Non-competitive antagonism.

<sup>n</sup>pA<sub>2</sub> = 8.87 for LTD<sub>4</sub> antagonism.

<sup>o</sup>Suppression of maximum aggregation response.



**Figure 1** DP<sub>1</sub> receptor antagonists. The natural ligand PGD<sub>2</sub> is shown in the box; the *trans*-orientation of the  $\alpha$  (upper) and  $\omega$  (lower) side-chains and (S) configuration at C15 are found in primary products of all COX/synthase systems. BW-A868C and ZK-138357 are each composed of four diastereoisomers (chiral centres at C8/C15 and C10/C15 respectively); compound **1** is racemic. Indomethacin is a lead compound for the non-prostanoid antagonists shown in the lower row; *N*-benzoyl-2-methyl-indol-3-yl-acetic acid templates are shown in red. The 3(S)-enantiomer of MK-0524 has 320-fold lower affinity for the human DP<sub>1</sub> receptor (Sturino *et al.*, 2007).

several structural classes. The most important of these (Figure 1) and their therapeutic applications are described below.

**Bicycloheptanes.** Chemists in Shionogi have synthesized selective DP<sub>1</sub> antagonists containing a bicyclo[2.2.1]heptane ring system akin to that present in PGH<sub>2</sub> (see inset in Figure 8) (Tsuru *et al.*, 1997; Honma *et al.*, 1998; Mitsumori *et al.*, 2003a). An initial lead was the racemic compound **1** previously shown to be a TP antagonist (Narisada *et al.*, 1988). Subsequent structure–activity relationship (SAR) studies revealed that a 6,6-dimethylbicyclo[3.1.1]heptane (pinane) system could substitute for the bicycloheptane ring (Tsuru *et al.*, 1997; Mitsumori *et al.*, 2003b; Yoshikawa *et al.*, 2005) and carbonylamino or sulphonylamino linkages to the  $\omega$ -aryl moiety were required for potent DP<sub>1</sub> antagonism (Tsuru *et al.*, 1997; Honma *et al.*, 1998; Mitsumori *et al.*, 2003b). Accordingly, S-5751 has high affinity for the DP<sub>1</sub> receptor ( $pK_i = 8.8$ ) and is orally active in models of allergy and inflammation in the guinea pig (Tsuru *et al.*, 1997; Arimura *et al.*, 2001; Mitsumori *et al.*, 2003b; Yasui *et al.*, 2008). In addition, lead optimization of the (+)-isomer of compound **1**, which has much higher DP<sub>1</sub> selectivity than its mirror-image, led to the 1-methoxy-dibenzo[*b,d*]furan **2**. This antagonist is potent, orally bioavailable and efficacious in guinea pig models of conjunctivitis and allergen-induced bronchoconstriction (Mitsumori *et al.*, 2003a).

Allergan have also filed patents claiming DP<sub>1</sub> receptor antagonism based on a 1(S),4(S)-7-oxabicyclo[2.2.1]heptane scaffold (Krauss *et al.*, 2005). One of these, compound **3**, has a  $pA_2$  of  $\sim 7.4$  for the human rc-DP<sub>1</sub> receptor and represents a logical structure for lead optimization.

**Indole acetic acids.** A non-prostanoid exploited for DP<sub>1</sub> antagonism at Ono Pharmaceuticals is the cyclo-oxygenase (COX) inhibitor/non-steroidal anti-inflammatory drug (NSAID), indomethacin (Figure 1). Initial studies showed that the acetic acid moiety could be switched to position 4 on the indole ring (Torisu *et al.*, 2004a). Optimization led to the discovery of two benzoxazines, ONO-AE3-237 and compound **4**, with high DP<sub>1</sub> antagonist selectivity and (sub)nanomolar affinity (Torisu *et al.*, 2004b,c,d; Torisu *et al.*, 2005). Administered orally, both compounds effectively suppressed PGD<sub>2</sub>- and allergen-induced vascular permeability in the guinea pig conjunctiva (Torisu *et al.*, 2004c). Pharmacokinetic studies on ONO-AE3-237 given by the oral (10 mg·kg<sup>-1</sup>) and intravenous (1 mg·kg<sup>-1</sup>) routes to fasted rats afforded plasma half-lives of 7.8 and 9.2 h respectively. The compound has a high volume of distribution indicating good tissue penetration and is 48% bioavailable at a dose of 10 mg·kg<sup>-1</sup> p.o. (Torisu *et al.*, 2004c).

Merck Frosst (Wang *et al.*, 2002; Berthelette *et al.*, 2003) and Sanofi-Aventis (Yang *et al.*, 2008) also filed patents claiming DP<sub>1</sub> antagonists with further variation of the acetic acid position on the indole template. Screening of the Merck



compound collection identified a difluoro-indole that had low-nanomolar affinity for DP<sub>1</sub> and TP receptors (Sturino *et al.*, 2006) and this, and other analogues, were optimized resulting in two 7-methylsulphone derivatives with high DP<sub>1</sub> affinity ( $K_i$  ~2 nM) and at least 100-fold selectivity over other prostanoid receptors. However, both compounds displayed poor pharmacokinetics in the rat, in particular extensive biliary excretion (Sturino *et al.*, 2007). This liability was overcome when the 5-substituent and the 7-methylsulphone on the indole ring were replaced by methylsulphone and fluorine, respectively, to give MK-0542 (laropiprant; Figure 1). Laropiprant has very high affinity for the human rc-DP<sub>1</sub> receptor ( $pK_i$  = 10.5), with ~300-fold lower affinity for the corresponding TP receptor (Sturino *et al.*, 2007). Excellent pharmacokinetic profiles have been found in the rat, dog, monkey and man (Chang *et al.*, 2007; Karanam *et al.*, 2007; Sturino *et al.*, 2007; Lai *et al.*, 2008b). Given orally to healthy male volunteers, laropiprant at single doses up to 900 mg and multiple doses up to 450 mg is rapidly absorbed ( $T_{max}$  = 0.8–2 h), demonstrates dose-proportional systemic exposure, has a half-life of 12–18 h and is generally well tolerated; this pharmacokinetic profile is unaffected by food (Karanam *et al.*, 2007; Lai *et al.*, 2008b). At a dose of 6 mg, laropiprant was effective in antagonizing PGD<sub>2</sub>-induced cAMP accumulation in human platelets *ex vivo*, indicating an interaction with the desired molecular target. Evidence for TP receptor blockade was also detected, but this effect was deemed not to be clinically relevant (Lai *et al.*, 2008b).

Merck Frosst has also disclosed 'backup' DP<sub>1</sub> antagonists, in which the indole template present in laropiprant is inverted. The tetrahydropyridindole **5** (Figure 1) exhibited the best profile ( $pK_i$  = 9.0 and 6.8 at DP<sub>1</sub> and TP receptors respectively) and is considered a suitable candidate for development (Beaulieu *et al.*, 2008).

**Other structural classes.** Certain aminopyrimidines have also been claimed in the patent literature to be DP<sub>1</sub> antagonists but neither *in vitro* nor *in vivo* pharmacological data are yet available (Langevin *et al.*, 2007; Stefany *et al.*, 2007).

#### Therapeutic applications

PGD<sub>2</sub> is an established mediator of allergic disease. It is the major prostanoid released from mast cells (Lewis *et al.*, 1982; Peters *et al.*, 1982) and is also secreted, albeit in lower amounts, by T-lymphocytes of the Th2 subset (Tanaka *et al.*, 2000). In asthma, dermatitis and rhinitis, allergen challenge leads to the rapid production of PGD<sub>2</sub> (Naclerio *et al.*, 1983; Murray *et al.*, 1986; Charlesworth *et al.*, 1991) and PGD<sub>2</sub>, itself, can reproduce many symptoms associated with allergic phenomena (see Pettipher, 2008). However, therapeutic/commercial success has not yet been attained with DP<sub>1</sub> antagonists probably because functional and subsequently molecular evidence emerged for a second subtype of PGD<sub>2</sub>-sensitive receptor that is strongly implicated in several manifestations of allergic disease including eosinophil infiltration, mucus hyper-secretion and plasma extravasation (see next section).

In terms of allergic inflammation, activation of DP<sub>1</sub> receptors is known to mediate pathological changes in blood flow. In allergic rhinitis, vessels within the nasal mucosa become

engorged leading to congestion and the release of plasma proteins, which contribute to enhanced nasal secretions. These effects are mimicked by PGD<sub>2</sub>, which explains the limited efficacy of histamine H<sub>1</sub> antagonists in allergic rhinitis (see Pettipher, 2008). Sturino *et al.* (2007) have shown that laropiprant abolishes the marked increase in nasal airway resistance induced by intranasal instillation of PGD<sub>2</sub> in conscious sheep. Significantly, comparable data have also been obtained in 15 healthy, non-smoking male volunteers in whom laropiprant (25 mg or 100 mg q.d. for 3 days) significantly suppressed PGD<sub>2</sub>-induced nasal congestion (Van Hecken *et al.*, 2007). Interestingly, PGD<sub>2</sub> fails to lower diastolic blood pressure (BP) in human volunteers (Heavey *et al.*, 1984) although (BW)-192C86, a DP<sub>1</sub> partial agonist (Gray *et al.*, 1992), is an effective depressor and BW-245C evokes adverse cardiovascular effects consistent with vasodilatation (Al Sinawi *et al.*, 1985). The reason for this discrepancy is unknown, but given that PGD<sub>2</sub> is a potent vasoconstrictor in several species (Jones, 1976; 1978) and the DP<sub>2</sub> receptor is expressed in the aorta (Nagata and Hirai, 2003) and potentially other blood vessels, its activation may oppose the BP-lowering activity of PGD<sub>2</sub> acting via the DP<sub>1</sub> receptor.

The ability of PGD<sub>2</sub> to promote adverse vasodilatation is also associated with the therapeutic use of niacin (vitamin B<sub>3</sub>), which, in high doses, is used clinically to lower plasma cholesterol. Niacin may also have anti-oxidant and anti-inflammatory activity and is used, in conjunction with statins, to treat dyslipidaemia (see Kamanna *et al.*, 2008). The adverse effects of niacin, particularly flushing, are due to receptor (GPR109A)-mediated release from Langherhans' cells of PGD<sub>2</sub> and PGE<sub>2</sub>, which promote vasodilatation of dermal and cerebrovascular capillaries through activation of DP<sub>1</sub>, EP<sub>2</sub> and EP<sub>4</sub> receptors. Merck (Paolini *et al.*, 2008) and Sanofi-Aventis (Harris, 2008) have claimed that a DP<sub>1</sub> antagonist could limit the cardiovascular liability of niacin if given as a combination therapy, thereby providing a better-tolerated drug. Indeed, laropiprant significantly reduced niacin-induced flushing in normal and dyslipidaemic subjects when compared with niacin alone (Cheng *et al.*, 2006; Lai *et al.*, 2007; Paolini *et al.*, 2008). Accordingly, these and other findings led Merck to develop the investigational combination product *Cordaptive*, which significantly reduced the vascular side effects of niacin in patients with primary hypercholesterolaemia and mixed dyslipidaemia (see Kamanna *et al.*, 2008). However, in April 2008 the US Food and Drug administration did not approve Merck's application to market *Cordaptive* (see [http://www.merck.com/newsroom/press\\_releases/research\\_and\\_development/2008\\_0428.html](http://www.merck.com/newsroom/press_releases/research_and_development/2008_0428.html)), and at the same time, rejected the name of the combination therapy. The US Food and Drug administration's decision not to approve *Cordaptive*, now renamed *Tredaptive*, is unclear given that the European Medicines Agency has approved this new extended-release combination therapy (see <http://www.emea.europa.eu/humandocs/Humans/EPAR/tredaptive/tredaptive.htm>). One possibility is that the beneficial effects of laropiprant are not superior to aspirin (Kamanna *et al.*, 2008) and the risk/benefit ratio is a primary consideration when the long-term effects of a new drug class are unknown.

DP<sub>1</sub> antagonists may also have utility in the treatment of allergic asthma, although, currently, this is controversial (see

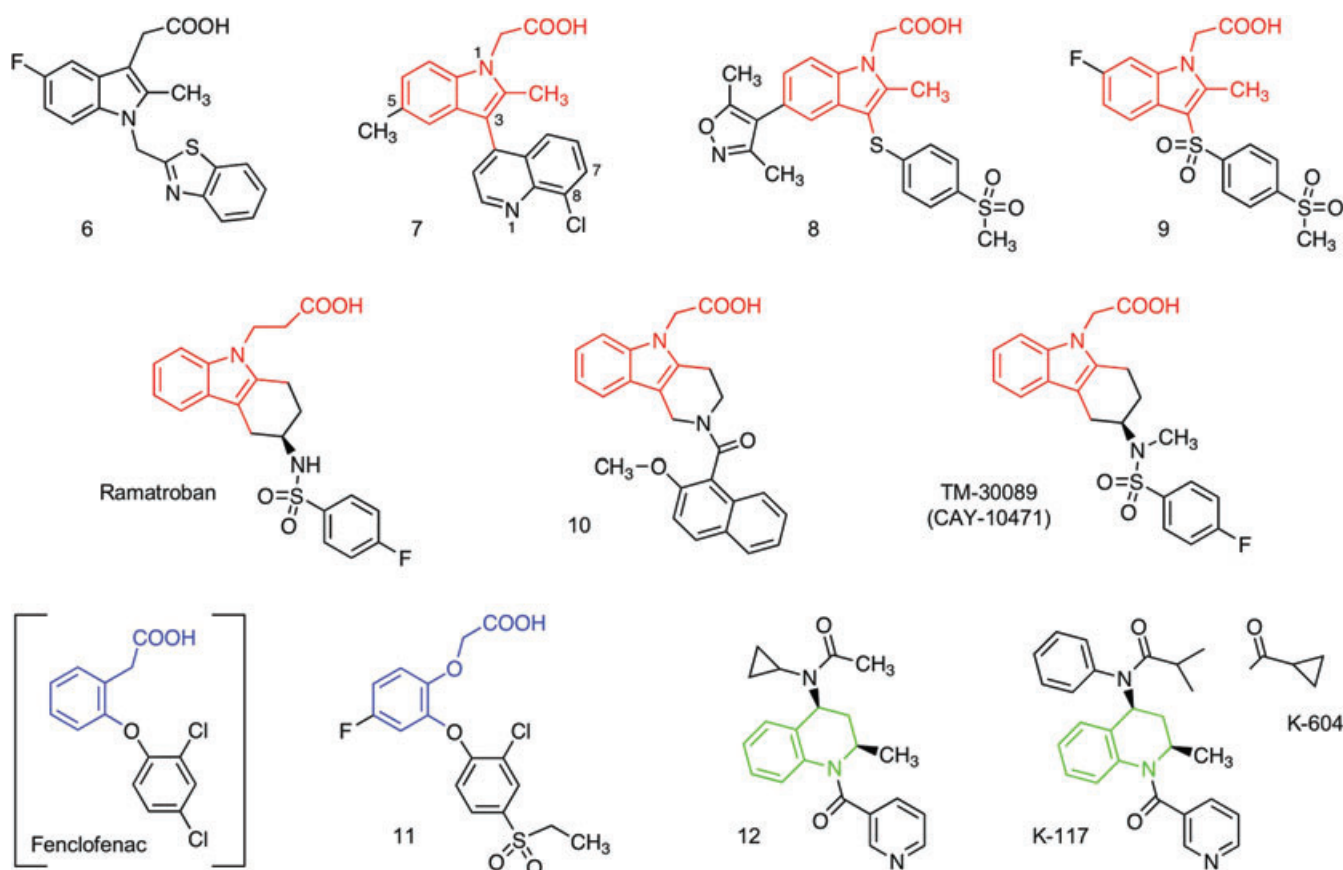
Pettipher, 2008). Persuasive evidence is available that activated mast cells, through their ability to generate PGD<sub>2</sub>, promote T-lymphocyte polarization towards a Th2 phenotype (Faith *et al.*, 2005; Hammad and Lambrecht, 2008). Mechanistically, it is believed that mast cell-derived PGD<sub>2</sub> activates DP<sub>1</sub> receptors on dendritic cells within the respiratory mucosa. This interaction then suppresses the generation of interleukin (IL)-12 (which normally favours a Th1 cell bias) leading to Th2 dominance (Kitawaki *et al.*, 2006; Theiner *et al.*, 2006). A cycle of chronic immunological activation then ensues through further production of IgE and mast cell activation. Indeed, these data are consistent with allergic airway responses being less intense in DP<sub>1</sub> receptor-deficient mice when compared with wild-type animals (Matsuoka *et al.*, 2000). However, the role of DP<sub>1</sub> receptors in regulating allergic reactions is complex. For example, the administration to wild-type mice of the DP<sub>1</sub> agonist, BW-245C, paradoxically *reduced* pulmonary allergic responses whereas DP<sub>1</sub> receptor null mice were unaffected (Hammad *et al.*, 2007). To explain this apparently contradictory result it has been proposed that DP<sub>1</sub> receptor-mediated inhibition of IL-12 release from dendritic cells during host sensitization promotes Th2 polarization (i.e. is pro-inflammatory). In contrast, during maintained airway inflammation, PGD<sub>2</sub> by suppressing dendritic cell function reduces Th2 cell function (i.e. is anti-inflammatory). Indeed, the DP<sub>1</sub> antagonist, S-5751, *attenuated* rather than exacerbated

allergen-induced inflammation in sensitized guinea pigs (Arimura *et al.*, 2001). Thus, the harmful/protective actions of PGD<sub>2</sub> may depend on when and where it is produced (Pettipher, 2008).

## DP<sub>2</sub> receptor antagonists

### Development

Studies conducted in the 1970s showed that PGD<sub>2</sub> elicited peripheral vasoconstrictor responses that were unlikely to be due to activation of, what we now understand to be, DP<sub>1</sub>, FP or TP receptors (Jones, 1976; 1978). In particular, 15-oxo PGD analogues (Jones and Wilson, 1978) had unexpectedly high agonist potency. Moreover, evidence for DP receptor heterogeneity was provided in 1985 by the finding that PGD<sub>2</sub> and six related analogues evoked functional responses across a variety of PGD<sub>2</sub>-sensitive systems that were not mimicked by the selective DP agonist, BW-245C (Narumiya and Toda, 1985). The results of several other studies also indicated the existence of multiple DP receptors (Woodward *et al.*, 1990a; 1993b; Rangachari and Betti, 1993; Fernandes and Crankshaw, 1995; Rangachari *et al.*, 1995). However, it was not until 1999 that the idea of multiple DP-receptors really began to gain general acceptance. Nagata *et al.* (1999b) identified a novel molecule expressed on the surface of minor populations



**Figure 2** DP<sub>2</sub> receptor antagonists. *Inverted* 2-methyl-indole-acetic acid residues (compare with **6**) are highlighted in red; ramatroban has an extra methylene (C2a). The phenylacetic acid moiety is shown in blue in fenclofenac, a lead molecule for compound **11**. Compound **12**, K-117 and K-604 contain a tetrahydroquinoline residue (green).

of CD4<sup>+</sup> T-lymphocytes, which resembled activated Th2 cells in that they released IL-4, IL-5 and/or IL-13 but not interferon- $\gamma$  on stimulation. This orphan site was named 'chemoattractant receptor-homologous molecule expressed on Th2 cells', or CRTh2, as primary sequence analysis showed significant amino acid homology to receptors that mediate chemoattraction (Hirai *et al.*, 2001). Indeed, despite the ability of PGD<sub>2</sub> to interact with the DP<sub>1</sub> subtype and CRTh2 with comparably high affinity ( $K_i$  = 45 and 61 nM respectively), the two cognate receptors are quite dissimilar at the amino acid level (Hirai *et al.*, 2001). A follow-up study by the same investigators discovered a mast cell-derived factor that could increase the cytosolic free Ca<sup>2+</sup> concentration in CRTh2-expressing cells (Nagata *et al.*, 1999a), which was identified as PGD<sub>2</sub> (Hirai *et al.*, 2001). This latter finding coincided with the publication of a pharmacological study in which Monneret *et al.* (2001) found that PGD<sub>2</sub> was chemotactic for eosinophils and also up-regulated the expression of CD11b and L-selectin by a mechanism that was neither mimicked by BW-245C nor blocked by the DP<sub>1</sub> antagonist, BW-A868C. Thus, a second DP receptor subtype (DP<sub>2</sub>, *aka* CD294) was identified on Th2 cells and eosinophils that mediated responses diametrically opposite to the inhibitory effects classically associated with DP<sub>1</sub> agonism. Given that CRTh2 is now known to be ubiquitously expressed within (Nagata *et al.*, 1999a) and outwith (Hirai *et al.*, 2001; Nagata and Hirai, 2003; Kostenis and Ulven, 2006; Kim and Luster, 2007) the immune system (Sawyer *et al.*, 2002; Nagata and Hirai, 2003), the term 'DP<sub>2</sub>' is a more appropriate designation and is used throughout this review.

The ability of PGD<sub>2</sub> to act as a chemoattractant for pro-inflammatory cells and to release Th2-like cytokines has resulted in a concerted effort by the pharmaceutical industry to synthesize selective DP<sub>2</sub> antagonists. Indeed, such compounds may be useful in suppressing a myriad of Th2-driven inflammatory pathologies including asthma, otitis, contact dermatitis and rhinitis. At the time of writing, in excess of 90 patents had been filed claiming selective DP<sub>2</sub> antagonists. In the sections below the main structural classes are described and affinity estimates of lead antagonists (Figure 2) are given in Table 2.

**Indole acetic acids.** In addition to providing a scaffold for the development of DP<sub>1</sub> antagonists, indomethacin (Figure 1) is also a selective, albeit weak, DP<sub>2</sub> agonist (Hirai *et al.*, 2002; Stubbs *et al.*, 2002). Exploiting this property, Pfizer first reported a benzothiazole derivative **6** that had a pA<sub>2</sub> of 6.8 and was ~40-fold selective for the DP<sub>2</sub> receptor (Bauer *et al.*, 2002). Subsequently, several patents describing highly potent and selective DP<sub>2</sub> antagonists were filed by AstraZeneca (Baxter *et al.*, 2003a,b; Birkinshaw *et al.*, 2003; Bonnert *et al.*, 2003; 2004; 2005a,b,c; Bonnert and Rasul, 2004). An initial hit was a 7-chloroquinoline derivative of indomethacin (pA<sub>2</sub> ~7 for human rc-DP<sub>2</sub> receptor), which also inhibited COX-1 with high potency (Birkinshaw *et al.*, 2006). Inversion of the indole template (Figure 2) and substitution of the 5-methoxy moiety by methyl increased antagonist potency by 23-fold and reduced COX-1 inhibition by a factor of 10 (Birkinshaw *et al.*, 2006). Addition of chlorine at position 8 of the quinoline to give **7** also

increased DP<sub>2</sub> affinity by 13-fold (IC<sub>50</sub> for inhibition of [<sup>3</sup>H]-PGD<sub>2</sub> binding = 2.3 nM). This derivative has a preferred biological profile with relatively weak binding to plasma proteins and good bioavailability in rats (76%) and dogs (100%), with half-lives of 1.7 and 5.3 h respectively (Birkinshaw *et al.*, 2006). Compounds having an arylthio substituent on position 3 of the indole nucleus also display very potent antagonism at the DP<sub>2</sub> receptor; compound **8** has a binding IC<sub>50</sub> of 0.4 nM (Bonnert and Rasul, 2004).

Related 1-acetic acid derivatives from Oxagen containing methylene or sulphonyl spacers between the aromatic moieties have been reported as potent DP<sub>2</sub> antagonists (Middlemiss *et al.*, 2005a,b,c,d; Armer *et al.*, 2006; Lovell, 2007), including compound **9**, which has a  $K_i$  of 68 nM and a DP<sub>2</sub>/DP<sub>1</sub>-selectivity ratio of approximately 150. Functionally, **9** potently inhibits DP<sub>2</sub> receptor-mediated human eosinophil shape change and Th2 cell chemotaxis with IC<sub>50</sub> values of 74 and 67 nM respectively (Armer *et al.*, 2005). Moreover, this compound is metabolically stable, has no inhibitory effect on five of the major cytochrome P450 enzymes (1A2, 2C19, 2C9, 2D6, 3A4) and fails to induce CYP3A4, CYP1A and CYP2C9. In rats, **9** is 56% bioavailable and has a half-life of 5.5 h following oral administration. Oxagen has reported the development of a lead compound, ODC9101 (*aka* OC459), which is in Phase IIa clinical trials for asthma. According to the company's website, ODC9101 has completed safety evaluations, is orally active and suitable for once-a-day dosing. Oxagen has also reported the development of a pre-clinical back-up molecule, OC499, and DP<sub>2</sub> antagonists for non-oral delivery (OC1768) and topical administration (OC2125, OC2184; see <http://www.oxagen.co.uk/pdfs/CRTH2summary.pdf>). The structures of these compounds have not been disclosed.

Athersys has also described a series of indole acetic acid derivatives with potent DP<sub>2</sub> antagonist activity (Bennani *et al.*, 2006) including substituted 3-benzylphthalazin-1(2H)-ones which have radioligand binding IC<sub>50</sub> values in the low nanomolar range; their development status is unknown.

**Ramatroban and analogues.** Ramatroban (Bay u3405) was originally described as a TP antagonist with a pA<sub>2</sub> of ~8.8 on human tissues (McKenniff *et al.*, 1991). Later studies revealed DP<sub>2</sub> antagonism (Sugimoto *et al.*, 2003), albeit of lower affinity (pA<sub>2</sub> = 7.44; Mathiesen *et al.*, 2006). The structural similarity of ramatroban to compounds **7–9** is clear. These observations were the impetus for the synthesis of compounds with increased selectivity for the DP<sub>2</sub> receptor (Arimura *et al.*, 2003). An example is **10**, in which the amide nitrogen atom is included in the tricyclic system. This compound has high affinity for the DP<sub>2</sub> receptor (pA<sub>2</sub> = 8.53) with significantly reduced TP receptor-blocking activity (Fretz *et al.*, 2005).

Other minor changes to ramatroban also resulted in compounds with a high degree of DP<sub>2</sub> selectivity. 7TM Pharma reported that *N*-methylating the sulphonamide or truncating the propionate moiety to acetate produces very selective (>1000-fold over DP<sub>1</sub> and TP), high-affinity DP<sub>2</sub> antagonists with  $K_i$  values of 1.9 (TM-30642) and 0.51 nM (TM-30643) respectively (Ulven and Kostenis, 2005). Furthermore, making both modifications to produce TM-30089 (*aka* CAY-10471)

preserved the DP<sub>2</sub> affinity of TM-30643 and further increased the DP<sub>2</sub>/TP-selectivity ratio to >10 000 (Ulven and Kostenis, 2005). Interestingly, in functional studies (e.g. [<sup>35</sup>S]-GTPγS binding/inositol phosphate accumulation; PGD<sub>2</sub>-induced eosinophil shape change), ramatroban and TM-30642 are surmountable competitive antagonists whereas TM 30643 and TM 30089 suppress the maximal response in a concentration-dependent manner (Mathiesen *et al.*, 2006). The insurmountable behaviour of TM-30089 and TM-30643 may be due to its slow dissociation from the DP<sub>2</sub> receptor, which also results in long-lasting antagonism (Mathiesen *et al.*, 2006). Whether such pharmacological behaviour occurs *in vivo* and would be therapeutically advantageous is currently unclear. However, this could be a desirable property as slowly dissociating drugs should act much longer than would be predicted from their plasma half-lives (Mathiesen *et al.*, 2006). 7TM Pharma, in partnership with Ortho-McNeil-Janssen Pharmaceuticals, has a compound in late-stage lead optimization although neither the structure nor the profile of the antagonist has been disclosed (see <http://www.7tm.com/News.aspx?M=News&PID=42&NewsID=39>).

**Phenyl acetic acids.** Another NSAID, fenclofenac (Figure 2), provided the starting point for the synthesis of DP<sub>2</sub> antagonists based on a phenylacetic acid template. An initial hit claimed in the original patent filed by Pfizer (Bauer *et al.*, 2002) had a 4-chlorophenylthio substituent resulting in a functional DP<sub>2</sub>/DP<sub>1</sub>-selectivity ratio of 40. Several other companies, including AstraZeneca, have since filed patents for bis-ether derivatives such as **11**, which has a binding pIC<sub>50</sub> of 9.0 (Bonnert *et al.*, 2005c).

**Tetrahydroquinolines.** Millennium and Warner-Lambert (now Pfizer) were the first to disclose DP<sub>2</sub> antagonists within the tetrahydroquinoline class (Awad *et al.*, 2004; Ghosh *et al.*, 2004; 2005; Kuhn *et al.*, 2004). These compounds are unique in that they are non-acidic indicating that a carboxylic acid moiety is not essential for DP<sub>2</sub> antagonism, as previously assumed (see Pettipher *et al.*, 2007). The 4-amino-tetrahydroquinoline **12** (Figure 2) is reported to gain access to the cerebrospinal fluid after oral dosing and also is efficacious in animal models of inflammation at an oral dose of 25 mg·kg<sup>-1</sup> (Corradini *et al.*, 2005). However, at the time of writing detailed pharmacological data on non-acidic DP<sub>2</sub> antagonists is sparse. Researchers at Kyowa Hakko Kogyo have reported K<sub>i</sub> values of 5.5 and 11 nM for K-117 and K-604 respectively, with minimal interaction with TP or DP<sub>1</sub> receptors at concentrations up to 1 μM (Mimura *et al.*, 2005).

#### Therapeutic applications

Arguably, allergic inflammation is the primary indication for antagonists that selectively block the DP<sub>2</sub> receptor. Indeed, the gene encoding this receptor shows a particularly strong association with asthma in Chinese and African-American populations (Huang *et al.*, 2004). Moreover, there is good evidence from *in vitro* and *in vivo* studies in laboratory animals that PGD<sub>2</sub>, acting via the DP<sub>2</sub> receptor, can mediate many of the cardinal features of allergic airways inflammation (see Ulven and Kostenis, 2006; Pettipher, 2008 for detailed

reviews). The most important observations that have led to this view can be summarized as follows:

1. PGD<sub>2</sub> and selective DP<sub>2</sub> agonists promote chemotaxis of eosinophils, basophils and CD4+ T-lymphocytes of the Th2 subset and this effect is abolished by a neutralizing anti-DP<sub>2</sub> receptor antibody (Hirai *et al.*, 2001; Monneret *et al.*, 2001).
2. PGD<sub>2</sub> promotes pulmonary eosinophilia in rats; this effect is mimicked by selective DP<sub>2</sub>, but not DP<sub>1</sub>, agonists and is abolished by ramatroban (Almishri *et al.*, 2005; Shiraishi *et al.*, 2005).
3. In guinea pigs, the DP<sub>2</sub> agonist, Δ<sup>12</sup>-PGJ<sub>2</sub>, mobilizes eosinophils from the bone marrow (Heinemann *et al.*, 2003).
4. The DP<sub>2</sub> agonist, 13,14-dihydro-15-oxo PGD<sub>2</sub>, promotes pulmonary eosinophilia and exacerbates histopathology in a murine model of allergic asthma (Spik *et al.*, 2005).
5. Ramatroban and DP<sub>2</sub> antagonists devoid of TP receptor-blocking activity reduce pulmonary eosinophilia in several animal species in response to allergen challenge (Nagai *et al.*, 1995; Uller *et al.*, 2007; Pettipher, 2008).
6. PGD<sub>2</sub> promotes the production of Th2 cytokines *in vivo* including IL-4, IL-5 and IL-13 (Fujitani *et al.*, 2002) and *in vitro*, this can occur in the absence of allergen or co-stimulatory molecules (Xue *et al.*, 2005).
7. The expression of the DP<sub>2</sub> receptor on eosinophils is up-regulated in atopic individuals (see Kostenis and Ulven, 2006).
8. High concentrations of PGD<sub>2</sub> are present in the airways of asthmatic subjects after antigen challenge (Murray *et al.*, 1986).
9. The expression of the DP<sub>2</sub> receptor on Th2 T-lymphocytes is up-regulated in individuals sensitized to pollen or house dust mite or who have atopic dermatitis (Iwasaki *et al.*, 2002).

The proof of this line of argument is the marketing of ramatroban in Japan under the trade name *Baynas* for the treatment of perennial allergic rhinitis; its clinical efficacy (e.g. reduction of symptoms and of chronic nasal swelling) has been attributed to DP<sub>2</sub> receptor blockade (Terada *et al.*, 1998).

In addition to anti-allergic indications, DP<sub>2</sub> antagonists may have utility in combating neuropathic pain (Corradini *et al.*, 2005) where the up-regulation of COX-2 and the subsequent formation of PGs are central to disease pathophysiology (Camu *et al.*, 2003).

#### Hybrid DP<sub>1</sub>/DP<sub>2</sub> receptor antagonists

In considering the SAR data described in the preceding sections, indole acetic acids may be a fruitful starting point for antagonists that block both DP<sub>1</sub> and DP<sub>2</sub> receptors. Indeed, in the context of allergic diseases, a hybrid antagonist may exert clinically relevant, beneficial effects that are not achieved when just one DP receptor is targeted. Thus, blockade of the DP<sub>1</sub> subtype would prevent PGD<sub>2</sub> from inhibiting the generation of IL-12 from dendritic cells, thereby inhibiting the polarization of T-lymphocytes to a Th2 phenotype that occurs during host sensitization. Antagonism of the DP<sub>2</sub> receptor

would, at the same time, suppress the chemotactic activity of  $\text{PGD}_2$  towards eosinophils, basophils and T-lymphocytes and so reduce pulmonary leukocyte burden and inflammatory status. Theoretically, this latter action would overcome any  $\text{DP}_1$  receptor-mediated anti-inflammatory effect of  $\text{PGD}_2$  on dendritic cells during maintained inflammation (Pettipher, 2008). It is also tempting to speculate that a non-selective  $\text{DP}$  antagonist that also has TP receptor-blocking activity could be efficacious in allergic asthma. In this scenario, TP receptor blockade would reduce the ability of  $\text{PGD}_2$  to induce bronchoconstriction, which in humans is mediated through the TP receptor (Beasley *et al.*, 1989). Again, indole acetic acids and ramatroban analogues may provide good templates for optimization.

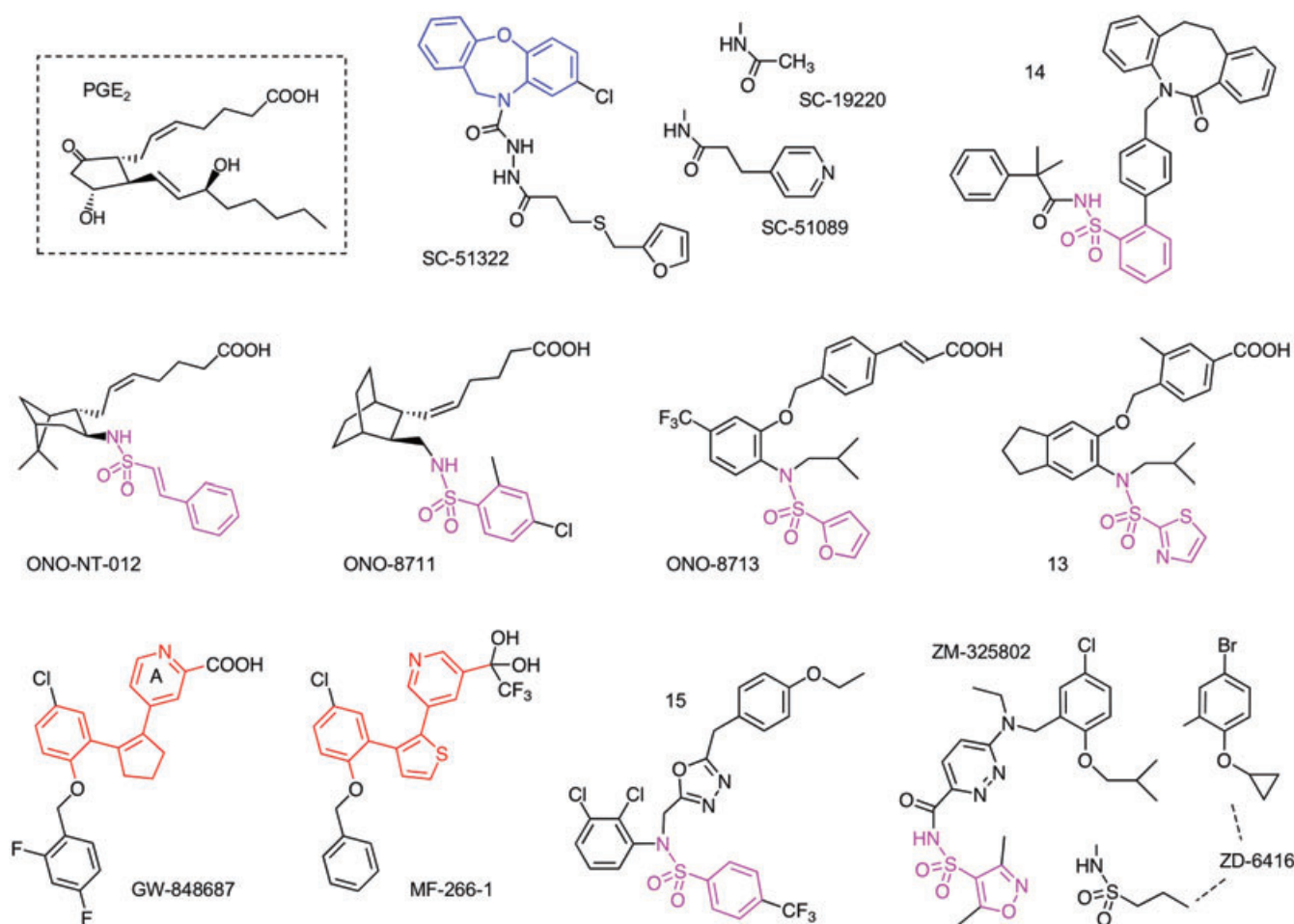
### $\text{EP}_1$ receptor antagonists

#### Development

Figure 3 shows the commonly used  $\text{EP}_1$  antagonists; while some may be considered as prostanoids, none is structurally close to  $\text{PGE}_2$ . The first  $\text{EP}_1$  antagonist was SC-19220, a diben-

zoxazepine hydrazide (Sanner, 1969). Although of low affinity ( $\text{pA}_2 = 5.5$ ), SC-19220 proved useful in the early characterization and elucidation of the roles of EP receptors. For example, on guinea pig trachea, 3–50  $\mu\text{M}$  SC-19220 (and indomethacin) suppressed inherent tone, indicating that  $\text{PGE}_2$  acting via  $\text{EP}_1$  receptors is the likely mediator (Farmer *et al.*, 1974). Also, SC-19220 at 10  $\mu\text{M}$  equally inhibited matching contractions of guinea pig trachea induced by 16,16-dimethyl  $\text{PGE}_2$ , iloprost and isocarbacyclin, but had no effect on contractions induced by U-46619 or histamine; these results demonstrated that certain  $\text{PGI}_2$  analogues could potentially activate  $\text{EP}_1$  receptors (Dong *et al.*, 1986).

SC-19220 (7.5–30  $\text{mg}\cdot\text{kg}^{-1}$  i.p.) inhibited carrageenan-induced inflammation in the rat without suppressing  $\text{PGE}_2$  levels at the injury site (Barbieri *et al.*, 1977). These and other observations were the stimulus for the synthesis of higher-affinity  $\text{EP}_1$  antagonists with potential as anti-inflammatory/analgesic drugs in man; alteration of the acetyl group was the most rewarding strategy. SC-51089 was unusual in the series in releasing hydrazine and was dropped from development (Hallinan *et al.*, 1993). The thioether SC-51322, which is much more potent than the corresponding sulphone (Hallinan *et al.*,



**Figure 3**  $\text{EP}_1$  receptor antagonists. The natural ligand  $\text{PGE}_2$  is shown in the box. The dibenzoxazepine residue in SC-51322 is shown in blue. Aryl-sulphonamido residues in antagonists with prostanoid and non-prostanoid structures are shown in cerise; ONO-NT-012 contains a styryl-sulphonamido moiety. The 1,2-biaryl-cyclopentene pharmacophores in GW-848687 and MF-266-1 are shown in red. Ring A in GW-848687 is part of a picolinic acid (pyridine-2-carboxylic acid) residue.

1994), has become the agent of choice for receptor characterization. It behaved competitively over the range 25–625 nM in a human rc-EP<sub>1</sub> receptor – reporter gene assay ( $pA_2 = 8.8$ ; Schild plot slope = 0.91; Durocher *et al.* (2000). In rat hepatocytes, SC-51322 at 100 nM abolished DNA synthesis induced by PGE<sub>2</sub> or 17-phenyl PGE<sub>2</sub> (Table 1), while 1  $\mu$ M SC-51322 did not affect responses to either PGF<sub>2 $\alpha$</sub>  or PGI<sub>2</sub>; the selective involvement of EP<sub>1</sub> receptors in the PGE response is clear (Kimura *et al.*, 2000; 2001). However, not all inferences about EP<sub>1</sub> receptor involvement are as secure. In a study on rat progenitor Leydig cells, 3 and 30  $\mu$ M SC-51322 suppressed IL-1 $\beta$  expression induced by 10  $\mu$ M 17-phenyl PGE<sub>2</sub> by 28% and 59% respectively (Walch *et al.*, 2003). Given that this system responded well to 100 nM cloprostenol, a potent FP agonist (Table 1), the action of 17-phenyl PGE<sub>2</sub> and its inhibition by SC-51322 may also have involved FP receptors. SC-51322 at 10  $\mu$ M did not inhibit PGE<sub>2</sub>-mediated inhibition of superoxide generation in human blood neutrophils (EP<sub>2</sub> system) (Kanamori *et al.*, 1997) or inhibit sulprostone-induced contraction of rat femoral artery (EP<sub>3</sub> system) at 1  $\mu$ M (Hung *et al.*, 2006).

Another widely used EP<sub>1</sub> antagonist is AH-6809. At 0.1–10  $\mu$ M, it blocked EP<sub>1</sub>-mediated actions of PGE<sub>2</sub>, 16,16-dimethyl PGE<sub>2</sub> or 17-phenyl PGE<sub>2</sub> in an apparently competitive manner ( $pA_2 = 7.4$ ) (Coleman *et al.*, 1987; Eglon and Whiting, 1988; Lawrence *et al.*, 1992). It did not block EP<sub>3</sub> receptor-mediated contraction of guinea pig ileum (Lawrence *et al.*, 1992) and human pulmonary artery (Qian *et al.*, 1994) at 2 and 5  $\mu$ M respectively and had no effect on the pre-synaptic EP<sub>3</sub> action of sulprostone on rat trachea at 3  $\mu$ M (Racké *et al.*, 1992). However, AH-6809 blocks EP<sub>2</sub> receptors (see later) as well as DP<sub>1</sub> and TP receptors in human-washed platelets (Keery and Lumley, 1988) with  $pA_2$  values of  $\sim 6.3$  and  $\sim 5.9$  respectively (our calculation). Of further concern is the inhibitory effect of AH-6809 (3–10  $\mu$ M) on platelet-activating factor (PAF)- and ADP-induced aggregation, which was attributed to inhibition of phosphodiesterase(s) (Keery and Lumley, 1988). In our hands, AH-6809 at 3–10  $\mu$ M caused similar partial block of the contractile actions of phenylephrine ( $\alpha_1$ ), histamine (H<sub>1</sub>), U-46619 (TP) and ONO-AE-248 (EP<sub>3</sub>) on guinea pig aorta (R.L. Jones *et al.*, 2009, submitted). AH-6809 has often been used at even higher concentrations, possibly because of its high water solubility. For example, 30–300  $\mu$ M AH-6809 inhibited the contractile action of PGE<sub>2</sub> in pig large cerebral artery; EP<sub>1</sub> receptor involvement was inferred (Jadhav *et al.*, 2004). However, the  $pA_2$  corresponding to 30  $\mu$ M AH-6809 is only 5.2 (our calculation) and the block was insurmountable at the higher concentrations. With more potent and selective EP<sub>1</sub> antagonists now available, it is time to relegate AH-6809 to its place in the historical development of prostanoid antagonists.

The EP<sub>1</sub> antagonists developed by Ono Pharmaceuticals (Figure 3, middle row) demonstrate an interesting progression from the TP antagonist ONO-11120 (see Figure 8; Katsura *et al.*, 1983) to a related pinane analogue (ONO-NT-012) showing EP<sub>1</sub>, FP and TP antagonism (and EP<sub>3</sub> agonism), to a bicyclo[2.2.2]octane analogue (ONO-8711) showing EP<sub>1</sub>/EP<sub>3</sub> antagonism, and, finally, to the non-prostanoids ONO-8713 and compound **13** with high selectivity for the EP<sub>1</sub> receptor. ONO-8711, ONO-8713 and **13** have  $K_D$  values for mouse rc-EP<sub>1</sub> receptors of 1.7, 0.3 and 0.14 nM respectively (Watanabe *et al.*,

1999; 2000; Naganawa *et al.*, 2006). Small modifications to **13** can restore EP<sub>3</sub> antagonist affinity. In rat-cultured mesangial cells, ONO-8713 at 1  $\mu$ M abolished induction of the transforming growth factor- $\beta$ -fibronectin cascade elicited by PGE<sub>2</sub> under high-glucose conditions; a COX-2-PGE<sub>2</sub>-EP<sub>1</sub> receptor drive was postulated to contribute to deleterious changes in diabetes (Makino *et al.*, 2002). Ohnishi *et al.* (2001) showed that 10  $\mu$ M ONO-8713 partially inhibited PGE<sub>2</sub>-induced exocytosis in mucous cells from guinea pig antrum ( $IC_{50} \sim 1 \mu$ M). However, 17-phenyl PGE<sub>2</sub> was a very weak agonist. The authors postulated that EP<sub>1</sub> and EP<sub>4</sub> receptors co-operate to sustain the high exocytotic response to PGE<sub>2</sub>. Norel *et al.* (2004) also showed that 10  $\mu$ M ONO-8713 partially suppressed the contractile action of sulprostone on human pulmonary vein; functional EP<sub>1</sub> receptors were postulated, even though the selective EP<sub>1</sub> agonist ONO-DI-004 was a very weak agonist. Prostanoid receptor binding data alone do not guarantee functional selectivity at these high antagonist concentrations; inclusion of control agonists (both prostanoid and non-prostanoid) in the system under test is essential.

Other pharmacophores for potent EP<sub>1</sub> antagonism, with inactivity against cytochrome P450 enzymes and good penetration into the central nervous system being secondary goals (see later), have emerged within the last 10 years. The series reported by Merck (Ruel *et al.*, 1999) contains a tricyclic system akin to that in the Searle series. Again EP<sub>1</sub>/EP<sub>3</sub> selectivity can be readily modulated, as shown by replacement of the terminal phenyl group in compound **14** ( $K_i$  values for human EP<sub>1</sub> and EP<sub>3</sub> receptors = 10 and 4000 nM) by methyl (770 and 1000 nM). EP<sub>1</sub> antagonists with aryl groups attached to adjacent carbons of a 5-membered ring (cyclopentene, thiophene, pyrrole) have been reported by research groups at GlaxoSmithKline and Merck Frosst. GW-848687 has nanomolar affinity for the human EP<sub>1</sub> receptor, 30-fold lower affinity for the human TP receptor and >400 times lower affinity for other prostanoid receptors (Giblin *et al.*, 2007). The picolinic acid residue in GW-848687 (ring A) is highly acidic ( $pK_a \sim 1.0$ ) and there has been considerable SAR work by both research groups on modulating the acidity of this region. The Merck antagonist MF-266-1, with a *m*-C(CF<sub>3</sub>)-(OH)<sub>2</sub> substituent (Figure 3), is a weak acid ( $pK_a = 7.5$  for first ionization) that retains high EP<sub>1</sub> affinity (Ducharme *et al.*, 2005; Clark *et al.*, 2008). A *m*-C(CF<sub>3</sub>)<sub>2</sub>-OH substituent in the Merck series and a *p*-C(CF<sub>3</sub>)<sub>2</sub>-OH substituent in the GlaxoSmithKline series resulted in lower EP<sub>1</sub> affinity (Ducharme *et al.*, 2005; Hall *et al.*, 2007a). Further modifications to ring A (e.g. *m*-NH(C=O)CH<sub>2</sub>Ph/*p*-(C=O)NHCH(CH<sub>3</sub>)Ph substituents) resulted in non-acidic relatives with high EP<sub>1</sub> potency. Compound **15** is also a non-acidic EP<sub>1</sub> antagonist (Hall *et al.*, 2007b).

#### Therapeutic applications

The upper portion of Table 3 shows that parenteral administration of EP<sub>1</sub> antagonists of different chemical classes suppressed the allodynic/hyperalgesic signs of inflammation in the rat and mouse. Intraplantar injection of these antagonists also opposed pain-producing stimuli, although at the mouse EP<sub>1</sub> receptor AH-6809 had minimal affinity, which suggests an off-target effect. That said, Khasar *et al.* (1993) showed that PGE<sub>2</sub> and SC-19220 are only mutually antagonistic when they

Table 3 Anti-nociceptive and anti-inflammatory activities of EP receptor antagonists

Route/species	Experimental model	Noxious stimulus/site	Antagonist (nominal receptor)	Antagonist dosing	Efficacy	Reference
Oral Guinea pig	joint pain (mono-osteoarthritis)	Iodoacetate/shoulder joint	MF-498 (EP <sub>4</sub> )	30 mg·kg <sup>-1</sup>	+++	Clark <i>et al.</i> (2008)
	Thermal hyperalgesia	L-902688 <sup>8</sup> /paw	MF-498 (EP <sub>4</sub> )	0.1–30 mg·kg <sup>-1</sup>	+++	Clark <i>et al.</i> (2008)
	Writhing response	Acetic acid/peritoneal cavity	SC-19220 (EP <sub>1</sub> )	50–300 mg·kg <sup>-1</sup>	++	Drower <i>et al.</i> (1987)
	Behavioural response	Formalin/paw	SC-19220 (EP <sub>1</sub> )	50–150 mg·kg <sup>-1</sup>	++	Drower <i>et al.</i> (1987)
	Chronic mechanical/thermal hyperalgesia/allodynia	Nerve constriction/sciatic	ONO-8711 (EP <sub>1</sub> )	10–100 mg·kg <sup>-1</sup> per day	+++	Kawahara <i>et al.</i> (2001)
	Chronic hyperalgesia (weight-bearing)	Freund's adjuvant/knee joint	GW-848687 (EP <sub>1</sub> )	30 mg·kg <sup>-1</sup> per day	+++	Giblin <i>et al.</i> (2007)
	Chronic arthritis (oedema/radiology)	Freund's adjuvant/paw	Compound <b>20</b> (EP <sub>4</sub> )	0.005 mg·kg <sup>-1</sup> per day	+++	Burch <i>et al.</i> (2008)
	Local oedema	Freund's adjuvant/paw	MF-266-1 (EP <sub>1</sub> )	Each 0.008–20 mg·kg <sup>-1</sup> daily for 10 days	0	Clark <i>et al.</i> (2008)
			MF-266-3 (EP <sub>3</sub> )		0	
			MF-498 (EP <sub>4</sub> )		+++	
Mouse	Mechanical hyperalgesia	Carrageenan/paw	CJ-042794 (EP <sub>4</sub> )	1–30 mg·kg <sup>-1</sup>	+++	Murase <i>et al.</i> (2008a)
	Local oedema	Freund's adjuvant/paw	CJ-042794 (EP <sub>4</sub> )	1–30 mg·kg <sup>-1</sup>	+++	Murase <i>et al.</i> (2008a)
	Mechanical hyperalgesia	Carrageenan/paw	CJ-023423 (EP <sub>4</sub> )	3–100 mg·kg <sup>-1</sup>	+++	Nakao <i>et al.</i> (2007)
	Thermal hyperalgesia	PGE <sub>2</sub> /paw	CJ-023423 (EP <sub>4</sub> )	1–29 mg·kg <sup>-1</sup>	+++	Nakao <i>et al.</i> (2007)
	Chronic inflammation (weight-bearing)	Freund's adjuvant/paw	CJ-023423 (EP <sub>4</sub> )	10–57 mg·kg <sup>-1</sup>	+++	Nakao <i>et al.</i> (2007)
	Chronic inflammation (oedema, histology)	Freund's adjuvant/paw	CJ-023423 (EP <sub>4</sub> )	29–96 mg·kg <sup>-1</sup>	+++	Okumura <i>et al.</i> (2008)
	Writhing response	Phenylbenzylquinone/peritoneal cavity	SC-51089 (EP <sub>1</sub> )	1–30 mg·kg <sup>-1</sup>	+++	Hallinan <i>et al.</i> (1993)
	Writhing response	Phenylbenzylquinone/peritoneal cavity	SC-51322 (EP <sub>1</sub> )	1–30 mg·kg <sup>-1</sup>	+++	Hallinan <i>et al.</i> (1994)
	Migration of Langerhans cells into lymph node	Fluorescein isothiocyanate/skin	ONO-AE3-208 (EP <sub>4</sub> )	10 mg·kg <sup>-1</sup> twice daily	++	Kabashima <i>et al.</i> (2003)
	Ear swelling/histology	UVB/skin of ear	ONO-AE3-208 (EP <sub>4</sub> )	10 mg·kg <sup>-1</sup>	++	Kabashima <i>et al.</i> (2007)
Intravenous Rat	Sensory discharge	Acetic acid/bladder lumen	ONO-8711 (EP <sub>1</sub> )	1–3 mg·kg <sup>-1</sup>	++	Ikeda <i>et al.</i> (2006)
	Visceromotor reflex	Distension/bladder	DG-041 (EP <sub>3</sub> )	10 mg·kg <sup>-1</sup>	+++	Su <i>et al.</i> (2008a)
Intraperitoneal Rat	Mechanical hyperalgesia	Freund's adjuvant/paw	AH-23848 (EP <sub>4</sub> )	0.1–10 mg·kg <sup>-1</sup>	+++	Lin <i>et al.</i> (2006)
	Thermal hyperalgesia	Freund's adjuvant/paw	AH-23848 (EP <sub>4</sub> )	0.1–10 mg·kg <sup>-1</sup>	+++	Lin <i>et al.</i> (2006)
	Mechanical allodynia	Sulprostone/intrathecal	ZM-325802 (EP <sub>1</sub> )	0.03 µg·kg <sup>-1</sup>	+++	Gil <i>et al.</i> (2008)
Topical to skin Mouse	Oedema	Chronic UVB	ONO-8713 (EP <sub>1</sub> )	96 nmol	++	Tober <i>et al.</i> (2006)
	Intraplantar					

Rat	Paw-withdrawal Mechanical hyperalgesia Chronic mechanical hyperalgesia Chronic thermal hyperalgesia	PGE <sub>2</sub> /paw <sup>b</sup> Incision wound/paw Partial nerve transection/ sciatic Partial nerve transection/ sciatic	SC-19220 (EP <sub>1</sub> ) ONO-8711 (EP <sub>1</sub> ) SC-19220 (EP <sub>1</sub> ) SC-19220 (EP <sub>1</sub> )	2.3 nmol 4.5–114 nmol 2.3 nmol 2.3 nmol	++ +++ +++ ++	Khassar <i>et al.</i> (1993) Omote <i>et al.</i> (2001) Syriatowicz <i>et al.</i> (1999) Syriatowicz <i>et al.</i> (1999) Kassuya <i>et al.</i> (2007)
Mouse	Mechanical allodynia  Paw-licking	PGE <sub>2</sub> /paw Carrageenan/paw PGE <sub>2</sub> /paw	AH-6809 (EP <sub>1</sub> ) L-826266 (EP <sub>3</sub> ) L-161982 (EP <sub>4</sub> ) L-826266 (EP <sub>3</sub> ) L-161982 (EP <sub>4</sub> ) AH-6809 (EP <sub>1</sub> ) L-826266 (EP <sub>3</sub> ) L-161982 (EP <sub>4</sub> )	100 nmol 0.1–10 nmol 10 nmol 10 nmol 10 nmol 10–100 nmol 3–30 nmol 1–30 nmol	0 +++ 0 ++ ++ 0 ++ +++	Kassuya <i>et al.</i> (2007) Kassuya <i>et al.</i> (2007) Kassuya <i>et al.</i> (2007)
Intrathecal Rat	Mechanical allodynia  Mechanical hyperalgesia Thermal hyperalgesia Flinching (late phase) Mechanical hyperalgesia (late phase) Mechanical hyperalgesia (late phase) Visceromotor reflex	Nerve ligation/LS spinal root Incision wound/foot Incision wound/paw Formalin/paw Carrageenan/paw PGE <sub>2</sub> /intrathecal Distension/bladder	SC-51322 (EP <sub>1</sub> )  ONO-8711 (EP <sub>1</sub> ) ONO-8711 (EP <sub>1</sub> ) SC-58109 (EP <sub>1</sub> ) ONO-8711 (EP <sub>1</sub> ) ONO-8711 (EP <sub>1</sub> ) L-798106 (EP <sub>3</sub> ) DG-041 (EP <sub>3</sub> )	22–220 nmol  114–228 nmol 114–128 nmol 65–650 nmol 2.28–228 nmol 0.11–0.45 nmol 10–300 nmol 10–100 nmol	++  ++ 0 ++ ++ ++ ++ ++ ++	Hefferan <i>et al.</i> (2003); O'Reilly and Loomis (2007) Omote <i>et al.</i> (2002) Ikeda <i>et al.</i> (2006) Malmberg <i>et al.</i> (1994) Nakayama <i>et al.</i> (2002) Nakayama <i>et al.</i> (2004) Su <i>et al.</i> (2008b) Su <i>et al.</i> (2008b)
Mouse	Thermal hyperalgesia Writhing response Mechanical allodynia Thermal hyperalgesia Mechanical allodynia Thermal hyperalgesia	PGE <sub>2</sub> /intrathecal PGE <sub>2</sub> /intrathecal PGE <sub>2</sub> /intrathecal PGE <sub>2</sub> /intrathecal <sup>c</sup> PGE <sub>2</sub> /intrathecal PGE <sub>2</sub> /intrathecal	AH-6809 (EP <sub>1</sub> ) AH-6809 (EP <sub>1</sub> ) AH-6809 (EP <sub>1</sub> ) AH-6809 (EP <sub>1</sub> ) ONO-NT-012 (?) <sup>d</sup> ONO-NT-012 (?)	0.29–2.9 nmol 0.29–2.9 nmol 4.2 nmol 4.2 nmol 3 × 10 <sup>-5</sup> –0.03 nmol 3 × 10 <sup>-5</sup> –0.03 nmol	+++ +++ 0 +++ +++ 0	Uda <i>et al.</i> (1990) Uda <i>et al.</i> (1990) Minami <i>et al.</i> (1995) Minami <i>et al.</i> (1995) Minami <i>et al.</i> (1995) Minami <i>et al.</i> (1995)
Topical to spinal tissue Rat	Mechanical allodynia	Bicuculline/LS–L6 spine	SC-51322 (EP <sub>1</sub> )	0.22–17.5 nmol	+++	Zhang <i>et al.</i> (2001)

Upper and lower panels relate to parenteral and topical administration of antagonist.

<sup>a</sup>Selective EP<sub>4</sub> agonist.

<sup>b</sup>Intradermal.

<sup>c</sup>About 30-fold higher dose of PGE<sub>2</sub> required compared with mechanical allodynia.

<sup>d</sup>See EP<sub>1</sub> antagonist section for specificity; reported pA<sub>2</sub> of 9.96 in allodynia model is unlikely to be valid because antagonist concentrations in biophase are unknown.



are both injected intradermally and not subcutaneously. More recently, attention has focused on the role of EP receptors in central nociception (see Svensson and Yaksh, 2002; Hefferan *et al.*, 2003; Mebane *et al.*, 2003). Minami *et al.* (2001) showed that PGE<sub>2</sub> could induce hyperalgesia in the mouse when injected intrathecally over the dose-range 0.00035–350 pmol. Deletion of the EP<sub>3</sub> receptor gene removed the more sensitive component of the hyperalgesia. In addition, the selective EP<sub>3</sub> agonist ONO-AE-248 induced hyperalgesia at relatively low doses. Surprisingly, the EP<sub>1</sub> knock-out mouse showed a hyperalgesic response in the hot-plate test, thereby confounding the role of EP<sub>1</sub> receptors in mediating hyperalgesia (Minami *et al.*, 2001). EP<sub>1</sub> antagonists given intrathecally suppress flinching, and mechanical allodynia and hyperalgesia (Table 3), although in certain cases the doses required for these effects are high. For example, in the studies by Omote *et al.* (2002), 100 µg ONO-8711 was injected intrathecally in a volume of 10 µl, representing an injectate concentration of 23 mM! Even after dilution in the cerebrospinal fluid, EP<sub>3</sub> receptors are likely to be blocked as well.

Intravenous ONO-8711 also suppressed afferent nerve discharge to distension of the rat bladder sensitized with acetic acid (Ikeda *et al.*, 2006). The inference from these studies was that PG(E<sub>2</sub>) generated locally activates EP<sub>1</sub> receptors on peripheral sensory neurones. Again EP<sub>1</sub>/EP<sub>3</sub> selectivity is critical, given that EP<sub>3</sub> agonists augment bradykinin-induced sensory nerve discharge (Kumazawa *et al.*, 1996). Topical ONO-8713 (like celecoxib) inhibited UV-B-induced skin inflammation and tumour development in the mouse (Tober *et al.*, 2006); the higher selectivity of ONO-8713 supports EP<sub>1</sub> receptor involvement. PGE<sub>2</sub> production, but not COX-1/COX-2 expression, was suppressed by ONO-8713; the mechanism is not clear.

Despite continuing synthesis of potent EP<sub>1</sub> antagonists with testing in analgesic/anti-inflammatory models, there has been little solid evidence of clinical efficacy. Sarkar *et al.* (2003) reported that ZD-6416, which is related to ZM-325802 (Figure 3; Shaw *et al.*, 1999; Jenkins *et al.*, 2001), inhibited upper oesophageal pain threshold to electrical stimulation in human volunteers. However, ZD-6416 does not appear to be particularly useful in the clinical setting (development profile obtained from Pharmaprojects; <http://www.pharmaprojects.com>).

While the preferred indication for EP<sub>1</sub> antagonists has been for pain, additional therapeutic uses in cancer, osteoporosis, arthritis, and neurodegenerative and renal disorders have been suggested. COX-2 inhibitors are of potential value in reducing colorectal adenomas which, in turn, has created interest in using prostanoid antagonists as an alternative. The cardiovascular risk associated with celecoxib in clinical trials involving colorectal adenoma prevention (Solomon *et al.*, 2005) would intensify interest in using prostanoid antagonists for this indication. ONO-8711 inhibited formation of colonic crypts (Kawamori *et al.*, 2001) and reduced the frequency of polyp formation in APC1309 mice (Watanabe *et al.*, 1999; Kitamura *et al.*, 2003b), and aberrant crypt foci in oxymetazoline-treated mice (Watanabe *et al.*, 1999). Correspondingly, aberrant crypt foci were reduced by 60% in EP<sub>1</sub><sup>-/-</sup> receptor mice (Watanabe *et al.*, 1999). COX-2 up-regulation has also been considered a target for drug treatment of

pathologies involving neurological injury and neurodegeneration. The downstream effects of COX-2 neurotoxicity have been reported to be EP<sub>1</sub> receptor-mediated (Kawano *et al.*, 2006).

Blood pressure in the spontaneously hypertensive rat was reduced by SC-51322 (10 mg<sup>-1</sup>·kg<sup>-1</sup>·day<sup>-1</sup>, gavage) (Guan *et al.*, 2007). EP<sub>1</sub> receptors appear to regulate BP in the male but not the female mouse (Audoly *et al.*, 1999; Stock *et al.*, 2001). In a more detailed study, Guan *et al.* (2007) showed that SC-51322 and EP<sub>1</sub>-receptor gene deletion blunted pressor responses to 17-phenyl PGE<sub>2</sub> and sulphostone, whereas the pressor response to the PGE<sub>1</sub> analogue MB-28767 (claimed to be 'a pure EP<sub>3</sub> agonist') was the same in EP<sub>1</sub><sup>+/+</sup> and EP<sub>1</sub><sup>-/-</sup> mice. While we agree with the authors' contention that both EP<sub>1</sub> and EP<sub>3</sub> receptors contribute to the pressor effects, the utility of MB-28767 may be compromised by its moderate TP agonism (Lawrence and Jones, 1992). EP<sub>1</sub>-receptor gene deletion also reduced the elevated BP and cardiac hypertrophy following a 4-week infusion of angiotensin (Ang) II in the mouse, and 1 µM SC-51322 markedly attenuated the contractile action of Ang II on the isolated pre-glomerular arteriole (Guan *et al.*, 2007). How these findings for EP<sub>1</sub> receptors integrate with the modest hypertension and reduction of antihypertensive efficacy associated with NSAIDs and COX-2 inhibitors in the human setting (Johnson *et al.*, 1994; Ishiguro *et al.*, 2008) is not clear. Species and gender appear to be highly influential factors.

## EP<sub>2</sub> receptor antagonists

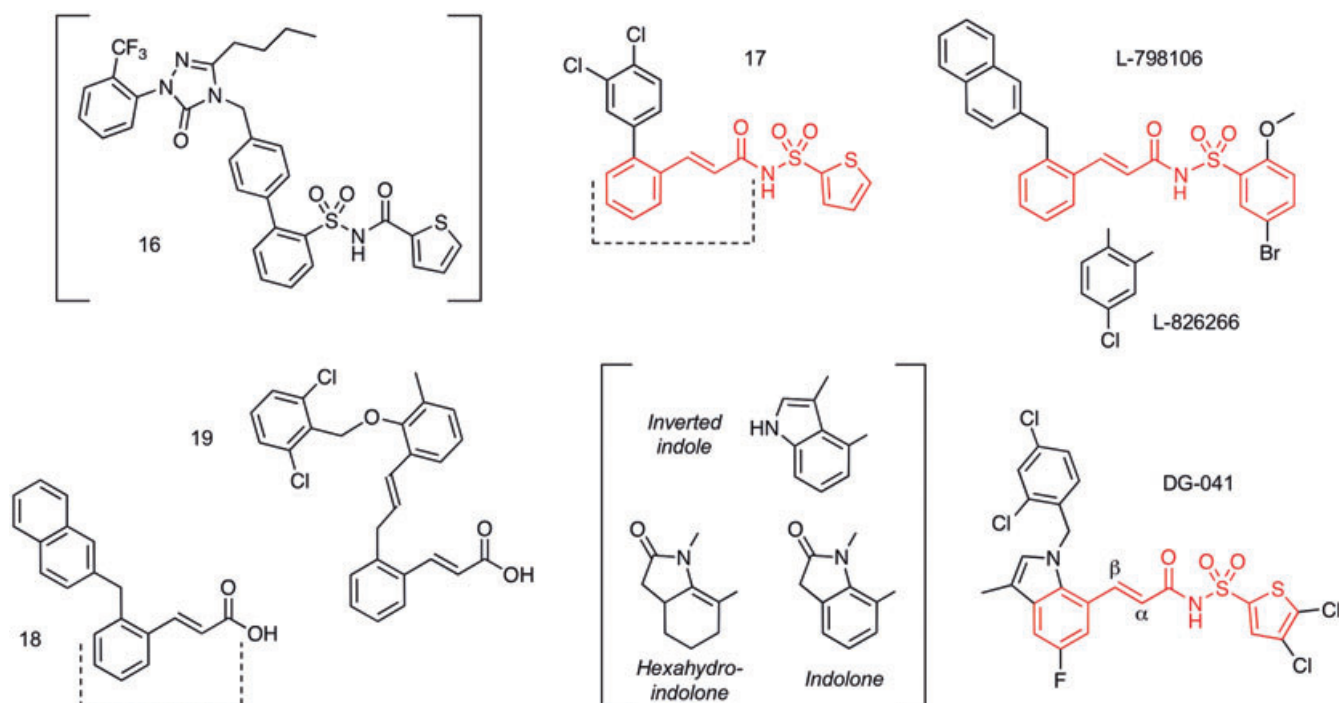
Selective EP<sub>2</sub> receptor antagonists are essentially unavailable. This may be partly due to a reluctance to inhibit the potential anti-inflammatory actions of endogenous PGE<sub>2</sub> (see Teixeira *et al.*, 1997) mediated via EP<sub>2</sub> receptors (Noguchi *et al.*, 1999; Nataraj *et al.*, 2001).

Woodward *et al.* (1995b) showed that AH-6809 has modest affinity for human rc-EP<sub>2</sub> receptors and antagonized PGE<sub>2</sub>-induced activation of adenylyl cyclase with a pA<sub>2</sub> of about 6.5. Lower affinities were found for inhibition of PGE<sub>2</sub>-induced relaxation of human bronchus (pA<sub>2</sub> = 5.78; Norel *et al.*, 1999) and guinea pig trachea (pA<sub>2</sub> = 5.6, KJ Ong and RL Jones, unpubl. obs.) (Table 2). This is consistent with its utility for distinguishing EP<sub>1</sub> receptors from other EP subtypes in non-primate pharmacological preparations (Coleman *et al.*, 1987; Eglen and Whiting, 1988; Lawrence *et al.* 1992). However, given the poor selectivity of AH-6809 in the low micromolar range, findings arising from its use in high concentration as an EP<sub>2</sub> antagonist (e.g. 100 µM in Aronoff *et al.*, 2004) should be approached with caution. K<sub>i</sub> values for rc-EP<sub>2</sub> receptors of about 1 µM have been found for some compounds in combinatorial studies (see Murase *et al.*, 2008b); it should be possible to build on these observations.

## EP<sub>3</sub> receptor antagonists

### Development

EP<sub>3</sub> antagonism in a series of biaryl-acylsulphonamides was reported in 2002 by Merck (Gallant *et al.*, 2002). The lead



**Figure 4** EP<sub>3</sub> receptor antagonists. The AT<sub>1</sub> receptor antagonist, compound **16**, is a lead molecule for the biaryl-ene-acyl-sulphonamide antagonists (pharmacophore in red). The left-hand portion of this pharmacophore corresponds to the cinnamic acid moiety in compounds **18** and **19** (see broken brackets). L-826266 is a chloro analogue of L-798106. The lower-middle brackets show modifications to the indole nucleus in the EP<sub>3</sub> antagonist series of DeCode Genetics.

compound was an AT<sub>1</sub> receptor antagonist **16** (Figure 4) with a binding  $K_D$  of 7  $\mu$ M for the human EP<sub>3</sub> receptor. In a combinatorial approach, reversal of the acylsulphonamide and insertion of an ethylene spacer yielded **17** with an EP<sub>3</sub>  $K_D$  of 25 nM and minimal binding to other prostanoid receptors. A related analogue, L-798106, at 0.2  $\mu$ M blocked the pre-synaptic (EP<sub>3</sub>) inhibitory actions of sulprostone on guinea pig vas deferens and trachea with  $pA_2$  values of 7.5 and 7.8 respectively (Clarke *et al.*, 2004). L-798106 (0.2–1  $\mu$ M) also caused parallel displacement of log concentration–response curves for contraction of rat femoral artery by sulprostone (in synergy with phenylephrine) (Hung *et al.*, 2006). L-826266, a chloro analogue of L-798106, has been used in other studies despite the lack of detailed information on its basic pharmacology. Schlemper *et al.* (2005) showed that L-826266 at 30  $\mu$ M inhibited both PGE<sub>2</sub>- and bradykinin-induced relaxation of guinea pig trachea, and proposed that bradykinin induces *de novo* synthesis of PGE<sub>2</sub>, which then activates EP<sub>3</sub> receptors. However, the specificity of L-826266 at this high concentration is unknown. An alternative explanation is that L-826266 modestly blocks EP<sub>2</sub> receptors thereby allowing the action of PGE<sub>2</sub> on the contractile EP<sub>1</sub> system in the trachea to dominate. Oliva *et al.* (2006) reported that L-826266 injected (as 0.1  $\mu$ L of 1.25–5 mM solutions in 20% DMSO/PSS) into the periaqueductal grey matter of the mouse brain suppressed the late hyperalgesic response to intradermal formalin. However, the concentration of L-826266 at the site(s) of action is indeterminate and interpretation of the finding is difficult because similar high ‘doses’ of EP<sub>1</sub> and EP<sub>4</sub> antagonists and AH-6809 also suppressed the formalin response.

DeCode Genetics has described a series of related molecules containing an indole nucleus, from which DG-041 (Figure 4) was selected for clinical investigation (Singh *et al.*, 2009). DG-041 had a  $IC_{50}$  of 8.1 nM in a EP<sub>3</sub>/Ca<sup>2+</sup> flux FLIPR assay; corresponding values in DP<sub>1</sub> and DP<sub>2</sub> assays were 131 and >10 000 nM respectively (Singh *et al.*, 2009). SAR studies involved modifications to the terminal aryl moieties, together with the indole unit. Compounds with an inverted indole nucleus retain high EP<sub>3</sub> affinity (Zhou *et al.*, 2009a,b), as do indolones and hexahydro-indolones (O’Connell *et al.*, 2009). Saturation of the  $\alpha,\beta$ -double bond also produced highly potent EP<sub>3</sub> antagonists, while further saturation of the remaining double bond in the hexahydro-indolone residue resulted in marked reduction in affinity. Much of DeCode’s work was directed towards improving water solubility: predicted *n*-octanol/water partition coefficients (ClogP) for DG-041, L-798106 and L-826266 are 6.6, 6.9 and 7.4 respectively (ChemAxon freeware). In this context, our recent studies on L-798106 and L-826266 have shown a slowly developing block of EP<sub>3</sub> agonist-induced contraction on guinea pig aorta, affording  $pA_2$  values of 7.96 and 7.58 respectively after 3-h contact (Jones *et al.*, 2008). As expected, highly potent antagonists such as BMY-180291 (TP,  $pA_2$  = 9.8) and doxepin (histamine H<sub>1</sub>,  $pA_2$  = 9.6) also had slow onsets at low-nanomolar concentrations. However, the slow onsets of L-798106 and L-826266 may be related to their high lipophilicity rather than their (moderate) receptor affinity.

Merck-Frosst researchers have also identified EP<sub>3</sub> antagonism in *ortho*-substituted cinnamic acid derivatives, which correspond to the left-hand portions of the ene-acyl-

sulphonamide antagonists shown in Figure 4. Compound **18** has a binding  $K_D$  for the human EP<sub>3</sub> receptor of 20 nM (Juteau *et al.*, 2001), while compound **19** has the highest binding affinity (3 nM) and behaved as a pure antagonist in a human EP<sub>3</sub>/adenylyl cyclase assay ( $pA_2 = 8.22$ ; Belley *et al.*, 2005). Compound **19** is also highly lipophilic (ClogP = 8.29).

ONO-AE3-240 is reported to be a highly selective EP<sub>3</sub> antagonist (mouse EP<sub>3</sub>/EP<sub>1</sub> selectivity ratio = 2500; Amano *et al.*, 2003), but its structure has not been disclosed.

#### Therapeutic applications

Like most prostanoid receptors, the EP<sub>3</sub> receptor has been implicated in pain of various aetiologies. These include allodynia produced by HIV-1 glycoprotein gp 120 (Minami *et al.*, 2003) and PGE<sub>2</sub> (Kassuya *et al.*, 2007), acute herpetic pain (Takasaki *et al.*, 2005), thermal hyperalgesia (Oka *et al.*, 1994) and formalin-induced hyperalgesia (Oliva *et al.*, 2006). A major role for EP<sub>3</sub> (and IP) receptors has been claimed in endotoxin-induced enhancement of pain perception (Ueno *et al.*, 2001). Intravenous DG-041 also suppressed the visceromotor reflex to bladder distension in the rat (Su *et al.*, 2008a); the authors defined this antagonist as brain non-penetrant, but no pharmacokinetic evidence was presented. Intrathecal application of L-798106 and DG-041 also produced a long-lasting suppression of the visceromotor reflex, while intracerebroventricular administration produced only a transient reduction (Su *et al.*, 2008b). However, EP<sub>3</sub> agonists are known to exert gastrointestinal cytoprotection and attenuate gastric acid secretion in animal models (Bunce *et al.*, 1990; Kunikata *et al.*, 2002). Although it is widely assumed that there is a similar involvement of EP<sub>3</sub> receptors in man, an extensive search of the literature provides only circumstantial evidence for this. For example, while misoprostol is clinically useful in suppressing gastroduodenal erosion (see Hawkey, 2000), its selectivity is not high enough to infer the involvement of EP<sub>3</sub> receptors (EP<sub>3</sub> ~ EP<sub>2</sub> ~ EP<sub>4</sub> >> EP<sub>1</sub> for misoprostol-free acid; Abramovitz *et al.*, 2000). A reduction in myocardial ischaemic damage was also achieved with EP<sub>3</sub> agonist treatment in the rat (Zacharowski *et al.*, 1999). Thus, it is feasible that the therapeutic use of EP<sub>3</sub> antagonists will result in a side effect profile at least comparable to that of COX inhibitors. This possibility does not seem to have hindered development work on other uses of EP<sub>3</sub> antagonists as discussed below.

It has been suggested that endogenous PGE<sub>2</sub> activates EP<sub>3</sub> receptors on stromal cells surrounding a tumour causing the release of vascular endothelial growth factor, which then promotes angiogenesis and tumour growth (Amano *et al.*, 2003). Injection of the EP<sub>3</sub> antagonist ONO-AE3-240 around sarcoma-180 tumours in the mouse markedly suppressed these effects in a manner similar to EP<sub>3</sub> receptor gene-deletion; ONO-8711 (EP<sub>1</sub> antagonist) and ONO-AE3-208 (EP<sub>4</sub> antagonist) were ineffective. In contrast, EP<sub>3</sub> agonists acting on all three mouse EP<sub>3</sub> receptor isoforms expressed in HEK-293 cells caused cell clustering and inhibited their proliferation via a G<sub>12</sub>-RhoA pathway (Macias-Perez *et al.*, 2008).

A further possible therapeutic application of an EP<sub>3</sub> antagonist is the treatment of pre-term labour. Based on the known activity of misoprostol (Sanchez-Ramos *et al.*, 1997) and sul-

prostone (Fruzzetti *et al.*, 1988), it appears that EP<sub>3</sub> receptor stimulation produces cervical ripening, a critical event that precedes parturition. An EP<sub>3</sub> antagonist could be effectively combined with a tocolytic, such as an EP<sub>2</sub> agonist (Senior *et al.*, 1993), to provide therapy for pre-term labour. The EP<sub>3</sub> receptor has been uniquely associated with febrile responses (Ushikubi *et al.*, 1998). However, development of an EP<sub>3</sub> antagonist for treating fever seems unlikely, given that low-cost COX inhibitors are highly effective in reducing body temperature.

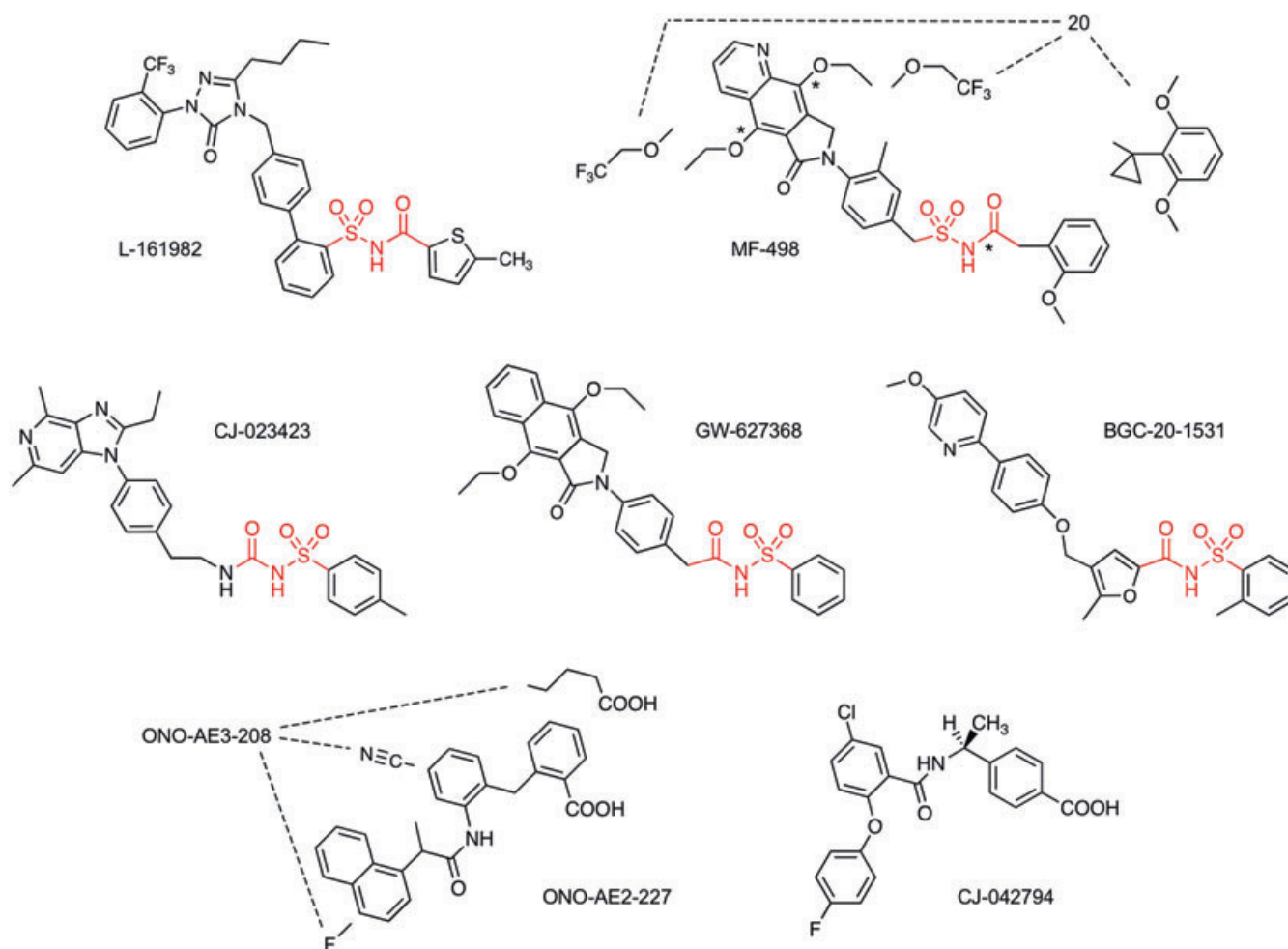
Finally, DG-041 has shown promise in the treatment of peripheral cardiovascular disease. Activation of EP<sub>3</sub> receptors on human platelets enhances aggregation induced by a variety of agents (in the presence of a TP antagonist); inhibition of adenylyl cyclase/priming of protein kinase C is thought to be the mechanism (Matthews and Jones, 1993; Vezza *et al.*, 1993). In addition, there is increased bleeding tendency and increased susceptibility to thromboembolism in the EP<sub>3</sub> receptor knock-out mouse (Ma *et al.*, 2001). DG-041 at 0.03–3  $\mu$ M inhibited the enhancement by sulprostone of ADP- or collagen-induced aggregation in human platelet-rich plasma (PRP) (Heptinstall *et al.*, 2008; Singh *et al.*, 2009). The  $pA_2$  value of 8.3 (our calculation from Heptinstall *et al.*, 2008 data) is probably an underestimate of the affinity owing to plasma protein binding of DG-041. In the rat, DG-041 at 5 or 60 mg·kg<sup>-1</sup> (by gavage; co-administration with clopidogrel) inhibited enhancement of platelet aggregation induced by PGE<sub>2</sub> *ex vivo*; there was no increase in bleeding time compared with clopidogrel alone (Singh *et al.*, 2009). A similar profile was obtained with DG-041 in Phase I trials in healthy subjects (reported in Heptinstall *et al.*, 2008).

## EP<sub>4</sub>-receptor antagonists

### Development

The first EP<sub>4</sub> antagonist to be reported was AH-23848 (Coleman *et al.*, 1994a), a close relative of the selective TP antagonist GR-32191 (Figure 8). Ligand binding studies on human rc-receptors indicated low EP selectivity for AH-23848, with  $K_i$  values for EP<sub>1</sub>, EP<sub>2</sub>, EP<sub>3</sub>, EP<sub>4</sub> and IP receptors being 45, 50, 4.4, 14 and >100  $\mu$ M respectively (Abramovitz *et al.*, 2000). However, its selectivity in functional assays appears to be higher and it has been of considerable utility in differentiating the EP<sub>2</sub>, EP<sub>4</sub> and IP agonist activities of prostanoid ligands. For example, Jones and Chan (2001) used AH-23848 at 30  $\mu$ M to demonstrate that the PGI<sub>2</sub> analogues cicaprost and AFP-07 relax certain vascular preparations by activating both EP<sub>4</sub> and IP receptors. Moreover, Lai *et al.* (2008a) showed that pulmonary artery smooth muscle cells from monocrotaline-treated rats have a reduced IP receptor density and that iloprost-induced cAMP elevation is blocked by AH-23848, and therefore likely to be due to activation of EP<sub>4</sub> receptors.

AH-23848 has since been overtaken by more potent agents. Antagonists in the major group contain an acyl-sulphonamide unit (Figure 5) and show some similarity to the ene-acyl-sulphonamide EP<sub>3</sub> antagonists. Indeed, L-161982 is a methyl analogue of the lead molecule **16** for the Merck EP<sub>3</sub>



**Figure 5** EP<sub>4</sub> receptor antagonists. L-161982 is a methyl analogue of compound **16** in Figure 4. Acyl-sulphonamido residues are shown in red. The bonds indicated by asterisks in MF-498 are subject to oxidative/hydrolytic attack *in vivo*; the corresponding substituents in compound **20** prevent these transformations.

antagonists. L-161982 was reported to have a pA<sub>2</sub> of about 8.5 in rc-EP<sub>4</sub> receptor – adenylyl cyclase assays (Machwate *et al.*, 2001). In a rat native EP<sub>4</sub> assay, L-161982 inhibited PGE<sub>2</sub>-induced cAMP accumulation with an IC<sub>50</sub> of about 30 nM, while forskolin-induced cAMP accumulation was unaffected by 10 μM L-161982. In similar functional assays, CJ-023423 afforded pA<sub>2</sub> values of 8.3 and 8.2 for human and rat rc-EP<sub>4</sub> receptors using the Schild protocol (Nakao *et al.*, 2007). In binding assays, CJ-023423 showed weak affinity for human EP<sub>2</sub> receptors and PAF receptors, while interactions with other prostanoid receptors and a range of non-prostanoid receptors were minimal.

GW-627368 has pK<sub>i</sub> values for binding of 7.0 and 6.8 for human rc-EP<sub>4</sub> and TP receptors respectively; binding to other prostanoid receptors is minimal (Wilson *et al.*, 2006). Correspondingly, a pA<sub>2</sub> of 7.9 was obtained for GW-627368 in a human rc-EP<sub>4</sub> receptor – adenylyl cyclase assay, with good evidence for competition. On human pulmonary vein, GW-627368 had a pA<sub>2</sub> of 7.06 against ONO-AE1-329 (Table 2; Foudi *et al.*, 2008); its higher affinity against PGE<sub>2</sub> may have been due to opposing contractile activity. GW-627368 had a higher affinity on piglet saphenous vein (pA<sub>2</sub> = 9.2) with a

linear Schild plot up to a concentration ratio of about 60; further rightward shift of the agonist curve was insignificant owing to PGE<sub>2</sub> activating a less sensitive EP<sub>2</sub> relaxant system (Wilson *et al.*, 2006). GW-627368 at 10 μM did not affect TP receptor-induced contraction under the same conditions. Results for rabbit saphenous vein were discrepant: Wilson *et al.* (2006) showed that 10 μM GW-627368 did not affect PGE<sub>2</sub>-induced relaxation implying the presence of an EP<sub>2</sub> system only, while Jones and Chan (2005) found a right-shift of about 1 log unit with 1 μM GW-627368, consistent with the presence of EP<sub>2</sub> and EP<sub>4</sub> systems; GW-627368 did not antagonize relaxation induced by either the selective EP<sub>2</sub> agonist ONO-AE1-259 or the PGI<sub>2</sub> analogue, taprostene in the latter experiments. MF-498 (Clark *et al.*, 2008), which is quite similar in structure to GW-627368, undergoes oxidative/hydrolytic metabolism at the three regions indicated in Figure 5; compound **20** was considerably more resistant to attack while retaining high EP<sub>4</sub> antagonist affinity (Burch *et al.*, 2008). The most recent addition to this group is BGC-20-1531 (Maubach *et al.*, 2009). It exhibits surmountable antagonism of PGE<sub>2</sub>-induced relaxation of human cerebral and middle meningeal and dog carotid and middle meningeal

arteries *in vitro*, while having no effect on PGE<sub>2</sub> (probably EP<sub>3</sub>)-induced contraction of human coronary, pulmonary and renal arteries.

A second looser group of amide-containing EP<sub>4</sub> antagonists is represented by ONO-AE2-227, ONO-AE3-208 and CJ-042794 (Figure 5). Binding studies indicate that ONO-AE2-227 and ONO-AE3-208 retain considerable affinity for EP<sub>3</sub> receptors (K<sub>i</sub> = 21 and 30 nM; Mutoh *et al.* 2002; Kabashima *et al.* 2003) and this must be borne in mind when interpreting *in vivo* data (see later). CJ-042794 behaved competitively in functional assays (Schild protocol) involving human and rat rc-EP<sub>4</sub> receptors (Murase *et al.*, 2008a,b). Its EP<sub>4</sub>/EP<sub>3</sub> selectivity ratio is very high, but it still retains measurable affinity for human EP<sub>2</sub> receptors (pK<sub>i</sub> = 6.2).

#### Therapeutic applications

Recent studies involving parenteral administration of several EP<sub>4</sub> antagonists (AH-23848, CJ-023423, CJ-042794, MF-498, ONO-AE3-208) have clearly demonstrated a major involvement of EP<sub>4</sub> receptors in small-animal models of inflammation (Table 3). Joint pain, mechanical and thermal hyperalgesia and oedema were markedly suppressed, often equivalent to the efficacy of selective COX-2 inhibitors such as rofecoxib. A peripheral site of action seems likely given that intraplantar administration of L-161982 suppressed carrageenan-induced mechanical allodynia in the mouse (Kassuya *et al.*, 2007). Whether block of the EP<sub>4</sub> receptor alone produces an effective anti-inflammatory drug in man remains to be seen. Moreover, PGE<sub>2</sub> may have a protective role in inflammation. Takayama *et al.* (2002) showed that PGE<sub>2</sub> suppressed chemokine production stimulated by lipopolysaccharide in human macrophages; L-161982 at 100 nM blocked this action.

Prostanoid EP<sub>4</sub> receptors, indeed nearly all EP receptor subtypes, have been implicated as contributors to colon tumorigenesis caused by excessive production of PGE<sub>2</sub> (Fujino and Regan, 2003; Majima *et al.*, 2003; Masataka *et al.*, 2003; Mutoh *et al.*, 2006). The selective EP<sub>4</sub> agonist ONO-AE1-329 (0.1–1 µM) increased colony formation in the human colon cancer cell line HCA-7 (Mutoh *et al.*, 2002) and L-161982 blocked PGE<sub>2</sub>-induced proliferation of HCA-7 cells (Cherukuri *et al.*, 2007). In both the oxymetazoline model of aberrant crypt foci (putative preneoplastic lesions) and the Min mouse model of intestinal polyp development, the EP<sub>4</sub> antagonist ONO-AE2-227 produced about a 67% reduction in the appropriate scores (Mutoh *et al.*, 2002). Aberrant crypt foci were similarly reduced in EP<sub>4</sub><sup>-/-</sup> mice. In the APC1309 mouse, ONO-AE2-227 had a preferential effect on polyp size, while the EP<sub>1</sub> antagonist ONO-8711 had a more pronounced effect on polyp number, and a combination of antagonists behaved additively (Kitamura *et al.*, 2003b). In the context of (tumour) vascularity, ONO-AE3-208 reduced IL-1β-induced angiogenesis in the mouse cornea at an oral dose of 1 mg·kg<sup>-1</sup> o.d. (Kuwano *et al.*, 2004). Finally, pretreatment of mouse mammary tumour cells with AH-23848 and ONO-AE3-208 followed by washing and immediate injection into immunologically compatible mice reduced pulmonary tumour score (Fulton *et al.*, 2006). These comprehensive studies provide encouragement that EP<sub>4</sub> receptor antagonists

may provide a safer replacement for COX-2 inhibitors in treating colon cancer. As, however, all EP receptor subtypes have been implicated in colon cancer, a pan-PGE synthase inhibitor may be more effective in preventing the key cell proliferative and angiogenic events. Arguably, and somewhat paradoxically, the EP<sub>4</sub> receptor has also been shown to participate in the maintenance of intestinal homeostasis by preserving mucosal integrity. In both EP<sub>4</sub><sup>-/-</sup> mice and wild-type mice treated with an EP<sub>4</sub> antagonist (ONO-AE3-208), susceptibility to the development of colitis was reported in a model of inflammatory bowel disease (Kabashima *et al.*, 2003; Narumiya, 2003).

There has been interest in using EP<sub>4</sub> agonists for their anabolic effects on bone (Raisz, 2006). The EP<sub>4</sub> antagonist L-161982 at a dose of 10 mg·kg<sup>-1</sup>·day<sup>-1</sup> reversed bone formation induced by PGE<sub>2</sub> in the rat, without affecting its diarrhoeal action (Machwate *et al.*, 2001). Also, human mesenchymal stem cells in culture secreted PGE<sub>2</sub> via COX-2, which was associated with production of bone morphogenetic protein-2 (BMP-2), a factor that stimulates differentiation of precursor mesenchymal cells into mature bone. The selective COX-2 inhibitor, NS-398, and the EP<sub>4</sub> antagonist, ONO-AE3-208, (concentration not specified) suppressed BMP-2 expression (Arikawa *et al.*, 2004). These findings identify bone loss as a potential side effect of EP<sub>4</sub> antagonists.

*In vitro*, EP<sub>4</sub> vasodilator systems typically exhibit high sensitivity to PGE<sub>2</sub> and this property is reflected in the role of PGE<sub>2</sub> in maintaining the open state of the ductus arteriosus during gestation in human and animal species. Strong expression of EP<sub>4</sub> (and IP) receptors is found in ductus tissue from the newborn infant and child (Leonhardt *et al.*, 2003). Towards the end of gestation, smooth muscle cells within the ductus migrate to the endothelial lining where they form intimal cushions. Declining PG levels at birth result in ductus constriction, thereby bringing the intimal cushions into close contact and effecting permanent closure. Intimal cushion formation is also driven by EP<sub>4</sub> receptor activation (Yokoyama *et al.*, 2006). The COX inhibitor, indomethacin, is commonly used to treat failure of ductus closure, but its efficacy is poor in a substantial proportion of cases, perhaps related to suppression of intimal cushion development (see Ivey and Srivastava, 2006). AH-23848 blocked PGE<sub>2</sub>-induced relaxation of the rabbit and sheep isolated ductus arteriosus preparations (Smith *et al.*, 1994; Bouayad *et al.*, 2001), while ONO-AE3-208 constricted the ductus of fetal and neonatal rats *in vivo* (Momma *et al.*, 2005). Whether an EP<sub>4</sub> antagonist would be better than a COX inhibitor in treating patent ductus in premature infants is not yet clear.

A second potential role for an EP<sub>4</sub> vasodilator system is the genesis of vascular headache in migraine. EP<sub>4</sub> antagonists block PGE<sub>2</sub>-induced relaxation of human-isolated middle cerebral artery (Davis *et al.*, 2004; Maubach *et al.*, 2009) and the picture has been enlarged to include the interaction of endogenous PGE<sub>2</sub> with calcitonin gene-related peptide release from trigeminal nerves (Maubach *et al.*, 2009). The use of AH-23848 and L-161982 also has provided evidence for a role of EP<sub>4</sub> receptors in substance P release by stretching of the renal pelvic wall (volume expansion) leading to activation of renal sensory afferents and subsequent diuresis/natriuresis (reno-renal reflex) (Kopp *et al.*, 2004).

## FP receptor and prostamide receptor antagonists

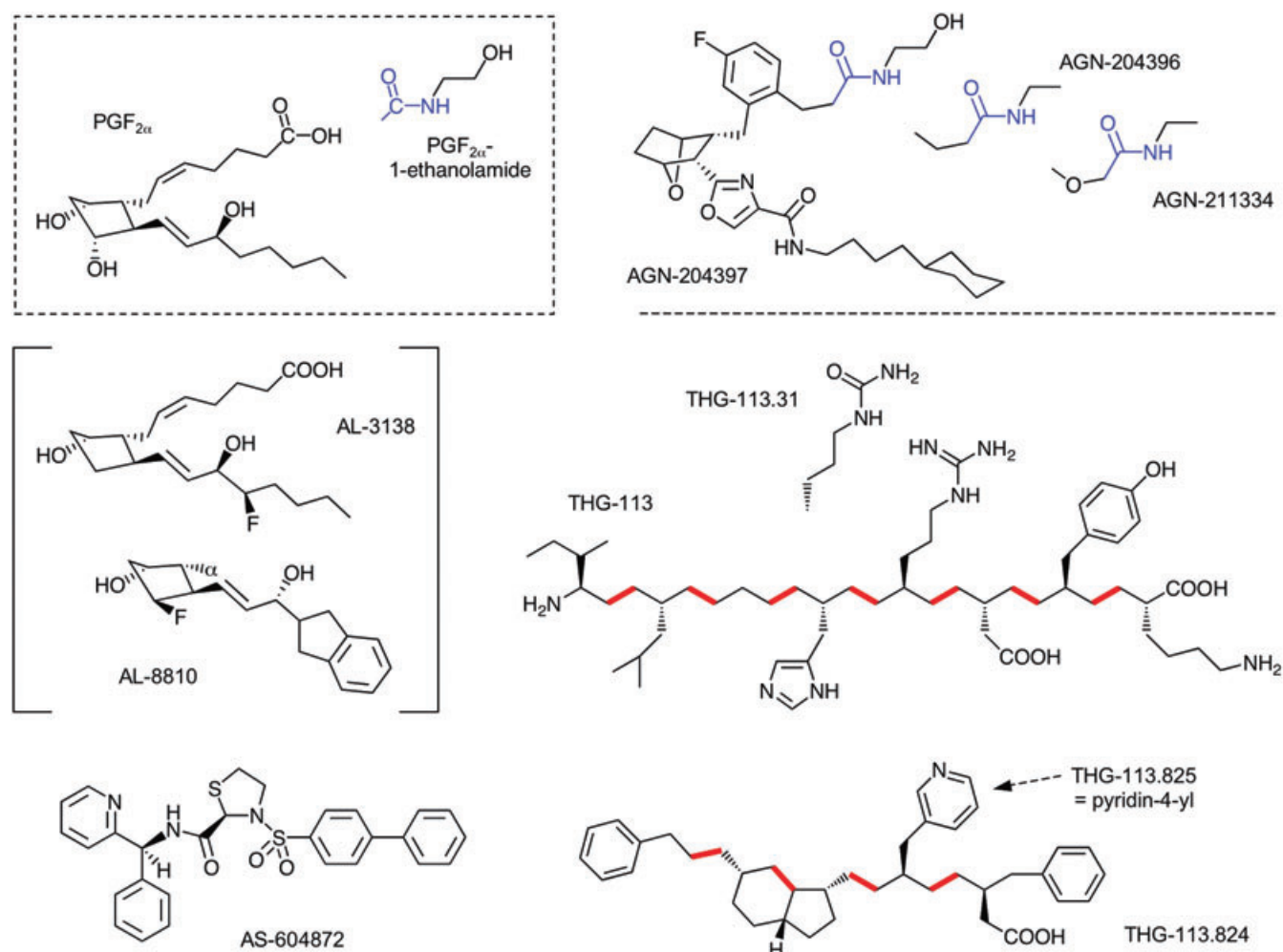
### Development

A variety of FP receptor antagonists have been reported in the past, but none has stood up to rigorous analysis. Thus, PGF<sub>2α</sub>-1-dimethylamine and PGF<sub>2α</sub>-1-dimethylamide (Maddox *et al.*, 1978; Stinger *et al.*, 1982) exhibited no meaningful FP antagonist or agonist activities (Sharif *et al.*, 2000; Woodward *et al.*, 2008). Phloretin (Kitanaka *et al.*, 1993) was non-selective and very weak in blocking fluprostenol-mediated phosphoinositide turnover in rat A7r5 vascular smooth muscle cells (Sharif *et al.*, 2000). Similarly glibenclamide (Delaey and Van de Voorde, 1995) was a weak, non-selective prostanoid antagonist (Sharif *et al.*, 2000).

AL-3138 and AL-8810, both PGF<sub>2α</sub> analogues (Figure 6), have received a degree of acceptance as FP antagonists (Griffin *et al.*, 1999; Sharif *et al.*, 2000). AL-8810 proved useful in studying PGF<sub>2α</sub>-mediated up-regulation of the orphan nuclear receptor Nur 77 (Liang *et al.*, 2004). However, further studies reveal that they are neither potent nor selective. For example, AL-8810 appears to block TP

receptors (Hutchinson *et al.*, 2003), an effect that has been confirmed in human rc-TP receptor stable transfectants (A.H. Krauss and D.F. Woodward, unpublished). In addition, both agents are FP partial agonists. This can be seen in the original report of AL-8810 on phosphoinositide turnover (Griffin *et al.*, 1999) and is corroborated by the finding of myogenic activity in the mouse uterus (Hutchinson *et al.*, 2003). AL-8810 induced a more pronounced Ca<sup>2+</sup> signal in human rc-FP stable transfectants (Y. Liang and D.F. Woodward, unpublished) and was a full agonist in the cat isolated iris preparation (Woodward *et al.*, 2007); the latter action was not blocked by a prostamide antagonist (Woodward *et al.*, 2007; see later), so it presumably reflects FP receptor agonism.

A series of octapeptides (THG-131 derivatives, Figure 6) have been claimed to possess selective FP antagonist activity (Chemtob and Peri, 2006; Peri *et al.*, 2006). In particular, THG-113.31 at 1 μM markedly inhibited PGF<sub>2α</sub>-induced contraction of pig retinal blood vessels, while having minimal effect on contraction to 17-phenyl PGE<sub>2</sub>, U-46619, phenylephrine, Ang II and endothelin-1; the inhibition of PGF<sub>2α</sub>



**Figure 6** FP receptor and prostamide receptor antagonists. The natural ligand PGF<sub>2α</sub> and its 1-ethanolamide derivative, prostamide F<sub>2α</sub>, are shown in the box. AL-3138 and AL-8810 are FP partial agonists in many systems; α = corresponding side-chain in PGF<sub>2α</sub>. The THG analogues are peptides: amide (CO-NH) residues are shown as red bars. The AGN analogues (upper right) are prostamide receptor antagonists; C1-amide residues are shown in blue.

contraction was insurmountable. In addition, THG-131.31 inhibited the associated phosphoinositide hydrolysis with an  $IC_{50}$  of about 30 nM. However, THG-131.31 competed poorly with [ $^3H$ ]-PGF $_{2\alpha}$  for binding to human rc-FP receptors (~13% at 10  $\mu$ M). Several other studies have cast doubt on the utility of THG-131.31. At 10  $\mu$ M, THG-113.31 showed fairly weak antagonism of the contractility of longitudinal and circular strips of sheep myometrium induced by PGF $_{2\alpha}$ ; there was no effect on PGE $_2$  contractions (Hirst *et al.*, 2005). Also, 10  $\mu$ M THG-113.31 had no effect on PGF $_{2\alpha}$ -induced contraction of human pregnant myometrium, while inhibiting spontaneous and oxytocin-induced contractions at much lower concentrations (Friel *et al.*, 2005). Finally, Doheny *et al.* (2007) showed that THG-113.31 at 10–50  $\mu$ M enhanced BK $_{Ca}$  channel opening in isolated myocytes from human uterus, an effect that was reversed by iberiotoxin. Several chemically simpler peptidomimetics in the THG series showed more potent block of PGF $_{2\alpha}$ -induced contraction than THG-113.31 (Figure 6, Table 2; Peri *et al.*, 2006); it would be of interest to have full pharmacological profiles.

Currently, the most convincing FP antagonist is the non-prostanoid AS-604872 (Cirillo *et al.*, 2007). It has  $K_i$  values of 35, 158 and 323 nM for human, rat and mouse rc-FP receptors and its selectivity was 20-fold for EP $_2$  receptors and greater than 300-fold for other prostanoid receptors. AS-604872 showed no agonist activity in a human FP receptor – inositol phosphate assay and had an  $IC_{50}$  of 47 nM against PGF $_{2\alpha}$ . *In vivo*, AS-604872 (1–30 mg·kg $^{-1}$ , i.v.) inhibited PGF $_{2\alpha}$ -induced uterine contraction in the non-pregnant rat; inhibition of oxytocin-induced contraction was slight.

Prostamide research originated from studies on neutral PGF $_{2\alpha}$  analogues and notably bimatoprost (17-phenyl PGF $_{2\alpha}$ -1-ethylamide), an effective anti-glaucoma drug (Woodward *et al.*, 2003; 2004). As previously mentioned, neutral PGF $_{2\alpha}$  analogues show only weak affinity for FP receptors (Maddox *et al.*, 1978; Schaaf and Hess, 1979), a profile subsequently confirmed for –OH, –OCH $_3$  and –CON(R) $_2$  C1-substitutes (Woodward *et al.*, 2000; 2008; Matias *et al.*, 2004). The pharmacology of bimatoprost was similar, with no meaningful activity at FP receptors but pronounced activity in certain preparations such as cat lung strip, cat iris, rabbit uterus, and human ciliary smooth muscle cells (Liang *et al.*, 2003; Woodward *et al.*, 2003b; Matias *et al.* 2004; Chen *et al.* 2005). The pharmacology of bimatoprost appeared indistinguishable from that of PGF $_{2\alpha}$ -1-amides, but this could not be rationalized until the discovery that anandamide (arachidonic acid-1-ethanolamide) was a substrate for COX-2 (Yu *et al.*, 1997).

PGE $_2$ -1-ethanolamide (prostamide E $_2$ ) was the first prostamide to be discovered and was identified as the major product following addition of anandamide to rc-COX-2 or cells expressing COX-2, but not COX-1 (Yu *et al.*, 1997). Subsequently, more extensive studies demonstrated that COX-2 oxidizes anandamide to endoperoxide intermediates, which are converted by specific PG synthases to the various prostamides (Kozak *et al.*, 2002; Koda *et al.*, 2004; Yang *et al.*, 2005; Moriuchi *et al.*, 2008). Despite being different terminal biosynthetic products, the evidence to date suggests that prostamides D $_2$ , E $_2$  and F $_{2\alpha}$  interact with a single receptor to exert their effects. Prostamide F $_{2\alpha}$  and its analogues are, however,

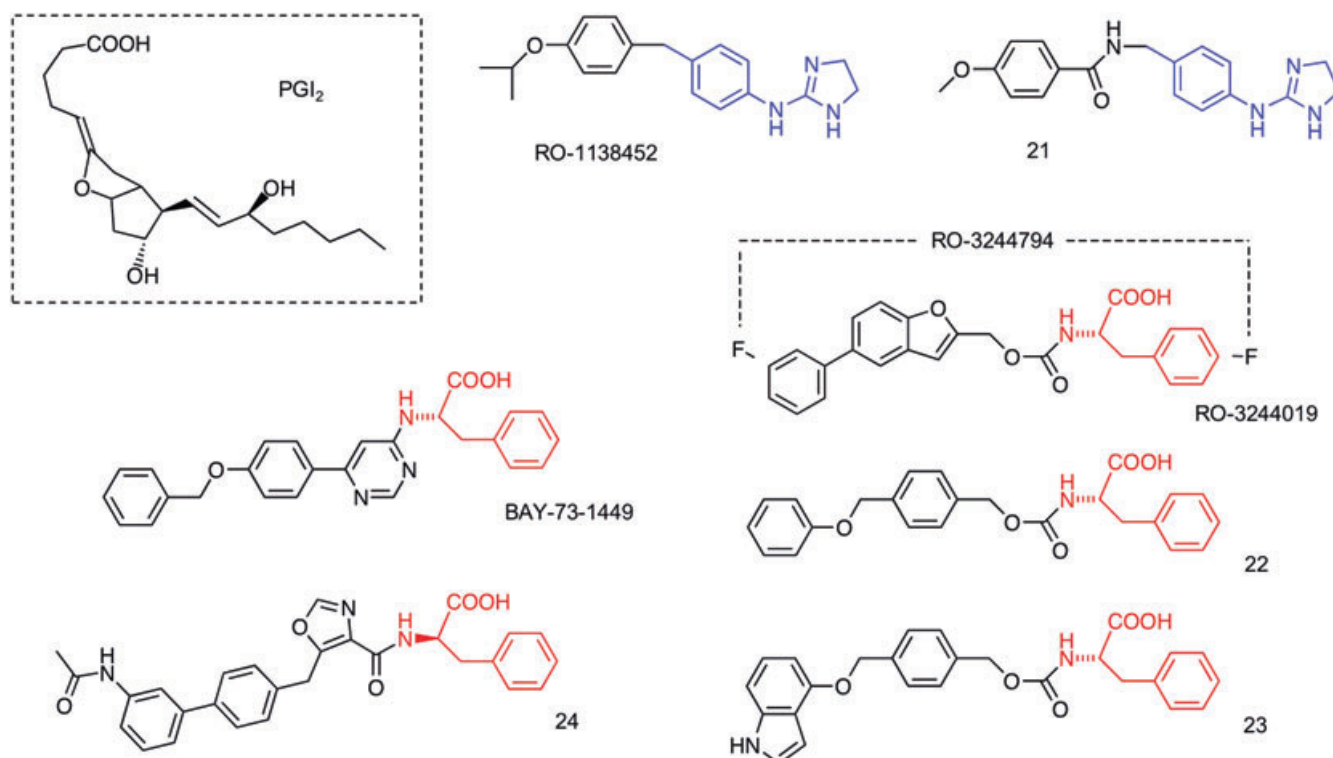
about 10 times more potent than prostamides D $_2$  and E $_2$  (Woodward *et al.*, 2007).

Initial pharmacological characterization of the prostamides, for example prostamide E $_2$  (Ross *et al.*, 2002), relied on agonist studies. In the context of PGF analogues, FP agonists (17-phenyl PGF $_{2\alpha}$  and PGF $_{2\alpha}$ ) and bimatoprost produced Ca $^{2+}$  signals in entirely different cells in a cat iris smooth muscle cell preparation (Spada *et al.*, 2005), suggesting the existence of a receptor with a distinct preference for prostamide F agonists. The receptor structure appears to involve heterodimerization of the wild-type and alternative mRNA splicing variants of the FP receptor, both encoded by *PTGFR*, the FP receptor gene (Liang *et al.*, 2008). This FP/alt-FP co-expression is analogous to the isoprostane binding site which is formed as a result of IP/TP receptor heterodimerization (Wilson *et al.*, 2004).

The strategy for discovering a prostamide antagonist was to identify antagonists in cluster 2 of the prostanoid receptor evolutionary tree (Narumiya *et al.*, 1999) and then form corresponding C1-amides. Cluster 2 contains the TP receptor, whose stable agonists have been a starting point for antagonists at other receptors (DP $_1$ , see Figure 1; EP $_1$ , see Figure 3). Based on the oxabicycloheptane analogue BMS-180291 (Figure 8) (Webb *et al.*, 1993), two prototype prostamide antagonists AGN-204396 (Figure 6) and AGN-204397 were identified (Woodward *et al.*, 2007; 2008). These agents showed good prostamide F/FP selectivity, but were of low affinity ( $pA_2$  ~5.5) and also blocked TP receptors. Substituting oxygen at C3 dramatically enhanced prostamide affinity, by as much as 100-fold for AGN-211334 and AGN-211335 (Wan *et al.*, 2007; Liang *et al.*, 2008; Woodward *et al.*, 2008). AGN-211334 and AGN-211335 potently inhibited prostamide F $_{2\alpha}$  and bimatoprost responses in cat iris preparations, but did not alter responses to FP agonists (Wan *et al.*, 2007; Liang *et al.*, 2008). AGN-211334 blocked the increase in conventional aqueous humour outflow produced by bimatoprost in the human perfused anterior segment preparation (Wan *et al.*, 2007), thereby demonstrating that the effects of bimatoprost in the human eye are prostamide receptor-mediated; previously, it has been suggested that bimatoprost's activity is dependent on deamidation to the FP-active free acid in ocular tissue (Camras *et al.*, 2004). AGN-211335 blocked the secondary Ca $^{2+}$  wave, myosin light chain phosphorylation, and Cyr 61 up-regulation induced by bimatoprost in the FP/alt-FP prostamide system (Liang *et al.*, 2008). These second-generation prostamide antagonists are likely to be sufficiently potent for *in vivo* studies.

#### Therapeutic applications

Prevention of pre-term labour (tocolysis) is probably the only therapeutic modality where a FP antagonist may be of value. This is a serious and unmet medical need given that premature birth accounts for 60–80% of perinatal deaths (Goldenberg, 2002). Parturition is prevented in FP $^{-/-}$  mice (Sugimoto *et al.*, 1997) confirming a significant role for PGF $_{2\alpha}$  (see Challis *et al.*, 2002 for a review of PG involvement). THG-113.31 is tocolytic in the pregnant sheep (Hirst *et al.*, 2005) and may be useful for delaying pre-term birth (Olson, 2005). AS-604872 suppressed spontaneous uterine contractions in late-term pregnant rats and delayed preterm birth caused by



**Figure 7** IP receptor antagonists. The natural ligand PGI<sub>2</sub> (prostaglandin I<sub>2</sub>) is shown in the box. 2-(Phenylamino)-imidazole moieties are shown in blue and phenylalanine residues in red (*S*-configuration in compound **24**). RO-3244794 is a difluoro analogue of RO-3244019.

mefipristone in pregnant mice; AS-604872 appeared to be more effective than the  $\beta_2$ -adrenoceptor agonist ritodrine (Chollet *et al.*, 2007). It is not yet clear whether these findings will translate into an effective drug in human reproduction.

Prostanoid FP receptors have been implicated in cardiomyocyte hypertrophy (Pönicke *et al.*, 2000; Xu *et al.*, 2008) and cancer (Fujino and Regan, 2001) and may play a role in regulating water and solute transport based on *in situ* hybridization studies in the mouse kidney (Saito *et al.*, 2003). The utility of a selective FP antagonist is, however, unclear.

The therapeutic utility of prostamide antagonists is similarly uncertain. Increased anandamide levels may result in the formation of prostamides as major products in inflammation and infection (Glass *et al.*, 2005). Testing of AGN 211334, or a close congener, in relevant animal models should provide evidence for or against a functional role of prostamides.

## IP receptor antagonists

### Development

IP receptor antagonists are a recent development based on the potential role of PGI<sub>2</sub> in mediating pain (Bley *et al.*, 1998). In an extensive series of studies, two structurally distinct classes of selective IP antagonist emerged from focused chemical library screening and synthetic chemistry (Bley *et al.*, 2006) (Figure 7). The 2-(phenylamino)-imidazole series is represented by RO-1138452 and compound **21** (Clark *et al.*, 2004; Keitz *et al.*, 2004), while the *N*-substituted phenylalanine series has a 'traditional' carboxylate in the phenylalanine

residue and is typified by RO-3244019 (Fitch *et al.*, 2004) and its difluoro analogue RO-3244794 (Bley *et al.*, 2006).

RO-1138452 has high affinity for human native (platelet) and rc-IP receptors, with pK<sub>i</sub> values of 9.3 and 8.7 respectively in studies utilizing [<sup>3</sup>H]-iloprost, although it also displays considerable affinity for PAF (7.9) and imidazole (8.3) receptors (Bley *et al.*, 2006). In functional studies in platelets (Jones *et al.*, 2006), pA<sub>2</sub> values were lower than K<sub>i</sub> values obtained in radioligand competition studies with platelet membrane preparations and for inhibition of carbacyclin-induced cAMP formation in cells over-expressing rc-IP receptors (Bley *et al.*, 2006); the difference was attributed to protein binding in studies involving PRP. The pA<sub>2</sub> values obtained in isolated blood vessel preparations (human pulmonary artery 8.20, guinea pig aorta 8.39 and rabbit mesenteric artery 8.12) were intermediate between values obtained in PRP and plasma membranes/cells in buffer (Jones *et al.*, 2006). The slight suppression of the cicaprost maximum response seen with higher concentrations of RO-1138452 was attributed to functional antagonism emanating from the (albeit weak) EP<sub>3</sub> agonist action of cicaprost. However, RO-1138452 displayed an insurmountable antagonist profile in studies of chemokine release from human airway epithelial cells using taprostene (see Table 1) as IP agonist (Ayer *et al.*, 2008). Moreover, RO-1138452 inhibition of taprostene-induced cAMP response element-dependent transcription was not reversed over a 20-h 'washout' period. These data could not be ascribed to covalent receptor inactivation, allosterism or a state of antagonist hemi-equilibrium and may be due to a pseudo-irreversible interaction with the IP receptor (Ayer *et al.*, 2008).



The N-substituted phenylalanines **22** and **23** (Nakae *et al.*, 2005) are weak IP antagonists, while the greater potency of **24** is associated with an S-configuration in the phenylalanine residue (Brescia *et al.*, 2007). RO-3244794 has no meaningful activity at EP<sub>1</sub>, EP<sub>3</sub>, EP<sub>4</sub> and TP receptors; no data were provided for DP<sub>1</sub>, DP<sub>2</sub>, EP<sub>2</sub> or FP receptors (Bley *et al.*, 2006). While RO-1138452 did not block EP<sub>2</sub> receptor-mediated relaxation in guinea pig aorta ( $pA_2 < 6.0$ ; Jones *et al.*, 2006), RO-3244794 had a  $pA_2$  of 6.92 for the human rc-EP<sub>2</sub> receptor expressed in HEK-293 cells (S.M. Hill and M.A. Giembycz, unpubl. data). Compound **24** did not bind to human EP<sub>2</sub> and EP<sub>4</sub> receptors (Brescia *et al.*, 2007). Strictly speaking, the missing data related to these compounds (e.g. RO-1138452 at DP<sub>2</sub> and FP receptors; Jones *et al.*, 2006) need to be addressed. In the context of systemic drug design, RO-3244794 has much greater oral bioavailability than RO-1138452 (51% vs. 0.7% in the rat) (Bley *et al.*, 2006).

#### Therapeutic applications

Pain has been the initial focus for IP antagonists based on IP receptor agonist effects and IP receptor distribution (reviewed by Bley *et al.*, 1998). Altered pain perception and inflammation were observed in IP<sup>-/-</sup> mice (Murata *et al.*, 1997). Subsequently, IP antagonists were shown to reduce pain responses in models where prostanoids have been implicated. These include acetic acid-induced abdominal constriction, mechanical hyperalgesia produced by carrageenan and pain associated with models of osteoarthritis and inflammatory arthritis (Bley *et al.*, 2006; Pulichino *et al.*, 2006). Such analgesic effects are accompanied by anti-inflammatory properties, which is invariably the case. In a collagen-induced arthritis model in mice, **21** produced effective inhibition when administered as a pretreatment, but was inactive when given after the initiation of the arthritis (Pulichino *et al.*, 2006). RO-3244794 and indomethacin were equi-effective in reducing carrageenan-induced rat paw oedema and more effective than rofecoxib in inhibiting the foot weight distribution change associated with intra-articular injection of monoiodoacetate (Bley *et al.*, 2006). Ostensibly, these results imply that IP receptors provide a singular target that would result in drugs that are at least as effective as NSAIDs and COX-2 inhibitors. The role of EP receptors in pain/inflammation has already been discussed. Despite a satisfactory preclinical profile, strong circumstantial evidence suggests that EP<sub>1</sub> antagonists were a failure in clinical trials. This, in turn, tends to cast doubt over the clinical prognosis for IP antagonists. Given the number of prostanoids that may be released locally and the even greater number of target receptors available, it may be regarded as counter-intuitive to propose that inflammation can be attributed to only one type of prostanoid receptor.

A role for PGI<sub>2</sub> in mediating the sensitized release of substance P from rat dorsal root ganglion neurons has been suggested based on studies with **23** (Nakae *et al.*, 2005). Related to this is the potential use of IP antagonists for bladder disorders. In rat models of bladder function, RO-3244019 dose-dependently decreased bladder contraction frequency and increased micturition threshold and voiding interval (Cefalu *et al.*, 2007). RO-3244019 was also effective in

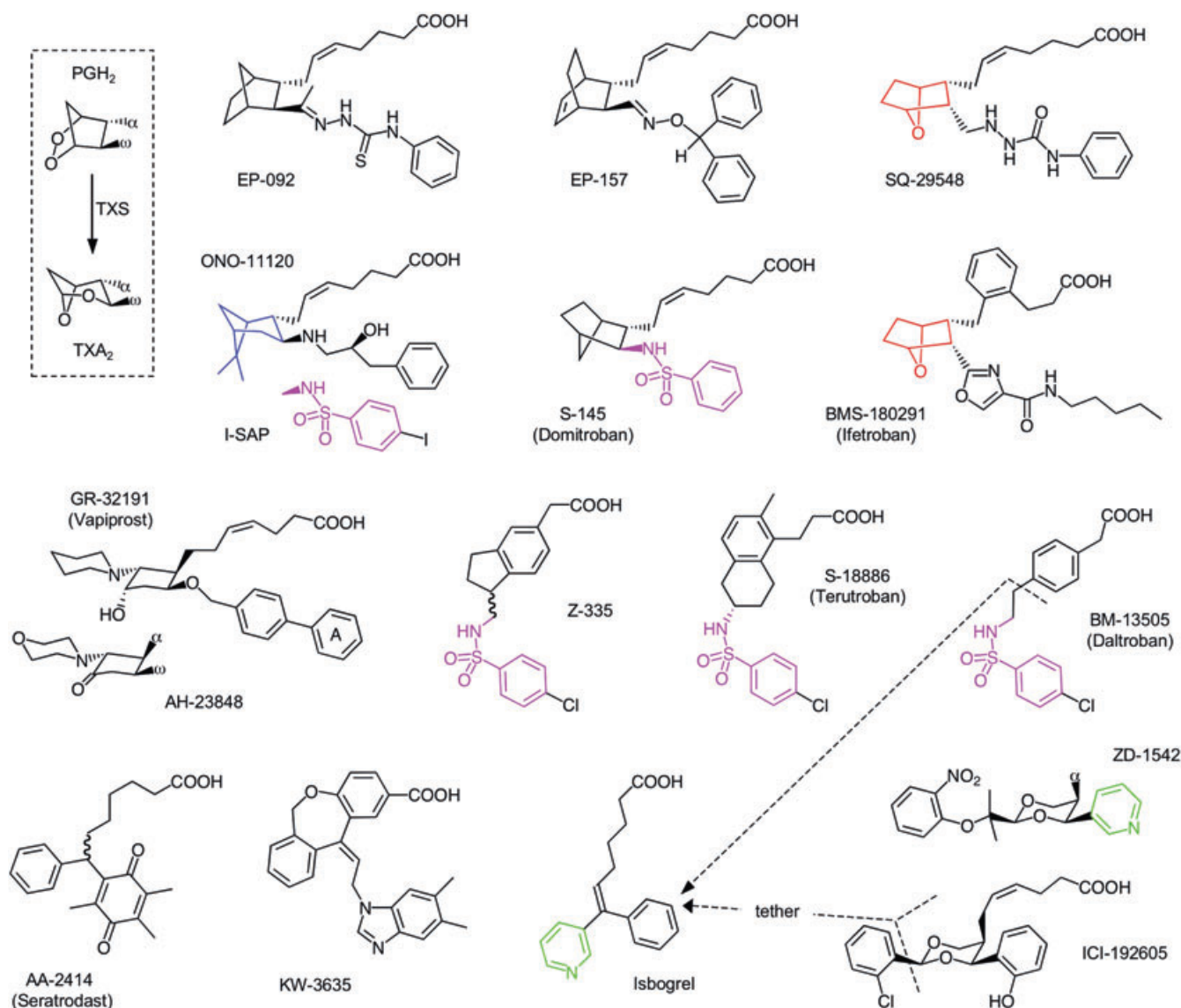
treating neurogenic detrusor overactivity arising from spinal cord injury in the rat (Khera *et al.*, 2007). Again, these results are of uncertain predictive value for clinical success, because indomethacin exhibited good activity in these models. The IP antagonist, BAY-73-1449 (Figure 7), was effective in acutely reducing shunt vessel blood flow in a rat model of portal hypertension (Bexis *et al.*, 2008); full details of its pharmacology are unavailable.

Finally, side effects originating from perturbation of the TXA<sub>2</sub>/PGI<sub>2</sub> balance could be greater with IP antagonists than with COX-2 inhibitors, including hypertension, stroke, myocardial infarction and atherosclerosis. Nephrotoxicity and K<sup>+</sup> and Na<sup>+</sup> retention also may occur (Nasrallah and Hébert, 2005), because prostacyclin synthase (PGIS) and COX-2 deficiencies produce similar renal toxicity. This suggests a protective role for PGI<sub>2</sub>, but it is important to note that no marked renal phenotype occurs in TP<sup>-/-</sup> or IP<sup>-/-</sup> mice (Breyer and Breyer, 2000; Yokoyama *et al.*, 2002; Nasrallah and Hébert, 2005).

## TP receptor antagonists

#### Development

Elucidation of the structures of PGH<sub>2</sub> and TXA<sub>2</sub> (Figure 8) was soon followed by a range of carba/thia bicyclic analogues (see Wilson and Jones, 1985). One aim was to produce a chemically stable TP agonist (e.g. U-46619, STA<sub>2</sub>); another was to investigate the potential for TP receptor antagonism. Thus, pinane-TXA<sub>2</sub> (PTA<sub>2</sub>) was reported to block both constriction of cat coronary artery and aggregation of human platelets elicited by U-46619 (Nicolaou *et al.*, 1979). However, this analogue often behaves as a partial agonist, inducing a 60% maximal activation in some TP systems (Jones *et al.*, 1982; Tymkewycz *et al.*, 1991). More useful TP antagonists emerged from additional modification of the  $\omega$ -chain. The pinane analogue ONO-11120 (Figure 8) is a true antagonist (Katsura *et al.*, 1983), while EP-045 (Jones *et al.*, 1982) and EP-092 (Armstrong *et al.*, 1985) were developed from the PG endoperoxide analogue, 9,11-etheno PGH<sub>2</sub>, also a partial agonist (Jones *et al.*, 1982). SQ-29548 (Ogletree *et al.*, 1985) and BMS-180291 (Ogletree *et al.*, 1993) have a hybrid bicyclic system (oxabicyclo[2.2.1]heptane) and differ from the natural agonists in the *cis*-orientation of the  $\alpha$  and  $\omega$ -chains. While GR-32191 (Lumley *et al.*, 1989) and ICI-192605 (Brewster *et al.*, 1988) still retain a prostanoid skeleton, other TP antagonists do not, including L-655240 (related to indomethacin; Hall *et al.*, 1987), BM-13505 (daltroban; Yanagisawa *et al.*, 1987), KW-3635 (Karasawa *et al.*, 1991a,b), AA-2414 (seratrodast; Ashida *et al.*, 1989) and even the K<sub>ATP</sub>-channel blocker glibenclamide (Cocks *et al.*, 1990). The attractively simple chemistry involved in the synthesis of daltroban has spurred the development of higher-potency, non-prostanoid antagonists containing a (*p*-halo)-benzenesulphonylaminomethyl residue at a critical distance from the carboxylate, for example, Z-335 (Tanaka *et al.*, 1998) and S-18886 (terutroban; Cimetière *et al.*, 1998). A similar residue in prostanoid molecules such as S-145 (domitroban; Mihara *et al.*, 1989), I-SAP (Naka *et al.*, 1992) (Figure 8), ONO-NT-126 (*p*-bromo;



**Figure 8** TP receptor antagonists. Conversion of PGH<sub>2</sub> to TXA<sub>2</sub> by thromboxane synthase (TXS) is shown in the box;  $\alpha$  and  $\omega$  represent natural 2-series side-chains. The pinane-thromboxane residue (related to 1(*S*)- $\alpha$ -pinene) is shown in blue; the 6-oxabicyclo(2.2.1)heptane system is in red. AH-23848 has the same  $\alpha$ - and  $\omega$ -chains as GR-32191. Benzenesulphonamide residues present in both prostanoid and non-prostanoid antagonists are shown in cerise. TP antagonists with two types of additional activity are presented. (A) IP agonism is conferred by the diphenylmethyl-heteroatomic unit in the bicyclo[2.2.2]octene analogue EP-157. (B) TXS inhibitory activity is conferred by the pyridin-3-yl residue (green) in isbogrel and ZD-1542 and by a similar replacement for ring A in relatives of GR-32191 (e.g. GR-83783; see text). Additionally, the broken arrows (lower right) typically indicate attachment of part of a TP antagonist to isbogrel (or ridogrel) to generate novel combined TP antagonist/TXS inhibitors; the tether (0–11 carbon units) has also been attached to the left-hand phenyl ring of ICI-192605 (2–8 carbon units) (Ackerley *et al.*, 1995).

Nakahata *et al.*, 1990) and oxa- and thia-bicyclo[3.1.0]hexane derivatives (Kamata *et al.*, 1990) also confers high TP affinity.

Differences in antagonist affinity constants between platelet and vascular smooth systems have stimulated much debate about the existence of TP receptor subtypes (Mais *et al.*, 1985; 1988; Swayne *et al.*, 1988; Morinelli *et al.*, 1989; Masuda *et al.*, 1991; Tymkewycz *et al.*, 1991; Folger *et al.*, 1992). Species heterology, the difficulty of distinguishing partial agonism from functional antagonism, and incomplete equilibration of high-affinity antagonists have been confounding factors. For example, antagonism of U-46619 by BMS-180291

on human platelets was surmountable for the shape-change response, but insurmountable for aggregation; the rate of aggregation was also slowed (Ogletree *et al.*, 1993). Similar profiles had been reported previously for EP-092 (Armstrong *et al.*, 1985) and GR-32191 (Lumley *et al.*, 1989) on human platelets, and for EP-169 and AH-23848 on human and rat platelets, but not on rabbit platelets where their affinities are lower (Tymkewycz *et al.*, 1991). However, against the slow-acting high-affinity TP agonist EP-171 (Jones *et al.*, 1989), GR-32191 did not alter the aggregation rate (Lumley *et al.*, 1989). It is likely that slow dissociation of a high-affinity antagonist from the TP receptor retards U-46619 occupancy

in the early stage of the aggregation response thereby favouring the disaggregation process and insurmountability; in contrast, shape change, which does not fade, allows a true measure of the equilibrium state. In pig platelets, longer pre-incubation times were required for S-145 than SQ-29548 owing to the smaller association rate constant of the former antagonist (Mihara *et al.*, 1989). BMS-180291 also had a Schild slope greater than unity on guinea pig aorta (Zhang *et al.*, 1996). However, in the low nanomolar range BMS-180291 requires up to 2 h to reach steady state on the aorta (Jones *et al.*, 2008) and the non-ideality may simply reflect retarded diffusion of a high-affinity ligand ( $pA_2 = 9.8$ , Table 2) through the extracellular space.

Radioligand binding studies of TP receptors coincided with these functional measurements. Two saturable binding sites were identified on human platelets using [ $^3$ H]-9,11-epoxymethano PGH<sub>2</sub>, the first radioligand developed for the TP receptor (Armstrong *et al.*, 1983; Pollock *et al.*, 1984). Binding to the more abundant site was displaced by EP-045 at concentrations similar to those required to inhibit [ $^3$ H]-phosphatidate formation, Ca<sup>2+</sup> elevation and aggregation induced by the TP agonist. A less abundant higher-affinity binding site was also identified, but it did not show the characteristic preference for a 15(S) configuration in the natural prostanoid  $\omega$ -chain. The presence of high- and low-affinity binding sites for agonist (but not antagonist) ligands was also evident using [ $^3$ H]-trimetoquinol (Ahn *et al.*, 1988) and [ $^3$ H]-SQ-29548 (Hedberg *et al.*, 1988). Further studies showed that the high-affinity site was associated with the platelet shape change (and increase in cytosolic Ca<sup>2+</sup>), while the lower-affinity site was associated with aggregation [and activation of phospholipase C (PLC)] (Dorn, 1989; Takahara *et al.*, 1990). [ $^3$ H]-GR-32191 played an important role in the elucidation of these relationships by binding reversibly to the 'shape change site' and irreversibly to the 'aggregation site' (Takahara *et al.*, 1990). It is difficult to explain this irreversibility given that GR-32191 does not obviously contain a chemically reactive group (Figure 8). Detailed studies have shown that 30-min exposure of human platelets to GR-32191 resulted in about 50% loss of binding sites for either [ $^3$ H]-GR-32191 or [ $^3$ H]-SQ-29548, while neither SQ-29548 nor BM-13177 affected B<sub>max</sub>. It was speculated that GR-32191 binds to internalized TP receptors (Armstrong *et al.*, 1993); the zwitterionic nature of GR-32191 at neutral pH may be relevant. A light-activated, covalent-bonding TP antagonist, azido-BSP, also discriminated these platelet sites by blocking aggregation but not shape change induced by U-46619 (Zehender *et al.*, 1988). The subsequent identification of a second TP receptor isoform (TP $\beta$ ) from a human umbilical vein endothelial cDNA library (Raychowdhury *et al.*, 1994) and the detection of mRNA for the  $\alpha$  and  $\beta$  isoforms in human platelets (Hirata *et al.*, 1996) would appear to complete the argument. However, these isoforms, which arise by alternative gene splicing and differ only in their cytoplasmic tails, do not show the ligand discrimination typical of the high- and low-affinity binding sites. Finally, significant expression of the TP $\alpha$  isoform only was found in human platelets (Habib *et al.*, 1999).

The seventh transmembrane domain (TM-7), which is strictly conserved in all the TP receptors characterized to date,

is critical to TP agonist and antagonist function. Point mutations in this domain in the human TP receptor severely suppressed the binding of SQ-29548 (Funk *et al.*, 1993b). Chimeric substitutions of the human TP receptor with the corresponding TM 1, 2 and 4 from rat resulted in modest suppression of SQ-29548 binding, lesser suppression of I-BOP (TP agonist) binding, and a poor correlation between the data sets (Dorn *et al.*, 1997).

Several different inhibitory properties have been combined with specific TP antagonism, either by chance or deliberately. For example, TP antagonists based on PGH<sub>2</sub> with diphenylmethyl-oxime (e.g. EP-157, Figure 8) or diphenylmethyl-azine residues in the  $\omega$ -chain were found to activate IP receptors in platelet and vascular systems (Armstrong *et al.*, 1986; 1989; Jones *et al.*, 1993). A diaryl-hetero(cyclic) moiety is critical to the IP agonism (Jones *et al.*, 1993). Related compounds lacking a prostanoid ring system (e.g. octimibate) showed similar profiles (Merritt *et al.*, 1991a,b), with BM-45778 being the most potent of a large series of non-prostanoid prostacyclin mimetics synthesized by Bristol-Myers Squibb (Meanwell *et al.*, 1994; Seiler *et al.*, 1997). Accurate estimation of the TP antagonist affinities of many of these compounds is difficult owing to their high lipophilicity, slow onset/offset (Jones *et al.*, 1997) and, in certain instances, an ability to inhibit (non-prostanoid) Gq-PLC-driven responses (Chow *et al.*, 2001).

Combining TP receptor antagonism with thromboxane synthase (TXS) inhibition has been an extensively investigated strategy, with the aim of balancing antagonistic/inhibitory activities several hours after dosing. TXS inhibition is expected to divert PGH<sub>2</sub> to PGD<sub>2</sub> and PGI<sub>2</sub> (Vermylen *et al.*, 1981; Smith, 1982), which both inhibit human platelet activation, and usually requires the presence of either a (N)-imidazole as found in dazoxiben (Randall *et al.*, 1981) or a pyridin-3-yl group as in ridogrel (Hoet *et al.*, 1990) and the related CV-4151 (isbogrel, Figure 8, Imura *et al.*, 1988). ZD-1542 (Brownlie *et al.*, 1993) is a pyridin-3-yl derivative related to ICI-192605, while GR-83783 (Campbell *et al.*, 1991a), a relative of GR-32191, has a 4-(pyridin-3-yl)-phenyl moiety (Figure 8). The (N)-benzimidazole KW-3635 did not inhibit cow platelet TXS at 100  $\mu$ M (Miki *et al.*, 1992). Several groups have successfully expanded the biaryl region of ridogrel (Cozzi *et al.*, 1994) or combined sulotroban/daltroban moieties with ridogrel/isbogrel moieties (Figure 8), resulting in GR-108774 (Campbell *et al.*, 1991b), CGS-22652 (Bhagwat *et al.*, 1993) and compound **36** in Soyka *et al.* (1993). In addition, Zeneca have used various tethers to connect the whole or part of the ICI-192605 nucleus to either a dazoxiben or an isbogrel nucleus (Figure 8) (Ackerley *et al.*, 1995). BM-531 and BM-573 are combined TP antagonist/TXS inhibitors lacking a carboxylic acid group (Dogné *et al.*, 2001; Rolin *et al.*, 2001).

Finally, YM-158 has similar high affinity for TP and cys-LT receptors in guinea pig trachea (Arakida *et al.*, 1998) (see later).

#### Therapeutic applications

The discovery of thromboxanes was Nobel prize-winning research. Thromboxane A<sub>2</sub> is undoubtedly important in

regulating cardiovascular homeostasis. Its biosynthesis by platelets and other tissues and highly potent actions on platelets and blood vessels provided a strong impetus for the development of TP antagonists (Patrono, 1990; Patscheke, 1990; Davis-Bruno and Halushka, 1994). However, these agents have not been a success to date. Clear evidence of benefit was not established in early studies (Misra, 1994), but economics was also a major factor. Low-dose aspirin, which targets platelet TXA<sub>2</sub> synthesis by irreversibly acetylating COX-1, has proven benefits (Patrignani *et al.*, 1982; Fitzgerald *et al.*, 1983; Reilly and Fitzgerald, 1987). More potent TP antagonists, with superior pharmacokinetic profiles, were developed, but these also do not appear successful at the commercial level. The reasons are not entirely clear. At the pharmacological level, a TP antagonist would appear preferable to low-dose aspirin, especially in the light of recent events surrounding COX-2 inhibitors.

1. The clinical side effects associated with COX-2 inhibitor therapy reveal the TXA<sub>2</sub>/PGI<sub>2</sub> balance as more delicate to perturbation and more important than was previously believed.
2. Low-dose aspirin does not always have the required TXS/PGIS selectivity (Knapp *et al.*, 1988; Patscheke, 1990).
3. Isoprostanes are alternative endogenous human TP receptor agonists and hence blockable by TP antagonists: their non-enzymatic formation would not be affected by aspirin, other COX inhibitors or TXS inhibitors (Kawikova *et al.*, 1996; Gardan *et al.*, 2000; Janssen *et al.*, 2001).
4. TP antagonists exhibit cardio-protective effects that are not shared by aspirin (Gomoll and Ogletree, 1994; Grover *et al.*, 1994).

Given these considerations, a potent, highly selective TP antagonist may be worth revisiting in large-scale clinical trials on cardiovascular disease. As oxidative stress and resultant formation of isoprostanes is now linked to atherogenesis (Dogné *et al.*, 2005), an initially unappreciated dimension to the TXA<sub>2</sub>/PGI<sub>2</sub> balance and cardiovascular risk is made manifest. Specifically, the clinical outcomes would be related to stroke, heart attack and angina. Thromboxane A<sub>2</sub> has also been implicated in hypertension occurring in pregnancy and TP antagonists (/TXS inhibitors) have been proposed as treatments for pre-eclampsia (Keith *et al.*, 1993; Dogné *et al.*, 2006).

In addition to the vasculature, TXA<sub>2</sub> potently stimulates other smooth muscles to contract. Thus, TP antagonists have been proposed as potential therapeutic modalities for asthma. Seratrodast (Figure 8) has received marketing approval for treatment of asthma in Japan (see Rolin *et al.*, 2006). Ramatroban and seratrodast have also been evaluated in Phase III clinical trials in the USA (Dogné *et al.*, 2002). The therapeutic rationale is to ameliorate the marked bronchoconstriction produced by TXA<sub>2</sub> and its involvement in bronchial hyper-responsiveness. However, TXA<sub>2</sub> and other prostanoids that activate TP receptors (e.g. PGD<sub>2</sub>) are not the only powerful bronchoconstrictors generated in asthma and it is unlikely that a TP antagonist alone would be adequate therapy in most patients. In this context, SQ-29548 and the cys-LT antagonist montelukast acting alone inconsistently inhibited contrac-

tion of human lung slices in culture challenged with antigen; a combination of the antagonists was much more effective, while an H<sub>1</sub> antagonist was ineffective (Wohlsen *et al.*, 2003). A combined TP/cys-LT antagonist (e.g. YM-158) may have greater clinical efficacy.

Activation of EP<sub>3</sub>, FP, TP and possibly EP<sub>1</sub> receptors causes contraction of the pregnant human myometrium, while DP<sub>1</sub>, EP<sub>2</sub> and IP receptors mediate relaxation (Senior *et al.*, 1993). Given this scenario, the potential of selective TP antagonists for treating labour-associated disorders, such as pre-term labour, would seem low. Recent studies on human myometrial specimens obtained at parturition provide a new perspective. It appears that there is a marked alteration in prostanoid receptor functional dynamics at term, prior to and following the onset of labour (Fischer *et al.*, 2008). Comparing the effects of U-46619, PGE<sub>2</sub> and PGF<sub>2α</sub>, a loss of FP but not TP responsiveness was apparent following the onset of labour (Fischer *et al.*, 2008). As PGE<sub>2</sub> produces a net inhibition of myogenic activity, this leaves TXA<sub>2</sub> as the only COX-derived product capable of exerting a contractile effect during labour. The role of TXA<sub>2</sub> in human parturition may be greater than previously envisaged.

The involvement of TXA<sub>2</sub> in inflammatory bowel disease may be significant (Rampton and Collins, 1993). The aetiology of ulcerative colitis and Crohn's disease is not fully understood, but initiating factors in pre-disposed individuals elicit severe and prolonged inflammation of the gut mucosa. Early studies with ridogrel and picotamide, combined TP antagonist/TXS inhibitors, claimed some clinical improvement associated with reduced TXA<sub>2</sub> (TXB<sub>2</sub>) release (Rampton and Collins, 1993). Given the multifactorial nature of these diseases and the number of other eicosanoids purported to be involved, pronounced beneficial effects of TP antagonists alone seem unlikely.

Evidence is available that activation of the TP receptor is implicated in chronic persistent cough (CPC). This is a detrimental and debilitating condition that serves no obvious function (French *et al.*, 2002), afflicts 9–33% of the population in Europe and North America and for which no satisfactory treatment is currently available (see Chung and Pavord, 2008). Thus, the mechanisms underlying CPC and the identification of new anti-tussive agents present a grossly neglected and unmet clinical need.

In humans, PGs have high tussive potency suggesting that they may be released locally in a variety of respiratory diseases where cough is a characteristic symptom. In subjects with asthma, both indomethacin and ozagrel (OKY-046; a TXS inhibitor) increase the threshold for cough when compared with placebo, suggesting that TXA<sub>2</sub> may be one of the cyclooxygenase products that sensitize the cough reflex (Fujimura *et al.*, 1995). Indomethacin and ozagrel also attenuate cough in subjects with hypertension treated with angiotensin-converting enzyme inhibitors (Fogari *et al.*, 1992; Malini *et al.*, 1997; Umemura *et al.*, 1997). Thus, the potential application for TP antagonists in alleviating CPC is clear. Indeed, in an animal model of asthma-related cough, the TP antagonist, seratrodast (Figure 8) significantly suppressed cough induced by capsaicin (Xiang *et al.*, 2002). Moreover, in 16 patients with stable chronic bronchitis, seratrodast (80 mg b.i.d. for 28 days) significantly increased the threshold for capsaicin-

induced cough (Ishiura *et al.*, 2003). Finally, ramatroban is reported to attenuate cough in subjects with cough variant asthma (Kitamura *et al.*, 2003), which could be due to blockade of both TP and DP<sub>2</sub> receptors (Gardiner and Browne, 1984).

A role for TXA<sub>2</sub> in cancer of the female breast and genital tract has also been suggested. Findings are reminiscent of the TXA<sub>2</sub>/PGI<sub>2</sub> balance in the cardiovascular system: TXA<sub>2</sub> promotes tumour growth and metastasis, while PGI<sub>2</sub> exerts a protective effect by maintaining vascular and platelet homeostasis (Honn and Meyer, 1981; Nigam *et al.*, 1992). A significant role in ovarian, cervical and other gynaecological cancers may exist (Nigam *et al.*, 1992), but there remains insufficient evidence to assign a pivotal role.

Thromboxanes have also been implicated in a number of other conditions, including glomerulonephritis (Patrono, 1990; Wardle, 1999), allergic conjunctivitis (Woodward *et al.*, 1990b), allergic rhinitis (Misra, 1994), diabetes (Dogné *et al.*, 2006), septic shock and pulmonary embolism (Ghuysen *et al.*, 2005). In summary, while numerous potential uses have been proposed over several years, clinical and economic success has been modest to date despite the availability of extremely potent and long-acting TP antagonists. What are the future therapeutic prospects? An argument could be made for superiority over low-dose aspirin. In order to establish this, extensive clinical trials would be essential. The bigger concern for the pharmaceutical industry would be 'how much better would a TP antagonist perform compared with low-dose aspirin?' If the difference is small, economics will dictate the fate of excellent TP antagonists. Perhaps the brightest new star in the galaxy of therapeutic options is the treatment of pre-term labour, given that TXA<sub>2</sub> is now known to be the major prostanoid involved in parturition. The only potential side effect that may occur with TP antagonist therapy is possible enhancement of hypersensitivity reactions (Narumiya, 2003). This would not be a major burden under most circumstances.

### Therapeutic future of prostanoid receptor antagonists

The future for selective prostanoid antagonists does not appear to provide an encouraging scenario. Potent and selective antagonists for every prostanoid receptor are now available, with the exception of the EP<sub>2</sub> subtype. Some selective antagonists have received regulatory approval as drugs but these instances tend to be few and not worldwide. In this category are DP<sub>2</sub> and TP antagonists. Drug approval filings/development have been discontinued on numerous TP antagonists, EP<sub>1</sub> antagonists and, most recently, an EP<sub>4</sub> antagonist. Arguments in favour of more potent and irreversible antagonists have been put forward but, in the global scheme of drug research, the current drugs seem adequate at the very least.

DP<sub>2</sub> antagonists apart, of those drugs designed as selective for a single prostanoid receptor, the TP blockers still appear to be the best prospect. This would require revisiting cardiovascular studies and daring to directly compare with low-dose

aspirin. The use of the TP antagonists for treating pre-term labour and even perhaps primary and secondary dysmenorrhoea should be contemplated in the light recent evidence (Fischer *et al.*, 2008). This is, however, a rather narrow spectrum of utility and does not fulfil three decades of high expectation.

Straightforward consideration of the current situation does not favour the widespread therapeutic utility of highly selective prostanoid receptor antagonists. Stated simply, where COX inhibitors are clinically effective, selective antagonists are not. This is not really astonishing. There are five major prostanoids biosynthesized by COX and additional active metabolites (e.g. 13,14-dihydro-15-oxo PGD<sub>2</sub>, 19(R)-OH PGE<sub>2</sub>). The isoprostanes are formed by non-enzymatic oxidation of arachidonic acid (Morrow *et al.*, 1990; 1994). Finally, there are the PG-ethanolamides (prostamides) and PG-glycerol esters, which are COX-2 products of the endocannabinoids anandamide and 2-arachidonyl glycerol. Expectations that one prostanoid and one dedicated receptor play a dominant, all-important role in most disease processes are lofty. It is unlikely that this all distils down to one important receptor activity in most diseases: a receptor widely controlling synergy or solely mediating a critical pathophysiological event.

Individual PGs and their receptors may play compensatory, fail-safe roles. Individual prostanoids may act sequentially to initiate and sustain disease states. They may subserve complementary roles. The role of DP<sub>1</sub> and DP<sub>2</sub> receptors in allergy, notably allergic rhinitis, provides an excellent example of two receptors behaving in a complementary manner to initiate and maintain the disease state. The combination of DP<sub>1</sub>/DP<sub>2</sub> antagonism in a single molecule appears a very promising therapeutic approach (Pettipher, 2008) and one that appears feasible. Equally so, combining TP and EP<sub>3</sub> antagonistic properties to prevent both platelet activation and vasoconstriction in cardiovascular disease states appears to be an achievable goal. Prostanoid-based drugs for pre-term labour may require even more versatility, with ideal therapy perhaps embodying the following attributes in a single molecule: (i) TP antagonism, (ii) EP<sub>3</sub> antagonism to block cervical ripening and (iii) EP<sub>2</sub> agonism to provide a tocolytic effect. Such a molecule would be a formidable medicinal chemistry challenge. Further challenges relate to diseases where COX inhibitors are widely and successfully used, because the ideal spectrum of antagonist properties may not be entirely clear. If the promise of prostanoid-based therapeutics is ever to be fulfilled, the role of prostaglandins may need to be carefully thought-out on a disease-by-disease basis.

### Conflicts of interest

The authors affirm no conflicts of interest.

### References

- Abramovitz M, Boie Y, Nguyen T, Rushmore TH, Bayne MA, Metters KM *et al.* (1994). Cloning and expression of a cDNA for the human prostanoid FP receptor. *J Biol Chem* **269**: 2632–2636.

- Abramovitz M, Adam M, Boie Y, Carrière M, Denis D, Godbout C *et al.* (2000). The utilization of recombinant prostanoid receptors to determine the affinities and selectivities of prostaglandins and related analogs. *Biochim Biophys Acta* **1483**: 285–293.
- Ackerley N, Brewster AG, Brown GR, Clarke DS, Foubister AJ, Griffin SJ *et al.* (1995). A novel approach to dual-acting thromboxane receptor antagonist/synthase inhibitors based on the link of 1,3-dioxane-thromboxane receptor antagonists and thromboxane synthase inhibitors. *J Med Chem* **38**: 1608–1628.
- Ahn CH, Wallace LJ, Miller DD, Feller DR (1988). Use of [<sup>3</sup>H]trimetoquinol as a radioligand in human platelets: interaction with putative endoperoxide/thromboxane A<sub>2</sub> receptor sites. *Thromb Res* **50**: 387–399.
- Alexander SPH, Mathie A, Peters JA (2008). Guide to Receptors and Channels (GRAC), 3rd edn. *Br J Pharmacol* **153** (Suppl. 2): S1–S209.
- Almishri W, Cossette C, Rokach J, Martin JG, Hamid Q, Powell WS (2005). Effects of prostaglandin D<sub>2</sub>, 15-deoxy-Δ<sup>12,14</sup> prostaglandin J<sub>2</sub>, and selective DP<sub>1</sub> and DP<sub>2</sub> receptor agonists on pulmonary infiltration of eosinophils in Brown Norway rats. *J Pharmacol Exp Ther* **313**: 64–69.
- Al Sinawi LA, Mekki QA, Hassan S, Hedges A, Burke C, Moody SG *et al.* (1985). Effect of a hydantoin prostaglandin analogue, BW 245C, during oral dosing in man. *Prostaglandins* **29**: 99–111.
- Amano H, Hayashi I, Endo H, Kitasato H, Yamashina S, Maruyama T *et al.* (2003). Host prostaglandin E<sub>2</sub>-EP<sub>3</sub> signaling regulates tumor-associated angiogenesis and tumor growth. *J Exp Med* **197**: 221–232.
- Angulo J, Cuevas P, La Fuente JM, Pomerol JM, Ruiz-Castañé E, Puigvert A *et al.* (2002). Regulation of human penile smooth muscle tone by prostanoid receptors. *Br J Pharmacol* **136**: 23–30.
- Arikawa T, Omura K, Morita I (2004). Regulation of bone morphogenetic protein-2 expression by endogenous prostaglandin E<sub>2</sub> in human mesenchymal stem cells. *J Cell Physiol* **200**: 400–406.
- Arakida Y, Suwa K, Ohga K, Yokota M, Miyata K, Yamada T *et al.* (1998). *In vitro* pharmacologic profile of YM158, a new dual antagonist for LTD<sub>4</sub> and TXA<sub>2</sub> receptors. *J Pharmacol Exp Ther* **287**: 633–639.
- Aronoff DM, Canetti C, Peters-Golden M (2004). Prostaglandin E<sub>2</sub> inhibits alveolar macrophage phagocytosis through an E-prostanoid 2 receptor-mediated increase in intracellular cyclic AMP. *J Immunol* **173**: 559–565.
- Arimura A, Yasui K, Kishino J, Asanuma F, Hasegawa H, Kakudo S *et al.* (2001). Prevention of allergic inflammation by a novel prostaglandin receptor antagonist, S-5751. *J Pharmacol Exp Ther* **298**: 411–419.
- Arimura A, Kishino J, Tanimoto N (2003). PGD<sub>2</sub> receptor antagonist. WO2003/03097042, Shionogi & Co.
- Arner RE, Ashton MR, Boyd EA, Brennan CJ, Brookfield FA, Gazi L *et al.* (2005). Indole-3-acetic acid antagonists of the prostaglandin D<sub>2</sub> receptor CRTH2. *J Med Chem* **48**: 6174–6177.
- Arner RE, Boyd AE, Hay PA (2006). 1-Acetic acid-indole derivatives with PGD<sub>2</sub> antagonist activity. WO2006/095183, Oxagen Ltd.
- Armour CL, Johnson PR, Alfredson ML, Black JL (1989). Characterization of contractile prostanoid receptors on human airway smooth muscle. *Eur J Pharmacol* **165**: 215–222.
- Armstrong RA, Jones RL, Wilson NH (1983). Ligand binding to thromboxane receptors on human platelets: correlation with biological activity. *Br J Pharmacol* **79**: 953–964.
- Armstrong RA, Jones RL, Pessapati V, Will SG, Wilson NH (1985). Competitive antagonism at thromboxane receptors in human platelets. *Br J Pharmacol* **84**: 595–607.
- Armstrong RA, Jones RL, MacDermot J, Wilson NH (1986). Prostaglandin endoperoxide analogues which are both thromboxane receptor antagonists and prostacyclin mimetics. *Br J Pharmacol* **87**: 543–551.
- Armstrong RA, Lawrence RA, Jones RL, Wilson NH, Collie A (1989). Functional and ligand binding studies suggest heterogeneity of platelet prostacyclin receptors. *Br J Pharmacol* **97**: 657–658.
- Armstrong RA, Humphrey PPA, Lumley P (1993). Characteristics of the binding of [<sup>3</sup>H]-GR32191 to the thromboxane (TP-) receptor of human platelets. *Br J Pharmacol* **110**: 539–547.
- Arner M, Högestätt ED, Uski TK (1991). Characterization of contraction-mediating prostanoid receptors in human hand veins: effects of the thromboxane receptor antagonists BM13,505 and AH23848. *Acta Physiol Scand* **141**: 79–86.
- Ashida Y, Matsumoto T, Kuriki H, Shiraishi M, Kato K, Terao S (1989). A novel anti-asthmatic quinone derivative, AA-2414 with a potent antagonistic activity against a variety of spasmogenic prostanoids. *Prostaglandins* **38**: 91–112.
- Audoly LP, Tilley SA, Goulet J, Key M, Nguyen H, Stock JL *et al.* (1999). Identification of specific EP receptors responsible for the hemodynamic effects of PGE<sub>2</sub>. *Am J Physiol* **277**: H924–H930.
- Awad MMA, Bazin M, Feru F, Goldstein SW, Kuhn CF (2004). Tetrahydroquinoline derivatives as CRTH2 antagonists. WO2004/035543, Warner-Lambert & Co.
- Ayer LM, Wilson SM, Traves SL, Proud D, Giembycz MA (2008). 4,5-Dihydro-1H-imidazol-2-yl)-[4-(4-isopropoxy-benzyl)-phenyl]-amine (RO1138452) is a selective, pseudo-irreversible orthosteric antagonist at the prostacyclin (IP)-receptor expressed by human airway epithelial cells: IP-receptor-mediated inhibition of CXCL9 and CXCL10 release. *J Pharmacol Exp Ther* **324**: 815–826.
- Barbieri EJ, Orzechowski RF, Rossi GV (1977). Measurement of prostaglandin E<sub>2</sub> in an inflammatory exudate: effects of nonsteroidal anti-inflammatory agents. *J Pharmacol Exp Ther* **201**: 769–777.
- Bastien L, Sawyer N, Grygorczyk R, Metters KM, Adam M (1994). Cloning, functional expression, and characterization of the human prostaglandin E<sub>2</sub> receptor EP<sub>2</sub> subtype. *J Biol Chem* **269**: 11873–11877.
- Bauer PH, Cheng JB, Gladue RP, Li B, Neote KS, Zhang J (2002). Methods for the identification of compounds useful for the treatment of disease states mediated by prostaglandin D<sub>2</sub>. EP1170594, Pfizer Products Inc.
- Baxter GS, Clayton JK, Coleman RA, Marshall K, Sangha R, Senior J (1995). Characterization of the prostanoid receptors mediating constriction and relaxation of human isolated uterine artery. *Br J Pharmacol* **116**: 1692–1696.
- Baxter A, Steele J, Teague S (2003a). Use of indole-3-acetic acids in the treatment of asthma, COPD and other diseases. WO2003/066046, AstraZeneca AB.
- Baxter A, Steele J, Teague S (2003b). Use of indole-3-acetic acids in the treatment of asthma, COPD and other diseases. WO2003/066047, AstraZeneca AB.
- Beasley RC, Featherstone RL, Church MK, Rafferty P, Varley JG, Harris A *et al.* (1989). Effect of a thromboxane receptor antagonist on PGD<sub>2</sub>- and allergen-induced bronchoconstriction. *J Appl Physiol* **66**: 1685–1693.
- Beaulieu C, Guay D, Wang Z, Leblanc Y, Roy P, Dufresne C *et al.* (2008). Identification of prostaglandin D<sub>2</sub> receptor antagonists based on a tetrahydropyridindole scaffold. *Bioorg Med Chem Lett* **18**: 2696–2700.
- Bellely M, Gallant M, Roy B, Houde K, Lachance N, Labelle M *et al.* (2005). Structure-activity relationship studies on *ortho*-substituted cinnamic acids, a new class of selective EP<sub>3</sub> antagonists. *Bioorg Med Chem Lett* **15**: 527–530.
- Bennani YL, Tumey LN, Gleason EA, Robarge MJ (2006). Indole acetic acids exhibiting CRTh2 receptor antagonism and uses thereof. US2006/0100425, Atherts Inc.
- Bennett A, Posner J (1971). Studies on prostaglandin antagonists. *Br J Pharmacol* **42**: 584–594.
- Berthelette C, Lachance N, Li L, Sturino CF, Wang Z (2003). Fluoro substituted cycloalkanoindoles, compositions containing such compounds and methods of treatment. WO2003/062200, Merck Frosst Canada & Co.
- Bexis S, McCormick PA, Docherty JR (2008). Vascular actions of the prostacyclin receptor antagonist BAY 73-1449 in the portal hypertensive rat. *Eur J Pharmacol* **590**: 322–326.

- Bhagwat SS, Gude C, Cohen DS, Dotson R, Mathis J, Lee W *et al.* (1993). Thromboxane receptor antagonism combined with thromboxane synthase inhibition. 5. Synthesis and evaluation of enantiomers of 8-[[4-(4-chlorophenyl)sulfonyl]amino]-4-(3-pyridinylalkyl) octanoic acid. *J Med Chem* **36**: 205–210.
- Billot X, Chateauneuf A, Chauret N, Denis D, Greig G, Mathieu M-C *et al.* (2003). Discovery of a potent and selective agonist of the prostaglandin EP<sub>4</sub> receptor. *Bioorg Med Chem Lett* **13**: 1129–1132.
- Birkinshaw TN, Bonnert R, Cook A, Rasul R, Sanganee H, Teague S (2003). Novel substituted indoles. WO2003/03101981, AstraZeneca AB.
- Birkinshaw TN, Teague SJ, Beech C, Bonnert RV, Hill S, Patel A *et al.* (2006). Discovery of potent CRTh2 (DP<sub>2</sub>) receptor antagonists. *Bioorg Med Chem Lett* **16**: 4287–4290.
- Bley KR, Hunter JC, Eglen RM, Smith JA (1998). The role of IP prostanoid receptors in inflammatory pain. *Trends Pharmacol Sci* **19**: 141–147.
- Bley KR, Bhattacharya A, Daniels DV, Gever J, Jahangir A, O'Yang C *et al.* (2006). RO1138452 and RO3244794: characterization of structurally distinct, potent and selective IP (prostacyclin) receptor antagonists. *Br J Pharmacol* **147**: 335–345.
- Boersma JI, Janzen KM, Oliveira L, Crankshaw DJ (1999). Characterization of excitatory prostanoid receptors in the human umbilical artery in vitro. *Br J Pharmacol* **128**: 1505–1512.
- Boie Y, Sawyer N, Slipetz DM, Metters KM, Abramovitz M (1995). Molecular cloning and characterization of the human prostanoid DP receptor. *J Biol Chem* **270**: 18910–18916.
- Boie Y, Stocco R, Sawyer N, Slipetz DM, Ungrin MD, Neuschafer-Rube F *et al.* (1997). Molecular cloning and characterization of the four rat prostaglandin E<sub>2</sub> prostanoid receptor subtypes. *Eur J Pharmacol* **340**: 227–241.
- Bonnert R, Rasul R (2004). Novel substituted 3-sulfur indoles. WO2004/106302, AstraZeneca AB.
- Bonnert R, Brough S, Cook T, Dickinson M, Rasul R, Sanganee H *et al.* (2003). Novel substituted indoles. WO2003/101961, AstraZeneca AB.
- Bonnert R, Dickinson M, Rasul R, Sanganee H, Teague S (2004). Indole-3-sulphur derivatives. WO2004/007451, AstraZeneca AB.
- Bonnert R, Cook AR, Luker TJ, Mohammed RS, Thom S (2005a). Substituted indole derivatives for pharmaceutical composition for treating respiratory diseases. WO2005/019171, AstraZeneca AB.
- Bonnert R, Mohammed RT, Teague S (2005b). 1-Acetic acid-indole, indazole and benzimidazole derivatives useful for the treatment of respiratory disorders. WO2005/054232, AstraZeneca AB.
- Bonnert RV, Patel A, Thom S (2005c). Phenoxyacetic acid derivatives. WO2005/018529A3, AstraZeneca AB.
- Bouayad A, Kajino H, Waleh N, Fouron J-C, Andelfinger G, Varma DR *et al.* (2001). Characterization of PGE<sub>2</sub> receptors in fetal and newborn lamb ductus arteriosus. *Am J Physiol* **280**: H2342–H2349.
- Brescia MR, Rokosz LL, Cole AG, Stauffer TM, Lehrach JM, Auld DS (2007). Discovery and preliminary evaluation of 5-(4-phenylbenzyl)oxazole-4-carboxamides as prostacyclin receptor antagonists. *Bioorg Med Chem Lett* **17**: 1211–1215.
- Brewster AG, Brown GR, Foubister AJ, Jessup R, Smithers MJ (1988). The synthesis of a novel thromboxane receptor antagonist 4(Z)-6-(2-O-chlorophenyl-4-O-hydroxyphenyl-1,3-dioxan-cis-5-yl) hexanoic acid, ICI 192605. *Prostaglandins* **36**: 173–178.
- Breyer MD, Breyer RM (2000). Prostaglandin receptors: their role in regulating renal function. *Curr Opin Nephrol Hypertens* **9**: 23–29.
- Brownlie RP, Brownrigg NJ, Butcher HM, Garcia R, Jessup R, Lee VJ *et al.* (1993). ZD1542, a potent thromboxane A<sub>2</sub> synthase inhibitor and receptor antagonist in vitro. *Br J Pharmacol* **110**: 1600–1606.
- Bunce KT, Clayton NM, Coleman RA, Collington EW, Finch H, Humphray JM *et al.* (1990). GR-63799X – a novel prostanoid with selectivity for EP<sub>3</sub> receptors. *Adv Prost Throm Leuk Res* **21A**: 379–382.
- Burch JD, Belley M, Fortin R, Deschênes D, Girard M, Colucci J *et al.* (2008). Structure-activity relationships and pharmacokinetic parameters of quinoline acylsulfonamides as potent and selective antagonists of the EP<sub>4</sub> receptor. *Bioorg Med Chem Lett* **18**: 2048–2054.
- Campbell IB, Collington EW, Finch H, Hayes R, Lumley P, Mills K *et al.* (1991a). Synthesis and pharmacological evaluation of combined thromboxane receptor antagonist/synthase inhibitors: pyridine-containing amino-prostanoids. *Bioorg Med Chem Lett* **1**: 695–698.
- Campbell IB, Collington EW, Finch H, Hayes R, Lumley P, Mills K *et al.* (1991b). Synthesis and pharmacological evaluation of combined thromboxane receptor antagonist/synthase inhibitors: pyridine-containing sulfonamido acids. *Bioorg Med Chem Lett* **1**: 699–704.
- Camras CB, Toris CB, Sjoquist B, Milleson M, Thorngren JO, Hejkal TW *et al.* (2004). Detection of the free acid of bimatoprost in aqueous humor samples from human eyes treated with bimatoprost before cataract surgery. *Ophthalmology* **111**: 2193–2198.
- Camu F, Shi L, Vanlersberghe C (2003). The role of COX-2 inhibitors in pain modulation. *Drugs* **63** (Suppl. 1): 1–7.
- Cefalu JS, Zhu QM, Eggers AC, Kaan TK, Ho MJ, Jett MF *et al.* (2007). Effects of the selective prostacyclin receptor antagonist RO3244019 on the micturition reflex in rats. *J Urol* **178**: 2683–2686.
- Challis JR, Sloboda DM, Alfaidy N, Lye SJ, Gibb W, Patel FA *et al.* (2002). Prostaglandins and mechanisms of preterm birth. *Reproduction* **124**: 1–17.
- Chan CL, Jones RL, Lau HY (2000). Characterization of prostanoid receptors mediating inhibition of histamine release from anti-IgE-activated rat peritoneal mast cells. *Br J Pharmacol* **129**: 589–597.
- Chan KM, Jones RL (2004). Partial agonism of taprostene at prostanoid IP receptors in vascular preparations from guinea-pig, rat, and mouse. *J Cardiovasc Pharmacol* **43**: 795–807.
- Chang SW, Reddy V, Pereira T, Dean BJ, Xia YQ, Seto C *et al.* (2007). The pharmacokinetics and disposition of MK-0524, a prostaglandin D<sub>2</sub> receptor 1 antagonist, in rats, dogs and monkeys. *Xenobiotica* **37**: 514–533.
- Charlesworth EN, Kagey-Sobotka A, Schleimer RP, Norman PS, Lichtenstein LM (1991). Prednisone inhibits the appearance of inflammatory mediators and the influx of eosinophils and basophils associated with the cutaneous late-phase response to allergen. *J Immunol* **146**: 671–676.
- Chemtob S, Peri AG (2006). Peptide antagonists of prostaglandin F<sub>2α</sub> receptor. US2006/6984719, Hospital Sainte-Justine, Montreal.
- Chen J, Woodward DF, Coleman RA, Jones RL, Lydford SJ (2001). Prostanoid receptor assays. In: Enna SJ, Williams M, Ferkany T, Kenakin T, Persolt RD, Sullivan JP (eds). *Current Protocols in Pharmacology*. John Wiley & Sons: New York, pp. 4.18.1–4.18.41.
- Chen J, Senior J, Marshall K, Abbas F, Dinh H, Dinh T *et al.* (2005). Studies using isolated uterine and other preparations show bimatoprost and prostanoid FP agonists have different activity profiles. *Br J Pharmacol* **144**: 493–501.
- Cheng K, Wu TJ, Wu KK, Sturino C, Metters K, Gottesdiener K *et al.* (2006). Antagonism of the prostaglandin D<sub>2</sub> receptor 1 suppresses nicotinic acid-induced vasodilation in mice and humans. *Proc Natl Acad Sci USA* **103**: 6682–6687.
- Cherukuri DP, Chen XB, Goulet AC, Young RN, Han Y, Heimark RL *et al.* (2007). The EP<sub>4</sub> receptor antagonist, L-161,982, blocks prostaglandin E<sub>2</sub>-induced signal transduction and cell proliferation in HCA-7 colon cancer cells. *Exp Cell Res* **313**: 2969–2979.
- Chollet A, Tos EG, Cirillo R (2007). Tocolytic effect of a selective FP receptor antagonist in rodent models reveals an innovative approach to the treatment of preterm labor. *BMC Pregnancy Childbirth* **7** (Suppl. 1): S16 (open access).
- Chow KB, Wong YH, Wise H (2001). Prostacyclin receptor-independent inhibition of phospholipase C activity by non-prostanoid prostacyclin mimetics. *Br J Pharmacol* **134**: 1375–1384.
- Chung KF, Pavord ID (2008). Prevalence, pathogenesis, and causes of chronic cough. *Lancet* **371**: 1364–1374.

- Cimetièrè B, Dubuffet T, Muller O, Descombes JJ, Simonet S, Laubie M *et al.* (1998). Synthesis and biological evaluation of new tetrahydronaphthalene derivatives as thromboxane receptor antagonists. *Bioorg Med Chem Lett* **8**: 1375–1380.
- Cirillo R, Tos EG, Page P, Missotten M, Quattropiani A, Scheer A *et al.* (2007). Arrest of preterm labor in rat and mouse by an oral and selective nonprostanoid antagonist of the prostaglandin F<sub>2α</sub> receptor (FP). *Am J Obstet Gynecol* **197**(54): e1–e9.
- Clark P, Rowland SE, Denis D, Mathieu MC, Stocco R, Poirier H *et al.* (2008). MF498 [N-[[4-(5,9-Diethoxy-6-oxo-6,8-dihydro-7H-pyrrolo [3,4-g]quinolin-7-yl)-3-methylbenzyl]sulfonyl]-2-(2-methoxyphenyl)acetamide], a selective E prostanoid receptor 4 antagonist, relieves joint inflammation and pain in rodent models of rheumatoid and osteoarthritis. *J Pharmacol Exp Ther* **325**: 425–434.
- Clark RD, Jahangir A, Severance D, Salazar R, Chang T, Chang D *et al.* (2004). Discovery and SAR development of 2-(phenylamino) imidazolines as prostacyclin receptor antagonists. *Bioorg Med Chem Lett* **14**: 1053–1056.
- Clarke DL, Giembycz MA, Patel HJ, Belvisi MG (2004). E-ring 8-isoprostanes inhibit ACh release from parasympathetic nerves innervating guinea-pig trachea through agonism of prostanoid receptors of the EP<sub>3</sub>-subtype. *Br J Pharmacol* **141**: 600–609.
- Cocks TM, King SJ, Angus JA (1990). Glibenclamide is a competitive antagonist of the thromboxane A<sub>2</sub> receptor in dog coronary artery *in vitro*. *Br J Pharmacol* **100**: 375–378.
- Coleman RA, Sheldrick RL (1989). Prostanoid-induced contraction of human bronchial smooth muscle is mediated by TP-receptors. *Br J Pharmacol* **96**: 688–692.
- Coleman RA, Humphrey PPA, Kennedy I, Lumley P (1984). Prostanoid receptors: the development of a working classification. *Trends Pharmacol Sci* **5**: 303–306.
- Coleman RA, Kennedy I, Sheldrick RLG (1987). New evidence with selective agonists and antagonists for the subclassification of PGE<sub>2</sub>-sensitive (EP) receptors. *Adv Prost Thromb Leukot Res* **17**: 467–470.
- Coleman RA, Grix SP, Head SA, Louttit JB, Mallett A, Sheldrick RL (1994a). A novel inhibitory prostanoid receptor in piglet saphenous vein. *Prostaglandins Leukot Essent Fatty Acids* **47**: 151–168.
- Coleman RA, Smith WL, Narumiya S (1994b). VIII International union of pharmacology classification of prostanoid receptors: properties distribution, and structure of the receptors and their subtypes. *Pharmacol Rev* **46**: 205–229.
- Colquhoun D (2007). Why the Schild method is better than Schild realised. *Trends Pharmacol Sci* **28**: 608–614.
- Corradini L, Field MJ, Kinloch RA, Williams-Jones BI (2005). Methods of treating neuropathic pain using a CRTH2 receptor antagonist. WO2005/102338, Warner-Lambert & Co.
- Cozzi P, Giordani A, Menichincheri M, Pillan A, Pinciroli V, Rossi A *et al.* (1994). Agents combining thromboxane receptor antagonism with thromboxane synthase inhibition: [[2-(1H-imidazol-1-yl)ethylidene]amino]oxy]alkanoic acids. *J Med Chem* **37**: 3588–3604.
- Craig DA (1993). The Cheng-Prusoff relationship: something lost in the translation. *Trends Pharmacol Sci* **14**: 89–91.
- Crider JY, Griffin BW, Sharif NA (1999). Prostaglandin DP receptors positively coupled to adenylyl cyclase in embryonic bovine tracheal (EBTr) cells: pharmacological characterization using agonists and antagonists. *Br J Pharmacol* **127**: 204–210.
- Daray FM, Minvielle AI, Puppo S, Rothlin RP (2003). Pharmacological characterization of prostanoid receptors mediating vasoconstriction in human umbilical vein. *Br J Pharmacol* **139**: 1409–1426.
- Davis RJ, Murdoch CE, Ali M, Purbrick S, Ravid R, Baxter GS *et al.* (2004). EP<sub>4</sub> prostanoid receptor-mediated vasodilatation of human middle cerebral arteries. *Br J Pharmacol* **141**: 580–585.
- Davis-Bruno KL, Halushka PV (1994). Molecular pharmacology and therapeutic potential of thromboxane A<sub>2</sub> receptor antagonists. *Adv Drug Res* **25**: 173–202.
- Delaey C, Van de Voorde J (1995). Prostanoid-induced contractions are blocked by sulfonylureas. *Eur J Pharmacol* **280**: 179–184.
- Dogné J-M, Rolin S, de Leval X, Benoit P, Neven P, Delarge J *et al.* (2001). Pharmacology of the thromboxane receptor antagonist and thromboxane synthase inhibitor BM-531. *Cardiovasc Drug Rev* **19**: 87–96.
- Dogné J-M, de Leval X, Benoit P, Rolin S, Pirotte B, Masereel B (2002). Therapeutic potential of thromboxane inhibitors in asthma. *Expert Opin Investig Drug* **11**: 275–281.
- Dogné J-M, Hanson J, Pratico D (2005). Thromboxane, prostacyclin and isoprostanes: therapeutic targets in atherogenesis. *Trends Pharmacol Sci* **26**: 639–644.
- Dogné J-M, Hanson J, de Leval X, Pratico D, Pace-Asciak CR, Drion P *et al.* (2006). From the design to the clinical application of thromboxane modulators. *Curr Pharm Des* **12**: 903–923.
- Doheny HC, O'Reilly MJ, Sexton DJ, Morrison JJ (2007). THG113.31, a specific PGF<sub>2α</sub> receptor antagonist, induces human myometrial relaxation and BKCa channel activation. *Reprod Biol Endocrinol* **5**: 10 (open access).
- Dong YJ, Jones RL, Wilson NH (1986). Prostaglandin E receptor subtypes in smooth muscle: agonist activities of stable prostacyclin analogues. *Br J Pharmacol* **87**: 97–107.
- Dorn GW (1989). Distinct platelet thromboxane A<sub>2</sub>/prostaglandin H<sub>2</sub> receptor subtypes. A radioligand binding study of human platelets. *J Clin Invest* **84**: 1883–1891.
- Dorn GW, Davis MG, D'Angelo DD (1997). Structural determinants for agonist binding affinity to thromboxane/prostaglandin endoperoxide (TP) receptors. Analysis of chimeric rat/human TP receptors. *J Biol Chem* **272**: 12399–12405.
- Drower EJ, Stapelfeld A, Mueller RA, Hammond DL (1987). The antinociceptive effects of prostaglandin antagonists in the rat. *Eur J Pharmacol* **133**: 249–256.
- Dubé GP, Mais DE, Jakubowski JA, Brune KA, Utterback BG, True TA *et al.* (1992). *In vitro* characterization of a novel TXA<sub>2</sub>/PGH<sub>2</sub> receptor ligand (S-145) in platelets and vascular and airway smooth muscle. *J Pharmacol Exp Ther* **262**: 784–791.
- Ducharme Y, Blouin M, Carrière MC, Chateaufneuf A, Côté B, Denis D *et al.* (2005). 2,3-Diarylthiophenes as selective EP<sub>1</sub>-receptor antagonists. *Bioorg Med Chem Lett* **15**: 1155–1160.
- Durocher Y, Perret S, Thibaudeau E, Gaumont MH, Kamen A, Stocco R *et al.* (2000). A reporter gene assay for high-throughput screening of G-protein-coupled receptors stably or transiently expressed in HEK293 EBNA cells grown in suspension culture. *Anal Biochem* **284**: 316–326.
- Eglen RM, Whiting RL (1988). The action of prostanoid receptor agonists and antagonists on smooth muscle and platelets. *Br J Pharmacol* **94**: 591–601.
- Faith A, McDonald J, Peek E, Richards D, Caulfield J, Chevretton E *et al.* (2005). Functional plasticity of human respiratory tract dendritic cells: GM-CSF enhances Th2 development. *J Allergy Clin Immunol* **116**: 1136–1143.
- Farmer JB, Farrar DG, Wilson J (1974). Antagonism of tone and prostaglandin-mediated responses in a tracheal preparation by indomethacin and SC-19220. *Br J Pharmacol* **52**: 559–565.
- Featherstone RL, Robinson C, Holgate ST, Church MK (1990). Evidence for thromboxane receptor mediated contraction of guinea-pig and human airways *in vitro* by prostaglandin (PG) D<sub>2</sub>, 9α,11β-PGF<sub>2</sub> and PGF<sub>2α</sub>. *Nauyn Schmied Arch Pharmacol* **341**: 439–443.
- Fernandes B, Crankshaw D (1995). Functional characterization of the prostanoid DP receptor in human myometrium. *Eur J Pharmacol* **283**: 73–81.
- Fischer DP, Hutchinson JA, Farrar D, O'Donovan PJ, Woodward DF, Marshall KM (2008). Loss of prostanoid F<sub>2α</sub>, but not thromboxane, responsiveness in pregnant human myometrium during labour. *Endocrinology* **197**: 171–179.
- Fitch WL, Berry PW, Tu Y, Tabatabaei A, Lowrie L, Lopez-Tapia F *et al.*



- (2004). Identification of glutathione-derived metabolites from an IP receptor antagonist. *Drug Metab Dispos* 32: 1482–1490.
- Fitzgerald GA, Oates JA, Hawiger J, Maas RL, Roberts LJ 2nd, Lawson JA *et al.* (1983). Endogenous biosynthesis of prostacyclin and thromboxane and platelet function during chronic administration of aspirin in man. *J Clin Invest* 71: 676–688.
- Fogari R, Zoppi A, Tettamanti F, Malamani GD, Tinelli C, Salvetti A (1992). Effects of nifedipine and indomethacin on cough induced by angiotensin-converting enzyme inhibitors: a double-blind, randomized, cross-over study. *J Cardiovasc Pharmacol* 19: 670–673.
- Folger WH, Halushka PV, Wilcox CS, Guzman NJ (1992). Characterization of rat glomerular thromboxane A<sub>2</sub> receptors: comparison to rat platelets. *Eur J Pharmacol* 227: 71–78.
- Foudi N, Kotelevets L, Louedec L, Lesèche G, Henin D, Chastre E *et al.* (2008). Vasorelaxation induced by prostaglandin E<sub>2</sub> in human pulmonary vein: role of the EP<sub>4</sub> receptor subtype. *Br J Pharmacol* 154: 1631–1639.
- French CT, Irwin RS, Fletcher KE, Adams TM (2002). Evaluation of a cough-specific quality-of-life questionnaire. *Chest* 121: 1123–1131.
- Fretz H, Fecher A, Hilpert K, Riederer M (2005). Tetrahydropyridoin-dole derivatives. WO2005/095397, Actelion Pharmaceuticals.
- Friel AM, O'Reilly MW, Sexton DJ, Morrison JJ (2005). Specific PGF<sub>2α</sub> receptor (FP) antagonism and human uterine contractility *in vitro*. *BJOG Int* 112: 1034–1042.
- Fruzzetti F, Melis GB, Strigini F, Vettori C, Ricci C, Fioretti P (1988). Use of sulprostone for induction of preoperative cervical dilation or uterine evacuation, a comparison among the effects of different treatment schedules. *Obstet Gynecol* 72: 704–708.
- Fujimura M, Kamio Y, Kasahara K, Bando T, Hashimoto T, Matsuda T (1995). Prostanoids and cough response to capsaicin in asthma and chronic bronchitis. *Eur Resp J* 8: 1499–1505.
- Fujino H, Regan JW (2001). FP Prostanoid receptor activation of a T-cell factor/β-catenin signaling pathway. *J Biol Chem* 276: 12489–12492.
- Fujino H, Regan JW (2003). Prostanoid receptors and phosphatidylinositol 3-kinase: a pathway to cancer. *Trends Pharmacol Sci* 24: 335–340.
- Fujitani Y, Kanaoka Y, Aritake K, Uodome N, Okazaki-Hatake K, Urade Y (2002). Pronounced eosinophilic lung inflammation and Th2 cytokine release in human lipocalin-type prostaglandin D synthase transgenic mice. *J Immunol* 168: 443–449.
- Fulton AM, Ma X, Kundu N (2006). Targeting prostaglandin E EP receptors to inhibit metastasis. *Cancer Res* 66: 9794–9797.
- Funk CD, Furci L, Fitzgerald GA, Grygorczyk R, Rochette C, Bayne MA *et al.* (1993a). Cloning and expression of a cDNA for the human prostaglandin E receptor EP<sub>1</sub> subtype. *J Biol Chem* 268: 26767–26772.
- Funk CD, Furci L, Moran N, Fitzgerald GA (1993b). Point mutation in the seventh hydrophobic domain of the human thromboxane A<sub>2</sub> receptor allows discrimination between agonist and antagonist binding sites. *Mol Pharmacol* 44: 934–949.
- Furci L, Fitzgerald DJ, Fitzgerald GA (1991). Heterogeneity of prostaglandin H<sub>2</sub>/thromboxane A<sub>2</sub> receptors: distinct subtypes mediate vascular smooth muscle contraction and platelet aggregation. *J Pharmacol Exp Ther* 258: 74–81.
- Gallant M, Carrière MC, Chateaufneuf A, Denis D, Gareau Y, Godbout C *et al.* (2002). Structure-activity relationship of biaryl acylsulfonamide analogues on the human EP<sub>3</sub> prostanoid receptor. *Bioorg Med Chem Lett* 12: 583–586.
- Gardan B, Crakowski JL, Sessa C, Hunt M, Stanke-Labesque F, Devillier P *et al.* (2000). Vasoconstrictor effects of iso-prostaglandin F<sub>2α</sub> type-III (8-iso-prostaglandin F<sub>2α</sub>) on human saphenous veins. *J Cardiovasc Pharmacol* 35: 729–734.
- Gardiner PJ, Browne JL (1984). Tussive activity of inhaled PGD<sub>2</sub> in the cat and characterisation of the receptor(s) involved. *Prostaglandins Leukot Med* 14: 153–159.
- Ghosh S, Elder AM, Carson KG, Sprott K, Harrison S (2004). PGD<sub>2</sub> receptor antagonists for the treatment of inflammatory diseases. WO2004/032848, Millennium Pharmaceuticals.
- Ghosh S, Elder AM, Carson KG, Sprott KT, Harrison SJ, Hicks FA *et al.* (2005). PGD<sub>2</sub> receptor antagonists for the treatment of inflammatory diseases. WO2005/100321, Millennium Pharmaceuticals.
- Ghuysen A, Dogné JM, Chiap P, Rolin S, Masereel B, Lambermont B (2005). Pharmacological profile and therapeutic potential of BM-573, a combined thromboxane receptor antagonist and synthase inhibitor. *Cardiovasc Drug Rev* 23: 1–14.
- Giblin GM, Bit RA, Brown SH, Chaignot HM, Chowdhury A, Chessell IP *et al.* (2007). The discovery of 6-[2-(5-chloro-2-[(2,4-difluorophenyl)methyl]oxy)phenyl]-1-cyclopenten-1-yl]-2-pyridinecarboxylic acid, GW848687X, a potent and selective prostaglandin EP<sub>1</sub> receptor antagonist for the treatment of inflammatory pain. *Bioorg Med Chem Lett* 17: 385–389.
- Gil DW, Cheevers CV, Donello JE (2008). Transient allodynia pain models in mice for early assessment of analgesic activity. *Br J Pharmacol* 153: 769–774.
- Giles H, Leff P (1992). Classification of platelet and vascular prostaglandin D<sub>2</sub> (DP) receptors: estimation of affinities and relative efficacies for a series of novel bicyclic ligands. With an appendix on goodness-of-fit analyses. *Br J Pharmacol* 106: 996–1003.
- Giles H, Leff P, Bolofo ML, Kelly MG, Robertson AD (1989). The classification of prostaglandin DP-receptors in platelets and vasculature using BW A868C, a novel, selective and potent competitive antagonist. *Br J Pharmacol* 96: 291–300.
- Glass M, Hong J, Sato TA, Mitchell MD (2005). Misidentification of prostamides as prostaglandins. *J Lipid Res* 46: 1364–1368.
- Goldenberg RL (2002). The management of preterm labor. *Obstet Gynecol* 100: 1020–1037.
- Gomoll AW, Ogletree ML (1994). Failure of aspirin to interfere with the cardioprotective effects of ifetroban. *Eur J Pharmacol* 271: 471–479.
- Gray SJ, Giles H, Posner J (1992). The effect of a prostaglandin DP-receptor partial agonist (192C86) on platelet aggregation and the cardiovascular system in healthy volunteers. *Br J Clin Pharmacol* 34: 344–351.
- Griffin BW, Klimko P, Crider JY, Sharif NA (1999). AL-8810: a novel prostaglandin F<sub>2</sub> alpha analog with selective antagonist effects at the prostaglandin F<sub>2</sub> alpha (FP) receptor. *J Pharmacol Exp Ther* 290: 1278–1284.
- Grover GJ, Schumacher WA, Ogletree ML (1994). Thromboxane receptor antagonist BMS-180291, but not aspirin, reduces the severity of pacing-induced ischaemia in dogs. *J Cardiovasc Pharmacol* 24: 493–499.
- Guan Y, Zhang Y, Wu J, Qi Z, Yang G, Dou D *et al.* (2007). Antihypertensive effects of selective prostaglandin E<sub>2</sub> receptor subtype 1 targeting. *J Clin Invest* 117: 2496–2505.
- Habib A, Fitzgerald GA, Maclouf J (1999). Phosphorylation of the thromboxane receptor alpha, the predominant isoform expressed in human platelets. *J Biol Chem* 274: 2645–2651.
- Hall A, Atkinson S, Brown SH, Chessell IP, Chowdhury A, Giblin GM *et al.* (2007a). Discovery of novel, non-acidic 1,5-biaryl pyrrole EP<sub>1</sub> receptor antagonists. *Bioorg Med Chem Lett* 17: 4450–4455.
- Hall A, Brown SH, Chowdhury A, Giblin GM, Gibson M, Healy MP *et al.* (2007b). Identification and optimization of novel 1,3,4-oxadiazole EP<sub>1</sub> receptor antagonists. *Bioorg Med Chem Lett* 17: 1200–1205.
- Hall RA, Gillard J, Guindon Y, Letts G, Champion E, Ethier D *et al.* (1987). Pharmacology of L-655,240 (3-[1-(4-chlorobenzyl)-5-fluoro-3-methyl-indol-2-yl]2,2-dimethylpropanoic acid); a potent, selective thromboxane/prostaglandin endoperoxide antagonist. *Eur J Pharmacol* 135: 193–201.
- Hallinan EA, Hagen TJ, Husa RK, Tsymbalov S, Rao SN, vanHoeck J-P *et al.* (1993). N-substituted dibenzoxazepines as analgesic PGE<sub>2</sub> antagonists. *J Med Chem* 36: 3293–3299.
- Hallinan EA, Stapelfeld A, Savage MA, Reichman M (1994). 8-Chlorodibenz[b,f][1,4]oxazepine-10(11H)-carboxylic acid, 2-[3-[2-

- (furanylmethyl)thio]-1-oxopropyl]hydrazide (SC-51322): a potent PGE<sub>2</sub> antagonist and analgesic. *Bioorg Med Chem Lett* 4: 509–514.
- Hammad H, Lambrecht BN (2008). Dendritic cells and epithelial cells: linking innate and adaptive immunity in asthma. *Nat Rev Immunol* 8: 193–204.
- Hammad H, Kool M, Soullie T, Narumiya S, Trottein F, Hoogsteden HC *et al.* (2007). Activation of the D prostanoid 1 receptor suppresses asthma by modulation of lung dendritic cell function and induction of regulatory T cells. *J Exp Med* 204: 357–367.
- Hanasaki K, Nakano K, Kasai H, Arita H, Ohtani K, Doteuchi M (1988). Specific receptors for thromboxane A<sub>2</sub> in cultured vascular smooth muscle cells of rat aorta. *Biochem Biophys Res Commun* 150: 1170–1175.
- Harris KJ (2008). A combination of niacin and a prostaglandin D<sub>2</sub> receptor antagonist. WO2008/039882, Sanofi-Aventis.
- Hawkey CJ (2000). Management of gastroduodenal ulceration caused by non-steroidal anti-inflammatory drugs. *Baillieres Best Pract Res Clin Gastroenterol* 14: 173–192.
- Heavey DJ, Lumley P, Barrow SE, Murphy MB, Humphrey PP, Dollery CT (1984). Effects of intravenous infusions of prostaglandin D<sub>2</sub> in man. *Prostaglandins* 28: 755–767.
- Hedberg A, Hall SE, Ogletree ML, Harris DN, Liu EC (1988). Characterization of [5,6-<sup>3</sup>H]SQ 29,548 as a high affinity radioligand, binding to thromboxane A<sub>2</sub>/prostaglandin H<sub>2</sub>-receptors in human platelets. *J Pharmacol Exp Ther* 245: 786–792.
- Hefferan MP, Carter C, Haley M, Loomis CW (2003). Spinal nerve injury activates prostaglandin synthesis in the spinal cord that contributes to early maintenance of tactile allodynia. *Pain* 101: 139–147.
- Heinemann A, Schuligoi R, Sabroe I, Hartnell A, Peskar BA (2003). Δ<sup>12</sup>-Prostaglandin J<sub>2</sub>, a plasma metabolite of prostaglandin D<sub>2</sub>, causes eosinophil mobilization from the bone marrow and primes eosinophils for chemotaxis. *J Immunol* 170: 4752–4758.
- Heptinstall S, Espinosa DI, Manolopoulos P, Glenn JR, White AE, Johnson A *et al.* (2008). DG-041 inhibits the EP<sub>3</sub> prostanoid receptor – a new target for inhibition of platelet function in atherosclerotic disease. *Platelets* 19: 605–613.
- Hirai H, Tanaka K, Yoshie O, Ogawa K, Kenmotsu K, Takamori Y *et al.* (2001). Prostaglandin D<sub>2</sub> selectively induces chemotaxis in T helper type 2 cells, eosinophils, and basophils via seven-transmembrane receptor CRTH2. *J Exp Med* 193: 255–261.
- Hirai H, Tanaka K, Takano S, Ichimasa M, Nakamura M, Nagata K (2002). Agonistic effect of indomethacin on a prostaglandin D<sub>2</sub> receptor, CRTH2. *J Immunol* 168: 981–985.
- Hirata M, Hayashi Y, Ushikubi F, Yokota Y, Kageyama R, Nakanishi S *et al.* (1991). Cloning and expression of cDNA for a human thromboxane A<sub>2</sub> receptor. *Nature* 349: 617–620.
- Hirata T, Ushikubi F, Kakizuka A, Okuma M, Narumiya S (1996). Two thromboxane A<sub>2</sub> receptor isoforms in human platelets. Opposite coupling to adenylyl cyclase with different sensitivity to Leu mutation. *J Clin Invest* 97: 949–956.
- Hirst JR, Parkington HC, Young IR, Palliser HK, Peri KG, Olson DM (2005). Delay of preterm birth in sheep by PHG 113.31, a prostaglandin F<sub>2α</sub> receptor antagonist. *Am J Obstet Gynecol* 193: 256–266.
- Hoet B, Falcon C, De Reys S, Arnout J, Deckmyn H, Vermylen J (1990). R68070, a combined thromboxane/endoperoxide receptor antagonist and thromboxane synthase inhibitor, inhibits human platelet activation *in vitro* and *in vivo*: a comparison with aspirin. *Blood* 75: 646–653.
- Honma T, Hiramatsu K, Arimura A (1998). Benzothiofenecarboxamide derivatives and PGD<sub>2</sub> antagonists comprising them. WO98/25919, Shionogi & Co Ltd.
- Honn KV, Meyer J (1981). Thromboxanes and prostacyclin: positive and negative modulators of tumour growth. *Biochem Biophys Res Commun* 102: 1122–1129.
- Horne WC (1984). Desacetyl-1-nantradol: a selective prostaglandin antagonist. *Prostaglandins Leukot Med* 15: 129–136.
- Huang JL, Gao PS, Mathias RA, Yao TC, Chen LC, Kuo ML *et al.* (2004). Sequence variants of the gene encoding chemoattractant receptor expressed on Th2 cells (CRTH2) are associated with asthma and differentially influence mRNA stability. *Hum Mol Genet* 13: 2691–2697.
- Hung GH, Jones RL, Lam FF, Chan KM, Hidaka H, Suzuki M *et al.* (2006). Investigation of the pronounced synergism between prostaglandin E<sub>2</sub> and other constrictor agents on rat femoral artery. *Prostaglandins Leukot Essent Fatty Acids* 74: 401–405.
- Hutchinson J, Marshall KM, Senior J (2003). Preliminary studies using a putative FP-receptor antagonist, AL-8810, on isolated mouse uterus. Proceedings of the British Pharmacological Society at <http://www.pa2online.org/Vol11Issue3abst038P.html>.
- Ikeda M, Kawatani M, Maruyama T, Ishihama H (2006). Prostaglandin facilitates afferent nerve activity via EP<sub>1</sub> receptors during urinary bladder inflammation in rats. *Biomed Res* 27: 49–54.
- Imura Y, Terashita Z, Shibouta Y, Nishikawa K (1988). The thromboxane A<sub>2</sub>/prostaglandin endoperoxide receptor antagonist activity of CV-1451, a thromboxane A<sub>2</sub> synthetase inhibitor. *Eur J Pharmacol* 147: 359–365.
- Imura Y, Terashita Z, Shibouta Y, Inada Y, Nishikawa K (1990). Antagonistic action of AA-2414 on thromboxane A<sub>2</sub>/prostaglandin endoperoxide receptor in platelets and blood vessels. *Jpn J Pharmacol* 52: 35–43.
- Ishiguro C, Fujita T, Omori T, Fujii Y, Mayama T, Sato T (2008). Assessing the effects of non-steroidal anti-inflammatory drugs on antihypertensive drug therapy using post-marketing surveillance database. *J Epidemiol* 18: 119–124.
- Ishiura Y, Fujimura Y, Yamamori C, Nobata K, Myou S, Kurashima K *et al.* (2003). Thromboxane antagonism and cough in chronic bronchitis. *Ann Med* 35: 135–139.
- Ito S, Okuda E, Sugama K, Negishi M, Hayaishi O (1990). Evaluation of ZK110841 and AH6809, an agonist and an antagonist of prostaglandin DP-receptors on human platelets, with a PGD<sub>2</sub>-responsive cell line from bovine embryonic trachea. *Br J Pharmacol* 99: 13–14.
- Itoh Y, Shindoh J, Horiba M, Kohno S, Ohata K, Ashida Y *et al.* (1993). Inhibitory effects of AA-2414, a thromboxane (Tx) A<sub>2</sub> receptor antagonist, on U-46619-, prostaglandin (PG) D<sub>2</sub>- and 9α,11β PGF<sub>2</sub>-induced contractions of guinea-pig tracheas and isolated human bronchi (Japanese). *Arerugi* 42: 1670–1676.
- Ivey KN, Srivastava D (2006). The paradoxical patent ductus arteriosus. *J Clin Invest* 116: 2863–2866.
- Iwasaki M, Nagata K, Takano S, Takahashi K, Ishii N, Ikezawa Z (2002). Association of a new-type prostaglandin D<sub>2</sub> receptor CRTH2 with circulating T helper 2 cells in patients with atopic dermatitis. *J Invest Dermatol* 119: 609–616.
- Jadhav V, Jabre A, Lin SZ, Lee TJ (2004). EP<sub>1</sub>- and EP<sub>3</sub>-receptors mediate prostaglandin E<sub>2</sub>-induced constriction of porcine large cerebral arteries. *J Cereb Blood Flow Metab* 24: 1305–1316.
- Janssen LJ, Premji M, Netherton S, Coruzzi J, Lu-Chao H, Cox PG (2001). Vasoconstrictor actions of isoprostanines via tyrosine kinase and Rho kinase in human and canine pulmonary vascular smooth muscles. *Br J Pharmacol* 132: 127–134.
- Janssens WJ, Cools FJ, Hoskens LA, Van Neuten JM (1990). Effect of ridogrel on vascular contractions caused by vasoactive substances released during platelet activation. *Thromb Haemost* 64: 91–96.
- Jenkins DW, Feniuk W, Humphrey PPA (2001). Characterization of the prostanoid receptor types involved in mediating calcitonin gene-related peptide release from cultured rat trigeminal neurones. *Br J Pharmacol* 134: 1296–1302.
- Johnson AG, Nguyen TV, Day RO (1994). Do nonsteroidal anti-inflammatory drugs affect blood pressure? A meta-analysis. *Ann Intern Med* 121: 289–300.
- Jones R, Woodward D, Wang J (2008). Investigation of the slow kinetics of the prostanoid EP<sub>3</sub> receptor antagonists L-798106 and L-826266 on guinea-pig aorta. *Fundam Clin Pharmacol* 22 (Suppl. 2): P078.

- Jones RL (1976). Cardiovascular actions of prostaglandins D and E in the sheep: evidence for two distinct receptors. *Adv Prostaglandin Thromboxane Res* 1: 221–230.
- Jones RL (1978). Definition of prostaglandin-sensitive arterial constrictor systems. *Acta Biol Med Ger* 37: 837–844.
- Jones RL (2004). An update on ligands for prostanoid receptors. *Sigma-RBI Newsletter for Cell Signalling and Neurosci Res* 20: 1–9. <http://www.sigmaaldrich.com/sigma/general%20information/prostanoidreceptors.pdf>
- Jones RL, Chan K (2001). Distinction between relaxations induced via prostanoid EP<sub>4</sub> and IP<sub>1</sub> receptors in pig and rabbit blood vessels. *Br J Pharmacol* 134: 313–324.
- Jones RL, Chan K (2005). Investigation of the agonist activity of prostacyclin analogues on prostanoid EP<sub>4</sub> receptors using GW 627368 and taprostene: evidence for species differences. *Prost Leukot Essent Fatty Acids* 72: 289–299.
- Jones RL, Wilson NH (1978). Tautomerism of the enedione system of 15-oxoprostaglandin D<sub>2</sub>. *J Org Chem (Perkin 1)*, 209–214.
- Jones RL, Peesapati V, Wilson NH (1982). Antagonism of the thromboxane-sensitive contractile systems of the rabbit aorta, dog saphenous vein and guinea-pig trachea. *Br J Pharmacol* 76: 423–438.
- Jones RL, Wilson NH, Lawrence RA (1989). EP 171: a high affinity thromboxane A<sub>2</sub>-mimetic, the actions of which are slowly reversed by receptor blockade. *Br J Pharmacol* 96: 875–887.
- Jones RL, Wilson NH, Marr CG, Muir G, Armstrong RA (1993). Diphenylmethylazine prostanoids with prostacyclin-like actions on human platelets. *J Lipid Mediat* 6: 405–410.
- Jones RL, Qian YM, Wise H, Wong HN, Lam WL, Chan HW *et al.* (1997). Relaxant actions of non-prostanoid prostacyclin mimetics on human pulmonary artery. *J Cardiovasc Pharmacol* 29: 525–535.
- Jones RL, Qian YM, Chan KM, Yip AP (1998). Characterization of a prostanoid EP<sub>3</sub>-receptor in guinea-pig aorta: partial agonist action of the non-prostanoid ONO-AP-324. *Br J Pharmacol* 125: 1288–1296.
- Jones RL, Wise H, Clark R, Whiting RL, Bley KR (2006). Investigation of the prostacyclin (IP) receptor antagonist RO1138452 on isolated blood vessel and platelet preparations. *Br J Pharmacol* 149: 110–120.
- Juteau H, Gareau Y, Labelle M, Sturino CF, Sawyer N, Tremblay N *et al.* (2001). Structure-activity relationship of cinnamic acylsulfonamide analogues on the human EP<sub>3</sub> prostanoid receptor. *Bioorg Med Chem* 9: 1977–1984.
- Kabashima K, Saji T, Murata T, Nagamachi M, Matsuoka T, Segi E *et al.* (2002). The prostaglandin receptor EP<sub>4</sub> suppresses colitis, mucosal damage and CD4 cell activation in the gut. *J Clin Invest* 109: 883–893.
- Kabashima K, Sakata D, Nagamachi M, Miyachi Y, Inaba K, Narumiya S (2003). Prostaglandin E<sub>2</sub>-EP<sub>4</sub> signaling initiates skin immune responses by promoting migration and maturation of Langerhans cells. *Nature Med* 9: 744–749.
- Kabashima K, Nagamachi M, Honda T, Nishigori C, Miyachi Y, Tokura Y (2007). Prostaglandin E<sub>2</sub> is required for ultraviolet B-induced skin inflammation via EP<sub>2</sub> and EP<sub>4</sub> receptors. *Lab Invest* 87: 49–55.
- Kam Y, Chow KB, Wise H (2001). Factors affecting prostacyclin receptor agonist efficacy in different cell types. *Cell Signal* 13: 841–847.
- Kamanna VS, Vo A, Kashyap ML (2008). Nicotinic acid: recent developments. *Curr Opin Cardiol* 23: 393–398.
- Kanamori Y, Niwa M, Kohno K, Al-Essa LY, Matsuno H, Kozawa O *et al.* (1997). Migration of neutrophils from blood to tissue: alteration of modulatory effects of prostanoid on superoxide generation in rabbits and humans. *Life Sci* 60: 1407–1417.
- Kamata S, Haga N, Tsuru T, Uchida K, Kakushi H, Arita H (1990). Synthesis of thromboxane receptor antagonists with bicyclo [3.1.0]hexane ring systems. *J Med Chem* 33: 229–239.
- Karanam B, Madeira M, Bradley S, Wenning L, Desai R, Soli E *et al.* (2007). Absorption, metabolism, and excretion of [<sup>14</sup>C]MK-0524, a prostaglandin D<sub>2</sub> receptor antagonist, in humans. *Drug Metab Dispos* 35: 1196–1202.
- Karasawa A, Kawakage M, Shirakura S, Higo K, Kubo K, Ohshima E *et al.* (1991a). Antiplatelet effects of the novel thromboxane A<sub>2</sub> receptor antagonist sodium (E)-11-[2-(5,6-dimethyl-1-benzimidazolyl)-ethylidene]-6,11-dihydrodibenz[b,e] oxepine-2-carboxylate monohydrate. *Arzneimittelforschung* 41: 1230–1236.
- Karasawa A, Shirakura S, Higo K, Kubo K (1991b). Actions of the novel thromboxane A<sub>2</sub> receptor antagonist sodium (E)-11-[2-(5,6-dimethyl-1-benzimidazolyl)-ethylidene]-6,11-dihydrodibenz[b,e]oxepine-1-carboxylate monohydrate on smooth muscle preparations. *Arzneimittelforschung* 41: 1237–1241.
- Kassuya CA, Ferreira J, Claudino RF, Calixto JB (2007). Intraplantar PGE<sub>2</sub> causes nociceptive behaviour and mechanical allodynia: the role of prostanoid E receptors and protein kinases. *Br J Pharmacol* 150: 727–737.
- Katsura M, Miyamoto T, Hamanaka N, Kondo K, Terada T, OHgaki Y *et al.* (1983). *In vitro* and *in vivo* effects of new powerful thromboxane antagonists (3-alkylamino pinane derivatives). *Adv Prost Thromb Leukot Res* 11: 351–357.
- Kawahara H, Sakamoto A, Takeda S, Onodera H, Imaki J, Ogawa R (2001). A prostaglandin E<sub>2</sub> receptor subtype EP<sub>1</sub> receptor antagonist (ONO-8711) reduces hyperalgesia, allodynia, and c-fos gene expression in rats with chronic nerve constriction. *Anaesth Analg* 93: 1012–1017.
- Kawamori T, Uchiya N, Kitamura T (2001). Evaluation of a selective prostaglandin E receptor EP<sub>1</sub> antagonist for potential properties in colon carcinogenesis. *Anticancer Res* 21: 3865–3869.
- Kawano T, Anrather J, Zhou P, Park L, Wang G, Frys KA *et al.* (2006). Prostaglandin E<sub>2</sub> EP<sub>1</sub> receptors: downstream effectors of COX-2 neurotoxicity. *Nat Med* 12: 225–229.
- Kawikova I, Barnes PJ, Takahashi T, Tadjkarimi S, Yacoub MH, Belvisi MG (1996). 8-Epi-PGF<sub>2α</sub>, a novel noncyclooxygenase-derived prostaglandin, constricts airways *in vitro*. *Am J Respir Crit Care Med* 153: 590–596.
- Keery RJ, Lumley P (1988). AH6809, a prostaglandin DP-receptor blocking drug on human platelets. *Br J Pharmacol* 94: 745–754.
- Keith JC, Spitz B, Van Assche FA (1993). Thromboxane synthetase inhibition as a new therapy for preeclampsia: animal and human studies minireview. *Prostaglandins* 45: 3–13.
- Keitz PF, Jahangir A, Lopez-Tapia FJ, O'Yang C (2004). Substituted arylamides as IP antagonists US 2004/0220247A1 Roche Palo Alto.
- Kemp BK, McPherson GA (1998). Interspecies differences in thromboxane A<sub>2</sub> receptors are distinguished by glibenclamide. *Eur J Pharmacol* 354: 173–178.
- Khasar SG, Green PG, Levine PD (1993). Comparison of intradermal and subcutaneous hyperalgesic effects of inflammatory mediators in the rat. *Neurosci Lett* 153: 215–218.
- Khera M, Boone TB, Salas N, Jett MF, Somogyi GT (2007). The role of the prostacyclin receptor antagonist RO3244019 in treating neurogenic detrusor overactivity after spinal cord injury in rats. *BJU Int* 99: 442–446.
- Kim N, Luster AD (2007). Regulation of immune cells by eicosanoid receptors. *Scientif World J* 7: 1307–1328.
- Kimura M, Osumi S, Ogihara M (2000). Stimulation of DNA synthesis and proliferation by prostaglandins in primary cultures of adult rat hepatocytes. *Eur J Pharmacol* 404: 259–271.
- Kimura M, Osumi S, Ogihara M (2001). Prostaglandin E<sub>2</sub> EP<sub>1</sub> receptor agonist-induced DNA synthesis and proliferation in primary cultures of adult rat hepatocytes: the involvement of TGFA. *Endocrinology* 142: 4428–4440.
- Kishino J, Hanasaki K, Nagasaki T, Arita H (1991). Kinetic studies on stereospecific recognition by the thromboxane A<sub>2</sub>/prostaglandin H<sub>2</sub> receptor of the antagonist, S-145. *Br J Pharmacol* 103: 1883–1888.
- Kitamura N, Hukuda R, Majima T, Horie T, Sugihara T (2003a). The 3 effective cases of cough variant asthma with ramatroban. *Jpn J Allergol* 52: 1089–1092.
- Kitamura T, Itoh M, Nada T, Tani K, Kobayashi M, Maruyama T *et al.*

- (2003b). Combined effects of prostaglandin E receptor subtype E<sub>1</sub> and subtype EP<sub>4</sub> antagonists on intestinal tumorigenesis in adenomatous polyposis coli gene knockout mice. *Comcer Sci* **94**: 618–621.
- Kitanaka J, Ishibashi T, Baba A (1993). Phloretin as an antagonist of prostaglandin F<sub>2α</sub> receptor in cultured rat astrocytes. *J Neurochem* **60**: 704–708.
- Kitawaki T, Kadowaki N, Sugimoto N, Kambe N, Hori T, Miyachi Y *et al.* (2006). IgE-activated mast cells in combination with pro-inflammatory factors induce Th2-promoting dendritic cells. *Int Immunol* **18**: 1789–1799.
- Knapp HR, Heally C, Lawson J, FitzGerald GA (1988). Effects of low-dose aspirin on endogenous eicosanoid formation in normal and atherosclerotic men. *Thromb Res* **50**: 377–386.
- Koda N, Tsutsui Y, Niwa H, Ito S, Woodward DF, Watanabe K (2004). Synthesis of prostaglandin F ethanamide by prostaglandin F synthase and identification of bimatoprost as a potent inhibitor of the enzyme. New enzyme method by LC/GSI/MS. *Arch Biochem Biophys* **424**: 128–136.
- Kopp UC, Cicha MZ, Nakamura K, Nusing RM, Smith LA, Hokfelt T (2004). Activation of EP<sub>4</sub> receptors contributes to prostaglandin E<sub>2</sub>-mediated stimulation of renal sensory nerves. *Am J Physiol Renal Physiol* **287**: F1269–F1282.
- Kostenis E, Ulven T (2006). Emerging roles of DP and CRTH2 in allergic inflammation. *Trends Mol Med* **12**: 148–158.
- Kozak KR, Crews BC, Morrow JD, Wang LH, Ma YH, Weinander R *et al.* (2002). Metabolism of the endocannabinoids, 2-arachidonylglycerol and anandamide, into prostaglandin, thromboxane, and prostacyclin glycerol esters and ethanalamides. *J Biol Chem* **277**: 44877–44885.
- Krauss AH, Woodward DF, Donde Y, Burk RM (2005). Prostaglandin D<sub>2</sub> antagonist. WO2005/079793, Allergan Inc.
- Kromer BM, Tippins JR (1996). Coronary artery constriction by the isopropane 8-epi prostaglandin F<sub>2α</sub>. *Br J Pharmacol* **119**: 1276–1280.
- Kuhn CF, Feru F, Bazin M, Awad MMA, Goldstein SW (2004). Quinoline derivatives as CRTH2 antagonists. EP1435356, Warner-Lambert & Co.
- Kumazawa T, Mizumura K, Koda H, Fukusako H (1996). EP receptor subtypes implicated in the PGE<sub>2</sub>-induced sensitization of poly-modal receptors in response to bradykinin and heat. *J Neurophysiol* **75**: 2361–2368.
- Kunikata T, Tanaka A, Miyazawa T, Kato S, Takeuchi K (2002). 16,16-dimethyl prostaglandin E<sub>2</sub> inhibits indomethacin-induced small intestinal lesions through EP<sub>3</sub> and EP<sub>4</sub> receptors. *Dig Dis Sci* **47**: 894–904.
- Kuwano T, Nakao S, Yamamoto H, Tsuneyoshi M, Yamamoto T, Kuwano M *et al.* (2004). Cyclooxygenase 2 is a key enzyme for inflammatory cytokine-induced angiogenesis. *FASEB J* **18**: 300–310.
- Lai E, De Lepeleire I, Crumley TM, Liu F, Wenning LA, Michiels N *et al.* (2007). Suppression of niacin-induced vasodilation with an antagonist to prostaglandin D<sub>2</sub> receptor subtype 1. *Clin Pharmacol Ther* **81**: 849–857.
- Lai E, Wenning LA, Crumley TM, De LI, Liu F, de Hoon JN *et al.* (2008b). Pharmacokinetics, pharmacodynamics, and safety of a prostaglandin D<sub>2</sub> receptor antagonist. *Clin Pharmacol Ther* **83**: 840–847.
- Lai YJ, Pullamsetti SS, Dony E, Weissmann N, Butrous G, Banat GA *et al.* (2008a). Role of prostanoid EP<sub>4</sub> receptor in iloprost-mediated vasodilatation in pulmonary hypertension. *Am J Respir Crit Care Med* **178**: 188–196.
- Langevin B, Orton E, Sherer D (2007). Dihydrogen phosphate salt of a prostaglandin D<sub>2</sub> receptor antagonist. WO2007/047378, Sanofi-Aventis.
- Lawrence RA, Jones RL (1992). Investigation of the prostaglandin E (EP-) receptor subtype mediating relaxation of the rabbit jugular vein. *Br J Pharmacol* **105**: 817–824.
- Lawrence RA, Jones RL, Wilson NH (1992). Characterization of receptors involved in the direct and indirect actions of prostaglandins E and I on the guinea-pig ileum. *Br J Pharmacol* **105**: 271–278.
- Lazareno S, Birdsall NJ (1993). Estimation of competitive antagonist affinity from functional inhibition curves using the Gaddum, Schild and Cheng-Prusoff equations. *Br J Pharmacol* **109**: 1110–1119.
- Leff P, Dougall IG (1993). Further concerns over Cheng-Prusoff analysis. *Trends Pharmacol Sci* **14**: 110–112.
- Leff P, Giles H (1992). Classification of platelet and vascular prostaglandin D<sub>2</sub> (DP) receptors: estimation of affinities and relative efficacies for a series of novel bicyclic ligands. With an appendix on goodness-of-fit analyses. *Br J Pharmacol* **106**: 996–1003.
- Leonhardt A, Glaser A, Wegmann M, Schranz D, Seyberth H, Nusing R (2003). Expression of prostanoid receptors in human ductus arteriosus. *Br J Pharmacol* **138**: 655–659.
- Lewis RA, Soter NA, Diamond PT, Austen KF, Oates JA, Roberts LJ (1982). Prostaglandin D<sub>2</sub> generation after activation of rat and human mast cells with anti-IgE. *J Immunol* **129**: 1627–1631.
- Liang Y, Li C, Guzman VM, Evinger AJ 3rd, Protzman CE, Krauss AH, Woodward DF (2003). Comparison of prostaglandin F<sub>2α</sub>, bimatoprost (prostanamide), and butaprost (EP<sub>2</sub> agonist) on Cyr61 and connective tissue growth factor gene expression. *J Biol Chem* **278**: 27267–27277.
- Liang Y, Li C, Guzman VM, Chang WW, Evinger AJ, Pablo JV, Woodward DF (2004). Upregulation of orphan nuclear receptor Nur77 following PGF<sub>2α</sub>, Bimatoprost, and Butaprost treatments. Essential role of a protein kinase C pathway involved in EP<sub>2</sub> receptor activated Nur77 gene transcription. *Br J Pharmacol* **142**: 737–748.
- Liang Y, Woodward DF, Guzman VM, Li C, Scott DF, Wang JW *et al.* (2008). Identification and pharmacological characterization of the prostaglandin FP receptor and FP receptor variant complexes. *Br J Pharmacol* **154**: 1079–1093.
- Lin CR, Amaya F, Barrett L, Wang H, Takada J, Samad TA *et al.* (2006). Prostaglandin E<sub>2</sub> receptor EP<sub>4</sub> contributes to inflammatory pain hypersensitivity. *J Pharmacol Exp Ther* **319**: 1096–1103.
- Liu YJ, Jackson DM, Blackham A (1996a). Effects of BW A868C, a selective prostaglandin DP receptor antagonist, in dog isolated vascular preparations. *Eur J Pharmacol* **303**: 187–192.
- Liu YJ, Jackson DM, Blackham A, Leff P (1996b). Partial agonist effects of BW A868C, a selective DP receptor antagonist, on Cl<sup>-</sup> secretion in dog tracheal epithelium. *Eur J Pharmacol* **304**: 117–122.
- Lovell JM (2007). Salts with CRTH2 antagonist activity. WO2007/107772, Oxagen Ltd.
- Lumley P, White BP, Humphrey PPA (1989). GR32191, a highly potent and specific thromboxane A<sub>2</sub> receptor blocking drug on platelets and vascular and airways smooth muscle *in vitro*. *Br J Pharmacol* **97**: 783–794.
- Lydford SJ, McKechnie KC (1994). Characterization of the prostaglandin E<sub>2</sub> sensitive (EP)-receptor in the rat isolated trachea. *Br J Pharmacol* **112**: 133–136.
- Lydford SJ, Li SW, McKechnie KC (1996a). Comparison of prostanoid DP receptors in the rabbit saphenous vein and human neutrophil. *Br J Pharmacol* **117**: 190P.
- Lydford SJ, McKechnie KC, Dougall IG (1996b). Pharmacological studies on prostanoid receptors in the rabbit isolated saphenous vein: a comparison with the rabbit isolated ear artery. *Br J Pharmacol* **117**: 13–20.
- Lydford SJ, McKechnie KC, Leff P (1996c). Interaction of BW A868C, a prostanoid DP-receptor antagonist, with two receptor subtypes in the rabbit isolated saphenous vein. *Prostaglandins* **52**: 125–139.
- Ma H, Hara A, Xiao CY, Okada Y, Takahata O, Nakaya K *et al.* (2001). Increased bleeding tendency and decreased susceptibility to thromboembolism in mice lacking the prostaglandin E receptor subtype EP<sub>3</sub>. *Circulation* **104**: 1176–1180.
- Maassen VanDenBrink A, Bax WA, Ferrari MD, Zijlstra FJ, Bos E *et al.* (1996). Augmented contraction of the human isolated coronary artery by sumatriptan: a possible role for endogenous thromboxane. *Br J Pharmacol* **119**: 855–862.

- Machwate M, Harada S, Leu CT, Seedor G, Labelle M, Gallant M *et al.* (2001). Prostaglandin receptor EP<sub>4</sub> mediates the bone anabolic effects of PGE<sub>2</sub>. *Mol Pharmacol* **60**: 36–41.
- Macias-Perez IM, Zent R, Carmosino M, Breyer MD, Breyer RM, Pozzi A (2008). Mouse EP<sub>3</sub>  $\alpha$ ,  $\beta$  and  $\gamma$  receptor variants reduce tumor cell proliferation and tumorigenesis *in vivo*. *J Biol Chem* **283**: 12538–12545.
- MacIntyre DE, Gordon JL (1977). Discrimination between platelet prostaglandin receptors with a specific antagonist of bisenoic prostaglandins. *Thromb Res* **11**: 705–713.
- McKenniff MG, Norman P, Cuthbert NJ, Gardiner PJ (1991). BAY u3405, a potent and selective thromboxane A<sub>2</sub> receptor antagonist on airway smooth muscle *in vitro*. *Br J Pharmacol* **104**: 585–590.
- McKenniff M, Rodger IW, Norman P, Gardiner PJ (1988). Characterisation of receptors mediating the contractile effects of prostanoids in guinea-pig and human airways. *Eur J Pharmacol* **153**: 149–159.
- Maddox YT, Ramwell PW, Shiner CS, Corey EJ (1978). Amide and 1-amino derivatives of F prostaglandins as prostaglandin antagonists. *Nature* **273**: 549–552.
- Mais DE, Saussy DLJ, Chaikhouni A, Kochel PJ, Knapp DR, Hamanaka N *et al.* (1985). Pharmacologic characterization of human and canine thromboxane A<sub>2</sub>/prostaglandin H<sub>2</sub> receptors in platelets and blood vessels: evidence for different receptors. *J Pharmacol Exp Ther* **233**: 418–424.
- Mais DE, DeHoll D, Sightler H, Halushka PV (1988). Different pharmacologic activities for 13-azapinane thromboxane A<sub>2</sub> analogs in platelets and blood vessels. *Eur J Pharmacol* **148**: 309–315.
- Majima M, Amano H, Hayashi I (2003). Prostanoid receptor signaling relevant to tumor growth and angiogenesis. *Trends Pharmacol Sci* **24**: 524–529.
- Makino H, Tanaka I, Mukoyama M, Sugawara A, Mori K, Muro S *et al.* (2002). Prevention of diabetic nephropathy in rats by prostaglandin E receptor EP<sub>1</sub>-selective antagonist. *J Am Soc Nephrol* **13**: 1757–1765.
- Malini PL, Stocchi E, Zanardi M, Milani M, Ambrosioni E (1997). Thromboxane antagonism and cough induced by angiotensin-converting-enzyme inhibitor. *Lancet* **350**: 15–18.
- Malmberg AB, Rafferty MF, Yaksh TL (1994). Antinociceptive effect of spinally delivered prostaglandin E receptor antagonists in the formalin test on the rat. *Neurosci Letts* **173**: 193–196.
- Masataka M, Amano H, Izumi H (2003). Prostanoid receptor signaling relevant to tumor growth and angiogenesis. *Trends Pharmacol Sci* **24**: 524–529.
- Masuda A, Mais DE, Oatis JE, Halushka PV (1991). Platelet and vascular thromboxane A<sub>2</sub>/prostaglandin H<sub>2</sub> receptors. Evidence for different subclasses in the rat. *Biochem Pharmacol* **42**: 537–544.
- Matias I, Chen J, De Petrocellis L, Bisogno T, Ligresti A, Fezza F *et al.* (2004). Prostaglandin ethanolamides (prostamides): *in vitro* pharmacology and metabolism. *J Pharmacol Exp Ther* **309**: 745–757.
- Mathiesen JM, Christopoulos A, Ulven T, Royer JF, Campillo M, Heinemann A *et al.* (2006). On the mechanism of interaction of potent surmountable and insurmountable antagonists with the prostaglandin D<sub>2</sub> receptor CRTH2. *Mol Pharmacol* **69**: 1441–1453.
- Matsuoka T, Hirata M, Tanaka H, Takahashi Y, Murata T, Kabashima K *et al.* (2000). Prostaglandin D<sub>2</sub> as a mediator of allergic asthma. *Science* **287**: 2013–2017.
- Matthews JS, Jones RL (1993). Potentiation of aggregation and inhibition of adenylate cyclase in human platelets by prostaglandin E analogues. *Br J Pharmacol* **108**: 363–369.
- Maubach KA, Clark DE, Fenton G, Lockey PM, Clark KL, Oxford AW *et al.* (2009). BCG20-1531, a novel, potent and selective prostanoid EP<sub>4</sub> receptor antagonist; a putative new treatment for migraine headache. *Br J Pharmacol* **156**: 316–327.
- Meanwell NA, Romine JL, Seiler SM (1994). Non-prostanoid prostacyclin mimetics. *Drugs Future* **19**: 361–385.
- Mebane H, Turnbach ME, Randich A (2003). Spinal EP receptors mediating prostaglandin E<sub>2</sub>-induced mechanical hyperalgesia, thermal hyperalgesia, and touch-evoked allodynia in rats. *J Pain* **4**: 392–399.
- Merritt JE, Brown AM, Bund S, Cooper DG, Egan JW, Hallam TJ *et al.* (1991a). Primate vascular responses to octimibate, a non-prostanoid agonist at the prostacyclin receptor. *Br J Pharmacol* **102**: 260–266.
- Merritt JE, Hallam TJ, Brown AM, Boyfield I, Cooper DG, Hickey DM *et al.* (1991b). Octimibate, a potent non-prostanoid inhibitor of platelet aggregation, acts via the prostacyclin receptor. *Br J Pharmacol* **102**: 251–259.
- Middlemiss D, Ashton MR, Boyd AE, Brookfield FA (2005a). Compounds with PGD<sub>2</sub> antagonist activity. WO2005/040112, Oxagen Ltd.
- Middlemiss D, Ashton MR, Boyd AE, Brookfield FA (2005b). Substituted indol-3-yl acetic acid derivatives. GB 2 407 318, Oxagen Ltd.
- Middlemiss D, Ashton MR, Boyd AE, Brookfield FA (2005c). Use of CRTh2 antagonist compounds in therapy. WO2005/044260, Oxagen Ltd.
- Middlemiss D, Ashton MR, Boyd AE, Brookfield FA, Armer RE (2005d). Compounds having CRTh2 antagonist activity. WO2005/040114, Oxagen Ltd.
- Mihara S, Hara S, Ueda M, Ide M, Fujimoto M (1989). Antagonistic actions of S-145 on vascular and platelet thromboxane A<sub>2</sub> receptors. *Eur J Pharmacol* **171**: 179–187.
- Miki I, Kishibayashi N, Nonaka H, Ohshima E, Takami H, Obase H *et al.* (1992). Effects of KW-3635, a novel dibenzoxepin derivative of a selective thromboxane A<sub>2</sub> antagonist, on human, guinea pig and rat platelets. *Jpn J Pharmacol* **59**: 357–364.
- Mimura H, Ikemura T, Kotera O, Sawada M, Tashiro S, Fuse E *et al.* (2005). Inhibitory effect of the 4-aminotetrahydroquinoline derivatives, selective chemoattractant receptor-homologous molecule expressed on T helper 2 cell antagonists, on eosinophil migration induced by prostaglandin D<sub>2</sub>. *J Pharmacol Exp Ther* **314**: 244–251.
- Minami T, Nishihara I, Sakamoto K, Ito S, Hyodo M, Hayashi O (1995). Blockade by ONO-NT-012, a unique prostanoid analogue, of prostaglandin E<sub>2</sub>-induced allodynia in conscious mice. *Br J Pharmacol* **115**: 73–76.
- Minami T, Nakano H, Kobayashi T, Sugimoto Y, Ushikubi F, Ichikawa A *et al.* (2001). Characterization of EP receptor subtypes responsible for prostaglandin E<sub>2</sub>-induced pain responses by use of EP<sub>1</sub> and EP<sub>3</sub> receptor knockout mice. *Br J Pharmacol* **133**: 438–444.
- Minami T, Matsumura S, Mabuchi T, Kobayashi T, Sugimoto Y, Ushikubi F *et al.* (2003). Functional evidence for interaction between prostaglandin EP<sub>3</sub> and kappa-opioid receptor pathways in tactile pain induced by human immunodeficiency virus type-1 (HIV-1) glycoprotein gp120. *Neuropharmacology* **45**: 96–105.
- Misra RA (1994). Recent progress in the clinical development of thromboxane A<sub>2</sub> receptor antagonists. *Exp Opin Invest Drugs* **3**: 469–480.
- Mitsumori S, Tsuru T, Honma T, Hiramatsu Y, Okada T, Hashizume H *et al.* (2003a). Synthesis and biological activity of various derivatives of a novel class of potent, selective, and orally active prostaglandin D<sub>2</sub> receptor antagonists. 1. Bicyclo[2.2.1]heptane derivatives. *J Med Chem* **46**: 2436–2445.
- Mitsumori S, Tsuru T, Honma T, Hiramatsu Y, Okada T, Hashizume H *et al.* (2003b). Synthesis and biological activity of various derivatives of a novel class of potent, selective, and orally active prostaglandin D<sub>2</sub> receptor antagonists. 1. 6,6-Dimethylbicyclo[3.1.1]heptane derivatives. *J Med Chem* **46**: 2446–2455.
- Momma K, Toyoshima K, Takeuchi D, Imamura S, Nakanishi T (2005). *In vivo* constriction of the fetal and neonatal ductus arteriosus by a prostanoid EP<sub>4</sub>-receptor antagonist. *Pediatr Res* **58**: 971–975.
- Monneret G, Gravel S, Diamond M, Rokach J, Powell WS (2001). Prostaglandin D<sub>2</sub> is a potent chemoattractant for human eosinophils that acts via a novel DP receptor. *Blood* **98**: 1942–1948.
- Morinelli TA, Okwu AK, Mais DE, Halushka PV, John V, Chen CK *et al.* (1989). Difluorothromboxane A<sub>2</sub> and stereoisomers: stable derivatives of thromboxane A<sub>2</sub> with differential effects on platelets and blood vessels. *Proc Natl Acad Sci USA* **86**: 5600–5604.

- Moriuchi H, Koda N, Okuda-Ashitaka E, Daiyasu H, Ogasarawa K, Toh H *et al.* (2008). Molecular characterization of a novel type of prostamide/prostaglandin F synthase, belonging to the thioredoxin-like superfamily. *J Biol Chem* **283**: 792–801.
- Morrow DJ, Hill KE, Burk RF, Nammour TM, Badr KF, Roberts LJ (1990). A series of prostaglandin F<sub>2</sub>-like compounds are produced *in vivo* in humans by a non-cyclooxygenase, free radical-catalysed mechanism. *Proc Natl Acad Sci USA* **87**: 9383–9387.
- Morrow JD, Minton TA, Mukundan CR, Campbell MD, Zackert WE, Daniel VC *et al.* (1994). Free radical-induced generation of isoprostanes *in vivo*. Evidence for the formation of D-ring and E-ring isoprostanes. *J Biol Chem* **269**: 4317–4326.
- Müller B, Schneider J, Wilsmann K, Lintz W, Flohé L (1983). Role of renin release in the hemodynamic, renal and dipsogenic actions of the prostacyclin analogue CG 4203 in conscious rats. *Prostaglandins Leukot Med* **11**: 361–372.
- Murase A, Okumura T, Sakakibara A, Tonai-Kachi H, Nakao S, Takada J (2008a). Effect of prostanoid EP<sub>4</sub> receptor antagonist, CJ 042,794 in rat models of pain and inflammation. *Eur J Pharmacol* **580**: 116–121.
- Murase A, Taniguchi Y, Tonai-Kachi H, Nakao S, Takada J (2008b). *In vitro* pharmacological characterization of CJ-042794, a novel, potent, and selective prostaglandin EP<sub>4</sub> receptor antagonist. *Life Sci* **82**: 226–232.
- Murata T, Ushikubi F, Matsuoka T, Hirata M, Yamasaki A, Sugimoto Y *et al.* (1997). Altered pain perception and inflammatory response in mice lacking prostacyclin receptor. *Nature* **388**: 678–682.
- Murray JJ, Tonnel AB, Brash AR, Roberts LJ, Gosset P, Workman R *et al.* (1986). Release of prostaglandin D<sub>2</sub> into human airways during acute antigen challenge. *N Engl J Med* **315**: 800–804.
- Mutoh M, Watanabe K, Kitamura T, Shoji Y, Takahashi M, Kawamori T *et al.* (2002). Involvement of prostaglandin E receptor subtype EP<sub>4</sub> in colon carcinogenesis. *Cancer Res* **62**: 28–32.
- Mutoh M, Takahashi M, Wakabayashi K (2006). Roles of prostanoids in colon carcinogenesis and their potential targeting for cancer chemoprevention. *Curr Pharm Des* **12**: 2375–2382.
- Naclerio RM, Meier HL, Kagey-Sobotka A, Adkinson NF Jr, Norman DA, Meyers PS *et al.* (1983). Mediator release after nasal airway challenge with allergen. *Am Rev Respir Dis* **128**: 597–602.
- Nagai H, Takeda H, Yamaguchi S, Tanaka H, Matsuo A, Inagaki N (1995). The effect of a thromboxane A<sub>2</sub> receptor antagonist BAY-u3405 on experimental allergic reactions. *Prostaglandins* **50**: 75–87.
- Naganawa A, Matsui T, Ima M, Yoshida K, Tsuruta H, Yamamoto S *et al.* (2006). Optimization of sulfonamide derivatives as highly selective EP<sub>1</sub> receptor antagonists. *Bioorg Med Chem* **14**: 7774–7789.
- Nagata K, Hirai H (2003). The second PGD<sub>2</sub> receptor CRTH2: structure, properties, and functions in leukocytes. *Prostaglandins Leukot Essent Fatty Acids* **69**: 169–177.
- Nagata K, Hirai H, Tanaka K, Ogawa K, Aso T, Sugamura K *et al.* (1999a). CRTH2, an orphan receptor of T-helper-2-cells, is expressed on basophils and eosinophils and responds to mast cell-derived factor(s). *FEBS Lett* **459**: 195–199.
- Nagata K, Tanaka K, Ogawa K, Kemmotsu K, Imai T, Yoshie O *et al.* (1999b). Selective expression of a novel surface molecule by human Th2 cells *in vivo*. *J Immunol* **162**: 1278–1286.
- Naka M, Mais DE, Morinelli TA, Hamanaka N, Oatis JE Jr, Halushka PV (1992). 7-[(1R,2S,3S,5R)-6,6-dimethyl-3-(4-iodobenzesulfonylamino)bicyclo[3.1.1]hept-2-yl]-5(Z)-heptenoic acid: a novel high-affinity radiolabeled antagonist for platelet thromboxane A<sub>2</sub>/prostaglandin H<sub>2</sub> receptors. *J Pharmacol Exp Ther* **262**: 632–637.
- Nakae K, Saito K, Iino T, Yamamoto N, Wakabayashi M, Yoshikawa S *et al.* (2005). A prostacyclin receptor antagonist inhibits the sensitized release of substance P from rat sensory neurones. *J Pharmacol Exp Ther* **315**: 1136–1142.
- Nakahata N, Sato K, Abe MT, Nakanishi H (1990). ONO NT-1<sub>2</sub>6 is a potent and selective thromboxane A<sub>2</sub> antagonist in human astrocytoma cells. *Eur J Pharmacol* **184**: 233–238.
- Nakao K, Murase A, Ohshiro H, Okumura T, Taniguchi K, Murata Y *et al.* (2007). CJ-023, 23, a novel, potent and selective prostaglandin EP<sub>4</sub> receptor antagonist with anti-hyperalgesic properties. *J Pharm Exp Ther* **322**: 686–694.
- Nakayama Y, Omote K, Namiki A (2002). Role of prostaglandin receptor EP<sub>1</sub> in the spinal dorsal horn in carrageenan-induced inflammatory pain. *Anesthesiology* **97**: 1254–1262.
- Nakayama Y, Omote K, Kawamata T, Namiki A (2004). Role of prostaglandin receptor subtype EP<sub>1</sub> in prostaglandin E<sub>2</sub>-induced nociceptive transmission in the rat spinal dorsal horn. *Brain Res* **1010**: 62–68.
- Namba T, Oida H, Sugimoto Y, Kakizuka A, Negishi M, Ichikawa A *et al.* (1994). cDNA Cloning of a mouse prostacyclin receptor. *J Biol Chem* **269**: 9986–9992.
- Narisada M, Ohtani M, Watanabe F, Uchida K, Arita H, Doteuchi M *et al.* (1988). Synthesis and *in vitro* activity of various derivatives of a novel thromboxane receptor antagonist, (±)-(5Z)-7-[3-endo-[(phenylsulfonyl)amino]bicyclo[2.2.1]hept-2-exo-yl]heptenoic acid. *J Med Chem* **31**: 1847–1854.
- Narumiya S (2003). Prostanoids in immunity: roles revealed by mice deficient in their receptors. *Life Sci* **74**: 391–395.
- Narumiya S, Fitzgerald GA (2001). Genetic and pharmacological analysis of prostanoid receptor function. *J Clin Invest* **108**: 25–30.
- Narumiya S, Toda N (1985). Different responsiveness of prostaglandin D<sub>2</sub>-sensitive systems to prostaglandin D<sub>2</sub> and its analogues. *Br J Pharmacol* **85**: 367–375.
- Narumiya S, Okuma S, Ushikubi F (1986). Binding of a radioiodinated 13-azapinane thromboxane antagonist to platelets: correlation with antiaggregatory activity in different species. *Br J Pharmacol* **88**: 323–331.
- Narumiya S, Sugimoto Y, Ushikubi B (1999). Prostanoid receptors: structure, properties and functions. *Physiol Rev* **79**: 1193–1226.
- Nasrallah R, Hébert RL (2005). Prostaglandin signaling in the kidney: implications for health and disease. *Am J Physiol Renal Physiol* **289**: F235–F246.
- Nataraj C, Thomas DW, Tilley SL, Nguyen MT, Mannon R, Koller BH *et al.* (2001). Receptors for prostaglandin E<sub>2</sub> that regulate cellular immune responses in the mouse. *J Clin Invest* **108**: 1229–1235.
- Nicolaou KC, Magolda RL, Smith JB, Aharony D, Smith EF, Lefer AM (1979). Synthesis and biological properties of pinane-thromboxane A<sub>2</sub>, a selective inhibitor of coronary artery constriction, platelet aggregation, and thromboxane formation. *Proc Natl Acad Sci USA* **76**: 2566–2570.
- Nishigaki N, Negishi M, Honda A, Sugimoto Y, Namba T, Narumiya S *et al.* (1995). Identification of prostaglandin E receptor 'EP<sub>2</sub>' cloned from mouse mastocytoma cells as EP<sub>4</sub>. *FEBS Lett* **364**: 339–341.
- Nigam S, Zakrzewicz A, Eskafi S, Roscher A (1992). Clinical significance of prostacyclin and thromboxane in cancer of the female breast and genital tract. *Cancer Metastasis Rev* **11**: 411–420.
- Noguchi K, Iwasaki K, Shitashiqe M, Murato S, Ishikawa I (1999). Prostaglandin E<sub>2</sub> downregulates interferon-gamma-induced intercellular adhesion molecule-1 expression via EP<sub>2</sub> receptors in human gingival fibroblasts. *Inflammation* **23**: 481–493.
- Norel X, Walch L, Labat C, Gascard JP, Dulmet E, Brink C (1999). Prostanoid receptors involved in the relaxation of human bronchial preparations. *Br J Pharmacol* **126**: 867–872.
- Norel X, De Montpreville V, Brink C (2004). Vasoconstriction induced by activation of EP<sub>1</sub> and EP<sub>3</sub> receptors in human lung: effects of ONO-AE-248, ONO-DI-004, ONO-8711 or ONO-8713. *Prostaglandins Other Lipid Mediat* **74**: 101–112.
- Norman P, Cuthbert NJ, McKenniff MG, Gardiner PJ (1992). The thromboxane receptors of rat and guinea-pig lung. *Eur J Pharmacol* **229**: 171–178.
- O'Connell M, Zeller W, Burgeson J, Mishra RK, Ramirez J, Kiselyov AS *et al.* (2009). Peri-substituted hexahydro-indolones as novel, potent and selective human EP<sub>3</sub> receptor antagonists. *Bioorg Med Chem Lett* **19**: 778–782.

- Ogletree ML, Allen GT (1992). Interspecies differences in thromboxane receptors: studies with thromboxane receptor antagonists in rat and guinea pig smooth muscles. *J Pharmacol Exp Ther* **260**: 789–794.
- Ogletree ML, Harris DN, Greenberg R, Haslanger MF, Nakane M (1985). Pharmacological actions of SQ 29,548, a novel selective thromboxane antagonist. *J Pharmacol Exp Ther* **234**: 435–441.
- Ogletree ML, Harris DN, Schumacher WA, Webb ML, Mistra RN (1993). Pharmacological profile of BMS 180,291: a potent, long-acting, orally active thromboxane  $A_2$ /prostaglandin endoperoxide receptor antagonist. *J Pharmacol Exp Ther* **264**: 570–578.
- Ohnishi A, Shimamoto C, Katsu K, Ito S, Imai Y, Nakahara T (2001). EP<sub>1</sub> and EP<sub>4</sub> receptors mediate exocytosis evoked by prostaglandin E<sub>2</sub> in guinea-pig antral mucous cells. *Exp Physiol* **86**: 451–460.
- Oka T, Aou S, Hori T (1994). Intracerebroventricular injection of prostaglandin E<sub>2</sub> induces thermal hyperalgesia in rats: the possible involvement of EP<sub>3</sub> receptors. *Brian Res* **663**: 287–292.
- Okumura T, Murata Y, Taniguchi K, Murase A, Nii A (2008). Effects of the selective EP<sub>4</sub> antagonist, CJ-023,423 on chronic inflammation and bone destruction in rat adjuvant-induced arthritis. *J Pharm Pharmacol* **60**: 723–730.
- Oliva P, Berrino L, De Novellis V, Palazzo E, Marabesse I, Siniscalco D *et al.* (2006). Role of periaqueductal grey prostaglandin receptors in formalin-induced hyperalgesia. *Eur J Pharmacol* **530**: 40–47.
- Olson DM (2005). The promise of prostaglandins: have they fulfilled their potential as therapeutic targets for the delay of preterm birth? *J Soc Gynecol Invest* **12**: 466–478.
- Omote K, Kawamata T, Nakayama Y, Kawamata M, Hazama K, Namiki A (2001). The effects of peripheral administration of a novel selective antagonist for prostaglandin E receptor subtype EP<sub>1</sub>, ONO-8711, in a rat model of postoperative pain. *Anesth Analg* **92**: 233–238.
- Omote K, Yamamoto H, Kawamata T, Nakayama Y, Namiki A (2002). The effects of intrathecal administration of an antagonist for prostaglandin E receptor subtype EP<sub>1</sub> on mechanical and thermal hyperalgesia in a rat model of postoperative pain. *Anesth Analg* **95**: 1708–1712.
- O'Reilly DD, Loomis CW (2007). Spinal prostaglandins facilitate exaggerated A- and C-fiber-mediated reflex responses and are critical to the development of allodynia early after L5–L6 spinal nerve ligation. *Anesthesiology* **106**: 795–805.
- Palea S, Toson G, Pietra C, Trist DG, Artibani W, Romano O *et al.* (1998). Pharmacological characterization of thromboxane and prostanoid receptors in human isolated urinary bladder. *Br J Pharmacol* **124**: 865–872.
- Paolini JF, Mitchel YB, Reyes R, Kher U, Lai E, Watson DJ *et al.* (2008). Effects of laropiprant on nicotinic acid-induced flushing in patients with dyslipidemia. *Am J Cardiol* **101**: 625–630.
- Paralkar VM, Borovecki F, Ke HZ, Cameron KO, Lefker B, Grasser WA *et al.* (2003). An EP<sub>2</sub> receptor-selective prostaglandin E<sub>2</sub> agonist induces bone healing. *Proc Natl Acad Sci USA* **100**: 6736–6740.
- Patrignani P, Filabozzi P, Patrono C (1982). Selective cumulative inhibition of platelet thromboxane production by low-dose aspirin in healthy subjects. *J Clin Invest* **69**: 1366–1372.
- Patrono C (1990). Thromboxane synthesis inhibitors and receptor antagonists. *Thromb Res Suppl* **11**: 15–23.
- Patscheke H (1990). Thromboxane  $A_2$ /prostaglandin H<sub>2</sub> receptor antagonists, a new therapeutic principle. *Stroke* **21**: 139–142.
- Peri K, Polyak F, Lubell W, Thouin E, Chemtob S (2006). Peptides and peptidomimetics useful for inhibiting the activity of prostaglandin F<sub>2 $\alpha$</sub>  receptor. US 2006/0211626, Quebec.
- Peters SP, Schleimer RP, Kagey-Sobotka A, Naclerio RM, MacGlashan DW, Schulman ES *et al.* (1982). The role of prostaglandin D<sub>2</sub> in IgE-mediated reactions in man. *Trans Assoc Am Physicians* **95**: 221–228.
- Pettipher R (2008). The roles of the prostaglandin D<sub>2</sub> receptors DP<sub>1</sub> and CRTH2 in promoting allergic responses. *Br J Pharmacol* **153**: S191–S199.
- Pettipher R, Hansel TT, Armer R (2007). Antagonism of the prostaglandin D<sub>2</sub> receptors DP<sub>1</sub> and CRTH2 as an approach to treat allergic diseases. *Nat Rev Drug Discov* **6**: 313–325.
- Pfister SL, Pratt PE, Kurian J, Campbell WB (2004). Glibenclamide inhibits thromboxane-mediated vasoconstriction by thromboxane receptor blockade. *Vascul Pharmacol* **40**: 285–292.
- Pierce KL, Regan JW (1998). Prostanoid receptor heterogeneity through alternative mRNA splicing. *Life Sci* **62**: 1479–1483.
- Pollock WK, Armstrong RA, Brydon LJ, Jones RL, MacIntyre DE (1984). Thromboxane-induced phosphatidate formation in human platelets. Relationship to receptor occupancy and to changes in cytosolic free calcium. *Biochem J* **219**: 833–842.
- Pönicke K, Giessler C, Grapow M, Heinroth-Hoffman I, Becker K, Osten B *et al.* (2000). FP-receptor mediated trophic effects of prostanoids in rat ventricular cardiomyocytes. *Br J Pharmacol* **129**: 1723–1731.
- Pulichino AM, Rowland S, Wu T, Clark P, Xu D, Mathieu MC *et al.* (2006). Prostacyclin antagonism reduces pain and inflammation in rodent models of hyperalgesia and chronic arthritis. *J Pharmacol Exp Ther* **319**: 1043–1050.
- Qian YM, Jones RL, Chan KM, Stock AI, Ho JK (1994). Potent contractile actions of prostanoid EP<sub>3</sub>-receptor agonists on human isolated pulmonary artery. *Br J Pharmacol* **113**: 369–374.
- Racké K, Bähring J, Langer C, Bräutigam M, Wessler I (1992). Prostanoids inhibit release of endogenous norepinephrine from rat isolated trachea. *Am Rev Respir Dis* **146**: 1182–1186.
- Raisz LG (2006). Anabolic effect of prostaglandins. *Clin Rev Bone Min* **4**: 123–128.
- Rampton DS, Collins CE (1993). Thromboxane in inflammatory bowel disease – pathogenic and therapeutic implications. *Aliment Pharmacol Ther* **7**: 357–367.
- Randall MJ, Parry MJ, Hawkeswood E, Cross PE, Dickinson RP (1981). UK-37 248, a novel, selective thromboxane synthetase inhibitor with platelet anti-aggregatory and anti-thrombotic activity. *Thromb Res* **23**: 145–162.
- Rangachari PK, Betti PA (1993). Biological activity of metabolites of PGD<sub>2</sub> on canine proximal colon. *Am J Physiol* **264**: G886–G894.
- Rangachari PK, Betti P-A, Prior ET, Roberts LJ (1995). Effects of a selective DP receptor agonist (BW 245C) and antagonist (BW A868C) on the canine colonic epithelium: an argument for a different DP receptor? *J Pharmacol Exp Ther* **275**: 611–617.
- Raychowdhury MK, Yukawa M, Collins LJ, McGrail SH, Kent KC, Ware JA (1994). Alternative splicing produces a divergent cytoplasmic tail in the human endothelial thromboxane A<sub>2</sub> receptor. *J Biol Chem* **269**: 19256–19261.
- Regan JW, Bailey TJ, Pepper DJ, Pierce KL, Bogardus AM, Donello JE *et al.* (1994). Cloning of a novel human prostaglandin receptor with characteristics of the pharmacologically defined EP<sub>2</sub> subtype. *Mol Pharmacol* **46**: 213–220.
- Reilly IA, FitzGerald GA (1987). Inhibition of thromboxane formation in vivo and ex vivo: implications for therapy with platelet inhibitory drugs. *Blood* **69**: 180–186.
- Rolin S, Dogné J-M, Michaux C, Delarge J, Masereel B (2001). Activity of a novel dual thromboxane A<sub>2</sub>-receptor antagonist and thromboxane synthase inhibitor (BM-573) on platelet function and isolated smooth muscles. *Prostaglandins Leukot Essent Fatty Acids* **65**: 67–72.
- Rolin S, Masereel B, Dogné J-M (2006). Prostanoids as pharmacological targets in COPD and asthma. *Eur J Pharmacol* **533**: 89–100.
- Ross RA, Craib SJ, Stevenson LA, Pertwee RG, Henderson A, Toole J *et al.* (2002). Pharmacological characterization of the anandamide cyclooxygenase metabolite: prostaglandin E<sub>2</sub> ethanolamide. *J Pharmacol Exp Ther* **301**: 900–907.
- Ruel R, Lacombe P, Abramovitz M, Godbout C, Lamontagne S, Rochette C *et al.* (1999). New class of biphenylene dibenzazocinones as potent ligands for the human EP<sub>1</sub> prostanoid receptor. *Bioorg Med Chem Lett* **9**: 2699–2704.
- Saito O, Guan Y, Qi Z, Davis LS, Koernhoff M, Sugimoto Y *et al.* (2003).

- Expression of the prostaglandin F receptor (FP) gene along the mouse genitourinary tract. *Am J Physiol Renal Physiol* **284**: F1164–F1170.
- Sametz W, Hennerbichler S, Glaser S, Wintersteiger R, Juan H (2000). Characterization of prostanoid receptors mediating actions of the isoprostanes, 8-iso-PGE<sub>2</sub> and 8-iso-PGF<sub>2α</sub>, in some isolated smooth muscle preparations. *Br J Pharmacol* **130**: 1903–1910.
- Sanchez-Ramoz L, Kaunitz AM, Wears RL, Delke I, Gaudier FL (1997). Misoprostol for cervical ripening and labor induction: a meta-analysis. *Obstet Gynecol* **89**: 633–642.
- Sanner JH (1969). Antagonism of prostaglandin E<sub>2</sub> by 1-acetyl-2-(8-chloro - 10, 11 - dihydrobiphenyl [1, 4] oxazepine - 10-carbonyl) hydrazine (SC-19220). *Arch Int Pharmacodyn Ther* **180**: 46–56.
- Sarkar S, Hobson AR, Hughes A, Growcott J, Woolf CJ, Thompson DG *et al.* (2003). The prostaglandin E<sub>2</sub> receptor-1 (EP1) mediates acid-induced visceral pain hypersensitivity in humans. *Gastroenterology* **124**: 18–25.
- Sawyer N, Cauchon E, Chateaufneuf A, Cruz RP, Nicholson DW, Metters KM *et al.* (2002). Molecular pharmacology of the human prostaglandin D<sub>2</sub> receptor, CRTH2. *Br J Pharmacol* **137**: 1163–1172.
- Schlemper V, Medeiros R, Ferreira J, Campos MM, Calixto JB (2005). Mechanisms underlying the relaxation response induced by bradykinin in the epithelium-intact guinea-pig trachea in vitro. *Br J Pharmacol* **145**: 740–750.
- Schaaf TK, Hess HJ (1979). Synthesis and biological activity of carboxyl-terminus modified prostaglandin analogues. *J Med Chem* **22**: 1340–1346.
- Seiler SM, Brassard CL, Federici ME, Romini J, Meanwell NA (1997). [3-[4-(4,5-Diphenyl-2-oxazolyl)-5-oxazolyl]phenoxy]acetic acid (BMJ 45778) is a potent non-prostanoid prostacyclin partial agonist: effects on platelet aggregation, adenylyl cyclase, cAMP levels, protein kinase, and iloprost binding. *Prostaglandins* **53**: 21–35.
- Senchyna M, Crankshaw DJ (1996). Characterization of the prostanoid TP receptor population in human nonpregnant myometrium. *J Pharmacol Exp Ther* **279**: 262–270.
- Senior J, Sangha R, Baxter GS, Marshall K, Clayton JK (1992). *In vitro* characterization of prostanoid FP-, DP-, IP- and TP-receptors on the non-pregnant human myometrium. *Br J Pharmacol* **107**: 215–221.
- Senior J, Marshall K, Sangha R, Clayton JK (1993). *In vitro* characterization of prostanoid receptors on human myometrium at term pregnancy. *Br J Pharmacol* **108**: 501–506.
- Sharif NA, Crider JY, Davis TL (2000). AL-3138 antagonizes FP prostanoid receptor-mediated inositol phosphates generation: comparison with some purported FP antagonists. *J Pharm Pharmacol* **52**: 1229–1239.
- Sharif NA, Senchyna M, Xu SX (2002). Pharmacological and molecular biological (RT-PCR) characterization of functional TP prostanoid receptors in immortalized human non-pigmented ciliary epithelial cells. *J Ocul Pharmacol Ther* **18**: 141–162.
- Sharif NA, Kaddour-Djebbar I, Abdel-Latif A (2008). Cat iris sphincter smooth-muscle contraction: comparison of FP-class prostaglandin analog agonist activities. *J Ocul Pharmacol Ther* **24**: 152–163.
- Shaw JS, Barker AJ, Breault G, Griffen EJ, Heapy CG, McLaughlin S (1999). ZM325082: a potent and selective antagonist at the prostanoid EP1 receptor. 9th World Congress on Pain, Vienna, 1999. Abstract Book, p. 272.
- Shiraishi Y, Asano K, Nakajima T, Oguma T, Suzuki Y, Shiomi T *et al.* (2005). Prostaglandin D<sub>2</sub>-induced eosinophilic airway inflammation is mediated by CRTH2 receptor. *J Pharmacol Exp Ther* **312**: 954–960.
- Singh J, Zeller W, Zhou N, Hategen G, Mishra R, Polozov A *et al.* (2009). Antagonists of the EP<sub>3</sub> receptor for prostaglandin E<sub>2</sub> are novel antiplatelet agents that do not prolong bleeding. *ACS Chem Biol* **4**: 115–126.
- Smith GCS, Coleman RA, McGrath JC (1994). Characterization of dilator prostanoid receptors in the fetal rabbit ductus arteriosus. *J Pharmacol Exp Ther* **271**: 390–396.
- Smith JB (1982). Effect of thromboxane synthetase inhibitors on platelet function: enhancement by inhibition of phosphodiesterase. *Thromb Res* **28**: 477–485.
- Solomon SD, McMurray JVV, Pfeffer MA, Wittes J, Fowler R, Finn P *et al.* (2005). Cardiovascular risk associated with celecoxib in a clinical trial for colorectal adenoma prevention. *N Engl J Med* **352**: 1071–1080.
- Soyka R, Heckel A, Nickl J, Eisert W, Muller TH, Weisenberger H (1993). 6,6-Disubstituted hex-5-enoic acid derivatives as combined thromboxane A<sub>2</sub> receptor antagonists and synthase inhibitors. *J Med Chem* **37**: 26–39.
- Spada CS, Krauss AH, Woodward DF, Chen J, Protzman CE, Nieves AL *et al.* (2005). Bimatoprost and prostaglandin F<sub>2α</sub> selectivity stimulate intracellular calcium signaling in different cat iris sphincter cells. *Exp Eye Res* **80**: 135–145.
- Spik I, Brenuchon C, Angeli V, Staumont D, Fleury S, Capron M *et al.* (2005). Activation of the prostaglandin D<sub>2</sub> receptor DP<sub>2</sub>/CRTH2 increases allergic inflammation in mouse. *J Immunol* **174**: 3703–3708.
- Stanke F, Cracowski JL, Chavanon O, Magne JL, Blin D, Bessard G *et al.* (1998). Glibenclamide inhibits thromboxane A<sub>2</sub>-induced contraction in human internal mammary artery and saphenous vein. *Eur J Pharmacol* **341**: 65–71.
- Stefany D, Harris KJ, Gillespy TA, Gardner CJ, Aguiar JC (2007). 2,6-Substituted-4-monosubstituted aminopyrimidine as prostaglandin D<sub>2</sub> receptor antagonists. WO2007/121280, Sanofi-Aventis.
- Stinger RB, Fitzpatrick TM, Corey EJ, Ramwell PW, Rose JC, Kot PA (1982). Selective antagonism of prostaglandin F<sub>2α</sub>-mediated vascular responses by N-dimethylamino substitution of prostaglandin F<sub>2α</sub>. *J Pharmacol Exp Ther* **220**: 521–525.
- Stock JL, Shinjo K, Burkhardt J, Roach M, Taniguchi K, Ishikawa T *et al.* (2001). The prostaglandin E<sub>2</sub> EP1 receptor mediates pain perception and regulates blood pressure. *J Clin Invest* **107**: 325–331.
- Stubbs VE, Schratl P, Hartnell A, Williams TJ, Peskar BA, Heinemann A *et al.* (2002). Indomethacin causes prostaglandin D<sub>2</sub>-like and eotaxin-like selective responses in eosinophils and basophils. *J Biol Chem* **277**: 26012–26020.
- Sturino CF, Lachance N, Boyd M, Berthelette C, Labelle M, Li L *et al.* (2006). Identification of an indole series of prostaglandin D<sub>2</sub> receptor antagonists. *Bioorg Med Chem Lett* **16**: 3043–3048.
- Sturino CF, O'Neill G, Lachance N, Boyd M, Berthelette C, Labelle M *et al.* (2007). Discovery of a potent and selective prostaglandin D<sub>2</sub> receptor antagonist, [(3R)-4-(4-chloro-benzyl)-7-fluoro-5-(methylsulfonyl)-1,2,3,4-tetrahydrocyclopenta[b] indol-3-yl]-acetic acid (MK-0524). *J Med Chem* **50**: 794–806.
- Su X, Lashinger ES, Leon LA, Hoffman BE, Hieble JP, Gardner SD *et al.* (2008a). An excitatory role for peripheral EP<sub>3</sub> receptors in bladder afferent function. *Am J Physiol Renal Physiol* **295**: F585–F594.
- Su X, Leon LA, Wu CW, Morrow DM, Jaworski JP, Hieble JP *et al.* (2008b). Modulation of bladder function by prostaglandin EP<sub>3</sub> receptors in the central nervous system. *Am J Physiol Renal Physiol* **295**: F984–F994.
- Sugimoto H, Shichijo M, Iino T, Manabe Y, Watanabe A, Shimazaki M *et al.* (2003). An orally-bioavailable small molecule antagonist of CRTH2, ramatroban (BAY u3405), inhibits PGD<sub>2</sub>-induced eosinophil migration in vitro. *J Pharmacol Exp Ther* **305**: 347–352.
- Sugimoto Y, Yamasaki A, Segi E, Tsuboi K, Aze Y, Nishimura T *et al.* (1997). Failure of parturition in mice lacking the prostaglandin F-receptor. *Science* **277**: 681–683.
- Suzawa T, Miyaura C, Inada M, Maruyama T, Sugimoto Y, Ushikubi F *et al.* (2000). The role of prostaglandin E receptor subtypes (EP<sub>1</sub>, EP<sub>2</sub>, EP<sub>3</sub>, and EP<sub>4</sub>) in bone resorption; an analysis using specific agonists for the respective EPs. *Endocrinology* **141**: 1554–1559.
- Svensson CI, Yaksh TL (2002). The spinal phospholipase-cyclooxygenase-prostanoid cascade in nociceptive processing. *Annu Rev Pharmacol Toxicol* **42**: 553–583.



- Swayne GT, Maguire J, Dolan J, Raval P, Dane G, Greener M *et al.* (1988). Evidence for homogeneity of thromboxane A<sub>2</sub> receptor using structurally different antagonists. *Eur J Pharmacol* **152**: 311–319.
- Syriatowicz JP, Hu D, Walker JS, Tracey DJ (1999). Hyperalgesia due to nerve injury: role of prostaglandins. *Neuroscience* **94**: 587–594.
- Takahara K, Murray R, Fitzgerald GA, Fitzgerald DJ (1990). The response to thromboxane A<sub>2</sub> analogues in human platelets. Discrimination of two binding sites linked to distinct effector systems. *J Biol Chem* **265**: 6836–6844.
- Takasaki I, Nojima H, Shiraki K, Sugimoto Y, Ichikawa A, Ushikubi F *et al.* (2005). Involvement of cyclooxygenase-2 and EP<sub>3</sub> prostaglandin receptor in acute herpetic but not postherpetic pain in mice. *Neuropharmacology* **49**: 283–292.
- Takayama K, Garcia-Cardena G, Sukhova GK, Comander J, Gimbrone MA Jr, Libby P (2002). Prostaglandin E<sub>2</sub> suppresses chemokine production in human macrophages through the EP<sub>4</sub> receptor. *J Biol Chem* **277**: 44147–44154.
- Tanaka T, Fukuta Y, Higashino R, Sato R, Nomura Y, Fukuda Y *et al.* (1998). Antiplatelet effect of Z-335, a new orally active and long-lasting thromboxane receptor antagonist. *Eur J Pharmacol* **357**: 53–60.
- Tanaka K, Ogawa K, Sugamura K, Nakamura M, Takano S, Nagata K (2000). Differential production of prostaglandin D<sub>2</sub> by human helper T cell subsets. *J Immunol* **164**: 2277–2280.
- Tani K, Naganawa A, Ishida A, Sagawa K, Harada H, Ogawa M *et al.* (2002). Development of a highly selective EP<sub>2</sub>-receptor agonist. Part 1: identification of 16-hydroxy-17,17-trimethylene PGE<sub>2</sub> derivatives. *Bioorg Med Chem Lett* **10**: 1093–1106.
- Teixeira MM, al-Rashed S, Rossi AG, Hellewell PG (1997). Characterization of the prostanoid receptors mediating inhibition of PAF-induced aggregation of guinea-pig eosinophils. *Br J Pharmacol* **121**: 77–82.
- Terada N, Yamakoshi T, Hasegawa M, Tanikawa H, Maesako K, Ishikawa K *et al.* (1998). The effect of ramatroban (BAY u3405), a thromboxane A<sub>2</sub> receptor antagonist, on nasal cavity volume and minimum cross-sectional area and nasal mucosal hemodynamics after nasal mucosal allergen challenge in patients with perennial allergic rhinitis. *Acta Otolaryngol Suppl* **537**: 32–37.
- Theiner G, Gessner A, Lutz MB (2006). The mast cell mediator PGD<sub>2</sub> suppresses IL-12 release by dendritic cells leading to Th2 polarized immune responses *in vivo*. *Immunobiology* **211**: 463–472.
- Tober KL, Wilgus TA, Kusewitt DF, Thomas-Ahner JM, Maruyama T, Oberyszyn TM (2006). Importance of the EP<sub>1</sub>-receptor in cutaneous UVB-induced inflammation and tumor development. *J Invest Dermatol* **126**: 205–211.
- Tsuri T, Honma T, Hiramatsu Y, Okada T, Hashizume H, Mitsumori S *et al.* (1997). Bicyclo[2.2.1]heptane and 6,6-dimethylbicyclo[3.1.1]heptane derivatives: orally active, potent, and selective prostaglandin D<sub>2</sub> receptor antagonists. *J Med Chem* **40**: 3504–3507.
- Torisu K, Kobayashi K, Iwahashi M, Egashira H, Nakai Y, Okada Y *et al.* (2004a). Discovery of new chemical leads for prostaglandin D<sub>2</sub> receptor antagonists. *Bioorg Med Chem Lett* **14**: 4557–4562.
- Torisu K, Kobayashi K, Iwahashi M, Nakai Y, Onoda T, Nagase T *et al.* (2004b). Development of prostaglandin D<sub>2</sub> receptor antagonist: discovery of highly potent antagonists. *Bioorg Med Chem* **12**: 4685–4700.
- Torisu K, Kobayashi K, Iwahashi M, Nakai Y, Onoda T, Nagase T *et al.* (2004c). Discovery of a new class of potent, selective, and orally active prostaglandin D<sub>2</sub> receptor antagonists. *Bioorg Med Chem* **12**: 5361–5378.
- Torisu K, Kobayashi K, Iwahashi M, Nakai Y, Onoda T, Nagase T *et al.* (2004d). Discovery of orally active prostaglandin D<sub>2</sub> receptor antagonists. *Bioorg Med Chem Lett* **14**: 4891–4895.
- Torisu K, Kobayashi K, Iwahashi M, Egashira H, Nakai Y, Okada Y *et al.* (2005). Development of a prostaglandin D<sub>2</sub> receptor antagonist: discovery of a new chemical lead. *Eur J Med Chem* **40**: 505–519.
- Tymkewycz PM, Jones RL, Wilson NH, Marr CG (1991). Heterogeneity of thromboxane A<sub>2</sub> (TP-) receptors: evidence from antagonist but not agonist potency measurements. *Br J Pharmacol* **102**: 607–614.
- Uda R, Horiguchi S, Ito S, Hyodo M, Hayashi O (1990). Nociceptive effects induced by intrathecal administration of prostaglandin D<sub>2</sub>, E<sub>2</sub>, or F<sub>2α</sub> to conscious mice. *Brain Res* **510**: 26–32.
- Ueno A, Matsumoto H, Naraba H, Ikeda Y, Ushikubi F, Matsuoka T *et al.* (2001). Major roles of prostanoid receptors IP and EP<sub>3</sub> in endotoxin-induced enhancement of pain perception. *Biochem Pharmacol* **62**: 157–160.
- Uller L, Mathiesen JM, Alenmyr L, Korsgren M, Ulven T, Hogberg T *et al.* (2007). Antagonism of the prostaglandin D<sub>2</sub> receptor CRTH2 attenuates asthma pathology in mouse eosinophilic airway inflammation. *Respir Res* **8** (16): 1–10.
- Ulven T, Kostenis E (2005). Minor structural modifications convert the dual TP/CRTH2 antagonist ramatroban into a highly selective and potent CRTH2 antagonist. *J Med Chem* **48**: 897–900.
- Ulven T, Kostenis E (2006). Targeting the prostaglandin D<sub>2</sub> receptors DP and CRTH2 for treatment of inflammation. *Curr Top Med Chem* **6**: 1427–1444.
- Umemura K, Nakashima M, Saruta T (1997). Thromboxane A<sub>2</sub> synthetase inhibition suppresses cough induced by angiotensin converting enzyme inhibitors. *Life Sci* **60**: 1583–1588.
- Ushikubi F, Segi E, Sugimoto Y, Murata T, Matsuoka T, Kobayashi T *et al.* (1998). Impaired febrile response in mice lacking the prostaglandin E receptor subtype EP<sub>3</sub>. *Nature* **395**: 281–284.
- Van Hecken A, Depre M, De LI, Thach C, Oeyen M, Van Effen J *et al.* (2007). The effect of MK-0524, a prostaglandin D<sub>2</sub> receptor antagonist, on prostaglandin D<sub>2</sub>-induced nasal airway obstruction in healthy volunteers. *Eur J Clin Pharmacol* **63**: 135–141.
- Vermynen J, Defreyn G, Carreras LO, Machin SJ, Van Schaeren J, Verstraete M (1981). Thromboxane synthetase inhibition as anti-thrombotic strategy. *Lancet* **1** (8229): 1073–1075.
- Veza R, Roberti R, Nenci GG, Gressele P (1993). Prostaglandin E<sub>2</sub> potentiates platelet aggregation by priming protein kinase C. *Blood* **82**: 2704–2713.
- Walch L, Labat C, Gascard JP, de Montpreville V, Brink C, Norel X (1999). Prostanoid receptors involved in the relaxation of human pulmonary vessels. *Br J Pharmacol* **126**: 859–866.
- Walch L, de Montpreville V, Brink C, Norel X (2001). Prostanoid EP<sub>1</sub>- and TP-receptors involved in the contraction of human pulmonary veins. *Br J Pharmacol* **134**: 1671–1678.
- Walch L, Clavarini E, Morris PL (2003). Prostaglandin (PG) FP and EP<sub>1</sub> receptors mediate PGF<sub>2α</sub> and PGE<sub>2</sub> regulation of interleukin-1β expression in Leydig cell progenitors. *Endocrinology* **144**: 1284–1291.
- Wan Z, Woodward DF, Cornell CL, Fliri HG, Martos JS, Pettit SN *et al.* (2007). Bimatoprost, prostamide activity, and conventional drainage. *Invest Ophthalmol Vis Sci* **48**: 4107–4115.
- Wang Z, Beaulieu C, Dufresne C, Guay D, Leblanc Y (2002). Dihydropyrrrolo[1,2-a]indole and tetrahydropyrido[1,2-a]indole derivatives as prostaglandin D<sub>2</sub> receptor antagonists. WO2002/094830, Merck Frosst Canada & Co.
- Wardle EN (1999). Thromboxane in glomerulonephritis: what about therapy? *Am J Ther* **6**: 111–114.
- Watanabe K, Kawamori T, Nakatsugi S, Ohta T, Ohuchida S, Yamamoto H *et al.* (1999). Role of the prostaglandin E receptor subtype EP<sub>1</sub> in colon carcinogenesis. *Cancer Res* **59**: 5093–5096.
- Watanabe K, Kawamori T, Nakatsugi S, Ohta T, Ohuchida S, Yamamoto H *et al.* (2000). Inhibitory effect of a prostaglandin E receptor subtype EP<sub>1</sub> selective antagonist, ONO-8713, on development of azoxymethane-induced aberrant crypt foci in mice. *Cancer Lett* **156**: 57–61.
- Watts IS, Wharton KA, White BP, Lumley P (1991). Thromboxane (Tx) A<sub>2</sub> receptor blockade and TxA<sub>2</sub> synthase inhibition alone and in combination: comparison of anti-aggregatory efficacy in human platelets. *Br J Pharmacol* **102**: 487–505.

- Webb ML, Liu EC, Monshizadegan H, Hedberg A, Misra RN, Goldenberg H *et al.* (1993). Binding and function of a potent new thromboxane receptor antagonist, BMS 180,291, in human platelets. *J Pharmacol Exp Ther* **264**: 1387–1394.
- Westwick J, Webb H (1978). Selective antagonism of prostaglandin (PG) E<sub>1</sub>, PGD<sub>2</sub> and prostacyclin (PGI<sub>2</sub>) on human and rabbit platelets by di-4-phloretin phosphate (DPP). *Thromb Res* **12**: 973–978.
- Wheeldon A, Vardey CJ (1993). Characterization of the inhibitory prostanoid receptors on human neutrophils. *Br J Pharmacol* **108**: 1051–1054.
- Wilson NH, Jones RL (1985). Prostaglandin endoperoxide and thromboxane A<sub>2</sub> analogues. *Adv Prostaglandin Thromboxane Leukot Res* **14**: 393–425.
- Wilson RJ, Giblin GM, Roomans S, Rhodes SA, Cartwright KA, Shield VJ *et al.* (2006). GW627368X ((N-[2-[4-(4,9-dioxy-1-oxo-1,3-dihydro-2H-benzo[f]isoindol-2-yl)phenyl]acetyl] benzene sulpho-namide): a novel, potent and selective prostanoid EP<sub>4</sub> receptor antagonist. *Br J Pharmacol* **148**: 326–339.
- Wilson SJ, Roche AM, Kostetskaia E, Smyth EM (2004). Dimerization of the human receptors for prostacyclin and thromboxane facilitates thromboxane receptor-mediated cAMP generation. *J Biol Chem* **279**: 53036–53047.
- Wohlsen A, Martin C, Vollmer E, Branscheid D, Magnussen H, Becker W-M *et al.* (2003). The early allergic response in small airways of human precision-cut lung slices. *Eur Respir J* **21**: 1024–1032.
- Woodward DF, Hawley SB, Williams LS, Ralston TL, Protzman CE, Spada CS *et al.* (1990a). Studies on the ocular pharmacology of prostaglandin D<sub>2</sub>. *Invest Ophthalmol Vis Sci* **31**: 138–146.
- Woodward DF, Nieves AL, Williams LS (1990b). Thromboxane A<sub>2</sub>-mimetics are potent microvascular permeability factors in the conjunctiva. *J Pharmacol Exp Ther* **255**: 23–27.
- Woodward DF, Protzman CE, Krauss AH, Williams LS (1993a). Identification of 19(R)-OH prostaglandin E<sub>2</sub> as a selective prostanoid EP<sub>2</sub>-receptor agonist. *Prostaglandins* **46**: 371–383.
- Woodward DF, Spada CS, Hawley SB, Williams LS, Protzman CE, Nieves AL (1993b). Further studies on ocular responses to DP-receptor stimulation. *Eur J Pharmacol* **230**: 327–333.
- Woodward DF, Fairbairn CE, Krauss H-P, Lawrence RA, Protzman CE (1995a). Radioligand binding analysis of receptor subtypes in two FP receptors preparations that exhibit different functional rank orders of potency in response to prostaglandin. *J Pharmacol Exp Ther* **273**: 285–291.
- Woodward DF, Pepperl DJ, Burkley TH, Regan JW (1995b). 6-Isopropoxy-9-oxoxanthene-2-carboxylic acid (AH 6809), a human EP<sub>2</sub> receptor antagonist. *Biochem Pharmacol* **50**: 1731–1733.
- Woodward DF, Krauss AH-P, Chen J, Gil DW, Kedzie KM, Protzman CE *et al.* (2000). Replacement of the carboxylic acid group of prostaglandin F<sub>2α</sub> with a hydroxyl or methoxy substitute provides biologically unique compounds. *Br J Pharmacol* **130**: 1933–1943.
- Woodward DF, Krauss AH, Chen J, Liang Y, Li C, Protzman CE *et al.* (2003). Pharmacological characterization of a novel antiglaucoma agent, Bimatoprost (AGN 192024). *J Pharmacol Exp Ther* **305**: 772–785.
- Woodward DF, Phelps RL, Krauss AH, Weber A, Short B, Chen J *et al.* (2004). Bimatoprost: a novel antiglaucoma agent. *Cardiovasc Drug Rev* **22**: 103–120.
- Woodward DF, Krauss AH-P, Wang JW, Protzman CE, Nieves AL, Liang Y *et al.* (2007). Identification of an antagonist that selectively blocks the activity of prostamides (prostaglandin-ethanolamides) in the feline iris. *Br J Pharmacol* **150**: 342–352.
- Woodward DF, Liang Y, Krauss AH (2008). Prostamides (prostaglandin-ethanolamides) and their pharmacology. *Br J Pharmacol* **153**: 410–419.
- Xiang A, Uchida Y, Nomura A, Iijima H, Sakamoto T, Ishii Y *et al.* (2002). Involvement of thromboxane A<sub>2</sub> in airway mucous cells in asthma-related cough. *J Appl Physiol* **92**: 763–770.
- Xu W, Chou C.-L, Sun H, Fujino H, Chen QM, Regan JW (2008). FP prostanoid receptor-mediated induction of the expression of early growth response factor-1 by activation of a Ras/Raf/mitogen-activated protein kinase signaling cascade. *Mol Pharmacol* **73**: 111–118.
- Xue L, Gyles SL, Wetley FR, Gazi L, Townsend E, Hunter MG *et al.* (2005). Prostaglandin D<sub>2</sub> causes preferential induction of pro-inflammatory Th2 cytokine production through an action on chemoattractant receptor-like molecule expressed on Th2 cells. *J Immunol* **175**: 6531–6536.
- Yanagisawa A, Smith JA, Brezinski ME, Lefer AM (1987). Mechanism of antagonism of thromboxane receptors in vascular smooth muscle. *Eur J Pharmacol* **133**: 89–96.
- Yang J, Xia M, Goetzl EJ, An S (1994). Cloning and expression of the EP<sub>3</sub>-subtype of human receptors for prostaglandin E<sub>2</sub>. *Biochem Biophys Res Commun* **198**: 999–1006.
- Yang W, Ni J, Woodward DF, Tang-Liu DD, Ling KH (2005). Enzymatic formation of prostamide F<sub>2α</sub> from anandamide involves a newly identified intermediate metabolite, prostamide H<sub>2</sub>. *J Lipid Res* **46**: 2745–2751.
- Yang Z, Reiling S, Nieduzak TR, Mathew RM, Jackson S, Harris KJ (2008). 2-Phenylindoles as prostaglandin D<sub>2</sub> receptor antagonists. WO2008/014186, Sanofi-Aventis.
- Yasui K, Asanuma F, Hirano Y, Shichijo M, Deguchi M, Arimura A (2008). Contribution of prostaglandin D<sub>2</sub> via prostanoid DP receptor to nasal hyperresponsiveness in guinea pigs repeatedly exposed to antigen. *Eur J Pharmacol* **578**: 286–291.
- Yokoyama C, Yabuki T, Shimonishi M, Wada M, Hatae T, Ohkawa S *et al.* (2002). Prostacyclin-deficient mice develop ischaemic renal disorders, including nephrosclerosis and renal infarction. *Circulation* **106**: 2397–2403.
- Yokoyama U, Minamisawa S, Quan H, Ghatak S, Akaike T, Segi-Nishida E *et al.* (2006). Chronic activation of the prostaglandin receptor EP<sub>4</sub> promotes hyaluronan-mediated neointimal formation in the ductus arteriosus. *J Clin Invest* **116**: 3026–3034.
- Yoshida M, Sato Y, Shimura T, Ohkubo S, Honma S, Tanaka T *et al.* (2007). Distinct effects of Z-335, a new thromboxane A<sub>2</sub> receptor antagonist, on rabbit platelets and aortic smooth muscle. *Pharmacology* **79**: 50–56.
- Yoshikawa M, Takagi K, Ohinata K, Inui A, Asakawa A, Kakudo S (2005). Novel uses of prostaglandin D<sub>2</sub>, prostaglandin D<sub>2</sub> agonist and prostaglandin D<sub>2</sub> antagonist. US2005/0215609, Shionogi & Co.
- Yu M, Ives D, Ramesha CS (1997). Synthesis of prostaglandin E<sub>2</sub> ethanolamide from anandamide by cyclooxygenase-2. *J Biol Chem* **272**: 21181–21186.
- Zacharowski K, Olbrich A, Otto M, Hafner GS, Thiemermann C (1999). Effects of the prostanoid EP<sub>3</sub>-receptor agonists M&B 28767 and GR 63799X on infarct size caused regional myocardial ischemia in the anaesthetized rat. *Br J Pharmacol* **126**: 849–858.
- Zehender H, Witte EC, Wolff HP, Patscheke H (1988). Irreversible inhibition of the TXA<sub>2</sub>/PGH<sub>2</sub> receptor of human platelets by a photoaffinity ligand. *Biochem Pharmacol* **37**: 491–495.
- Zhang R, Ogletree ML, Moreland S (1996). Characterization of thromboxane A<sub>2</sub>/prostaglandin endoperoxide receptors in aorta. *Eur J Pharmacol* **317**: 91–96.
- Zhang Z, Hefferan MP, Loomis CW (2001). Topical bicuculline to the rat spinal cord induces highly localized allodynia that is mediated by spinal prostaglandins. *Pain* **92**: 351–361.
- Zhou N, Zeller W, Krohn M, Anderson H, Zhang J, Onua E *et al.* (2009a). 3,4-Disubstituted indole acylsulfonamides: a novel series of potent and selective human EP<sub>3</sub> receptor antagonists. *Bioorg Med Chem Lett* **19**: 123–126.
- Zhou N, Zeller W, Zhang J, Onua E, Kiselyov AS, Ramirez J *et al.* (2009b). 3-Acrylamide-4-aryloxyindoles: synthesis, biological evaluation and metabolic stability of potent and selective EP<sub>3</sub> receptor antagonists. *Bioorg Med Chem Lett* **19**: 1528–1531.